



Echinococcosis: Advances in the 21st Century

Hao Wen,^a Lucine Vuitton,^b Tuerhongjiang Tuxun,^c Jun Li,^{a,d} Dominique A. Vuitton,^b Wenbao Zhang,^{a,d} Donald P. McManus^e

^aState Key Laboratory of Pathogenesis, Prevention and Treatment of High Incidence Diseases in Central Asia and WHO Collaborating Centre for Prevention and Care Management of Echinococcosis, Urumqi, China

bWHO Collaborating Centre for Prevention and Treatment of Human Echinococcosis and French National Centre for Echinococcosis, University Bourgogne Franche-Comte and University Hospital, Besancon, France

Department of Liver and Laparoscopic Surgery, Digestive and Vascular Surgery Center, The First Affiliated Hospital of Xinjiang Medical University, Urumqi, China

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SUMMARY Echinococcosis is a zoonosis caused by cestodes of the genus *Echinococcus* (family Taeniidae). This serious and near-cosmopolitan disease continues to be a significant public health issue, with western China being the area of highest endemicity for both the cystic (CE) and alveolar (AE) forms of echinococcosis. Considerable advances have been made in the 21st century on the genetics, genomics, and molecular epidemiology of the causative parasites, on diagnostic tools, and on treatment techniques and control strategies, including the development and deployment of vaccines. In terms of surgery, new procedures have superseded traditional techniques, and total cystectomy in CE, *ex vivo* resection with autotransplantation in AE, and percutaneous and perendoscopic procedures in both diseases have im-

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Address correspondence to Donald P. McManus, Don.McManus@qimrberghofer.edu.au.

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^dClinical Medical Research Institute, The First Affiliated Hospital of Xinjiang Medical University, Urumqi, China

eMolecular Parasitology Laboratory, Infectious Diseases Division, QIMR Berghofer Medical Research Institute, Brisbane, Queensland, Australia

proved treatment efficacy and the quality of life of patients. In this review, we summarize recent progress on the biology, epidemiology, diagnosis, management, control, and prevention of CE and AE. Currently there is no alternative drug to albendazole to treat echinococcosis, and new compounds are required urgently. Recently acquired genomic and proteomic information can provide a platform for improving diagnosis and for finding new drug and vaccine targets, with direct impact in the future on the control of echinococcosis, which continues to be a global challenge.

KEYWORDS alveolar echinococcosis, cystic echinococcosis, echinococcosis, *Echinococcus, Echinococcus granulosus, Echinococcus multilocularis*, genetic epidemiology, genome, transcriptome, strains/genotypes, zoonosis

INTRODUCTION

chinococcosis refers principally to two severe zoonotic tapeworm diseases, cystic echinococcosis (CE) and alveolar echinococcosis (AE), caused by *Echinococcus* granulosus sensu lato and Echinococcus multilocularis, respectively (1). CE is cosmopolitan and more common, although a few island countries have declared elimination (2, 3). In areas of endemicity, the annual CE incidence ranges from <1 to 200 per 100,000, whereas that of AE ranges from 0.03 to 1.2 per 100,000 (4). Mortality in untreated or inadequately treated AE patients is >90% within 10 to 15 years of diagnosis (1). The CE mortality rate (2% to 4%) is lower but may increase considerably if inadequate care management is provided. Current estimates of the global burden average 285,500 disability-adjusted life years (DALYs) for human CE (5-7) (>1 million if underreporting is taken into account) and 666,434 DALYs for AE (7). The World Health Organization (WHO) has listed echinococcosis as one of the 17 neglected diseases targeted for control or elimination by 2050 (http://whqlibdoc.who.int/hq/2012/WHO_HTM_NTD _2012.1_eng.pdf). Indeed, major recent advances are set to revolutionize the care management and control of echinococcosis. Nevertheless, improved diagnosis and identification of new drug and vaccine targets are urgently required given the limitations of current diagnostic procedures, the toxicity and poor efficacy of available drugs, the often-inadequate surgical strategy, and the challenges in control and prevention.

In this review we outline the biology and life cycle characteristics of the *Echinococcus* spp. and consider the epidemiology, transmission, and clinical features of echinococcosis. We discuss recent advances in the diagnosis, treatment, care management, prevention, and control of CE and AE and show how genome and transcriptome studies are unravelling details of the developmental biology of *Echinococcus* spp. and their interactions with mammalian hosts, providing important information that can lead to the development of novel interventions and therapies against echinococcosis.

BIOLOGY AND LIFE CYCLE CHARACTERISTICS

The life cycles of the Echinococcus spp. are dependent on predator-prey associations involving two mammalian hosts (Fig. 1). Carnivores (canids and felids) serve as definitive hosts for the adult tapeworms, and their herbivorous prey (ungulates, rodents, and lagomorphs) act as intermediate hosts for the metacestodes; humans are generally not directly involved in the transmission of CE or AE, although under certain unique and unusual circumstances, such as reported in the Turkana region of Kenya, humans can act as intermediate hosts for E. granulosus (1). The developmental stages of the Echinococcus spp., exemplified by E. granulosus sensu lato, are shown in Fig. 2 (8, 9). Hundreds to thousands of 3- to 7-mm-long Echinococcus sp. adult worms develop in the intestines of their definitive hosts; the last segment (or proglottid) of each worm matures to produce eggs that are released in the carnivore's feces into the external environment. In turn, humans or the intermediate hosts ingest the eggs, which hatch in the intestine to release oncospheres that pass through the portal and lymphatic vessels and reach the liver, where they usually settle and develop as larvae (metacestodes or hydatid cysts); less frequently they may also reach the lungs, brain, bones, or any other organ of the human or intermediate host. Protoscoleces, the fertile forms of

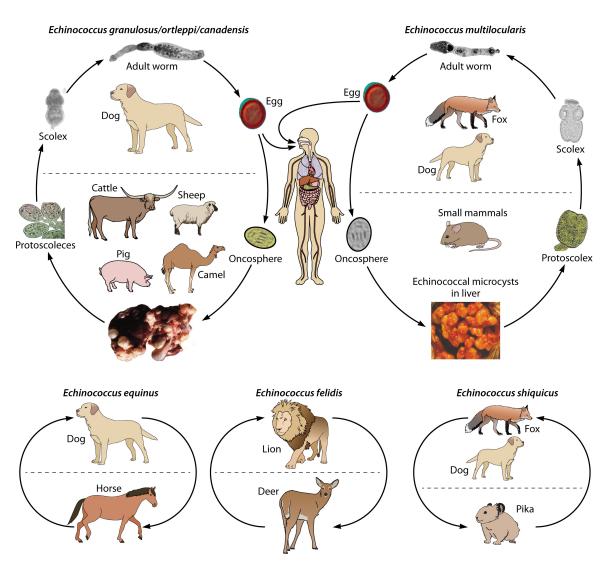


FIG 1 Life cycles of *Echinococcus* spp. Species responsible for human infection (*E. granulosus sensu stricto, E. ortleppi,* and *E. canadensis* [belonging to *E. granulosus sensu lato*] and *E. multilocularis*) are shown at the top. Species at the bottom (*E. shiquicus,* a species close to *E. multilocularis,* and *E. equinus* and *E. felidis,* belonging to *E. granulosus sensu lato*) are not known to cause disease in humans. Only the most common definitive and intermediate hosts which play a major role in life cycle/transmission are shown; other hosts may be encountered (especially wildlife hosts for *E. granulosus sensu lato* and domestic hosts for *E. multilocularis*). *E. vogeli* and *E. oligarthra,* which are responsible for polycystic echinococcosis in humans in Central and South America, are not represented in the figure.

the parasite, produced asexually by the metacestode, are released into the hydatid fluid; when ingested by the definitive host, protoscoleces evaginate their scoleces, aided by bile salts, and, after attaching to the intestinal wall, they develop into mature, egg-producing adult worms.

EPIDEMIOLOGY AND TRANSMISSION

Distribution of CE and AE

The pattern of distribution for CE has remained essentially unchanged over the past 2 decades, with areas of high endemicity, including western China, Central Asia, South America, Mediterranean countries and eastern Africa (Fig. 3), and the main risk factors being contact with dogs and raising livestock (3, 10, 11). However, studies in Africa have revealed a significant number of human cases and active transmission in animals, including wildlife, in countries hitherto considered not to be areas of endemicity (12, 13). Five thousand new CE cases are still diagnosed annually in Argentina, Brazil, Chile, Peru, and Uruguay (14, 15). Thirty years of dosing dogs with the anthelmintic drug praziquantel 8 times annually has significantly decreased transmission to humans, but

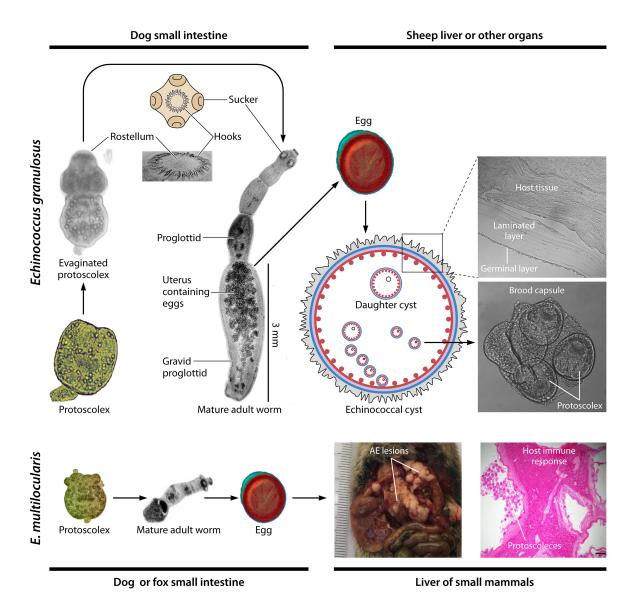


FIG 2 Different developmental stages in *Echinococcus granulosus* and *E. multilocularis*. Growth of the larval cyst is unlimited, and it can, for *E. granulosus*, grow to 30 cm or more in humans, while the adult worm, egg, and protoscolex are limited in size and shape. *Echinococcus* sp. tapeworms have no gut, circulatory, or respiratory organs and have a highly adapted relationship with their mammalian hosts which they exploit for nutrients, signaling pathways, and neuroendocrine hormones. Strobilization is a notable feature of cestode biology, whereby proglottids (segments) bud distally from the anterior scolex, resulting in the production of tandem reproductive units (proglottids) exhibiting increasing degrees of development. *Echinococcus* is monoecious, and the last segment (gravid proglottid) produces diploid eggs that give rise to ovoid embryos, the oncospheres. However, a striking feature of the biology of *Echinococcus* is that the protoscolex has the potential to develop in either of two directions: it may develop into an adult tapeworm producing sexually produced eggs in the dog gut, or, if a hydatid cyst ruptures within the intermediate or human host, each released protoscolex is capable of differentiating asexually into a new cyst, a process termed "secondary" echinococcosis. While a unilocular fluid-filled bladder (cyst) is a feature of *E. granulosus sensu lato* in its larval stage, the metacestode of *E. multilocularis* consists of a mass of small, multilocular vesicles embedded in the immune reaction of the host (granuloma and fibrosis). These multiple and aggregated vesicles grow by proliferation of cells in the germinal layer of the metacestode.

CE is still present in a number of areas in South America (14, 16). CE has been declared eliminated from New Zealand, and Tasmania in Australia is considered to be provisionally free of the disease (17); nevertheless, *E. granulosus* is present on the Australian mainland and is still found in Tasmanian wild and rural dogs, but at low prevalence (18). In Western Europe and North America, most human cases are imported, although an autochthonous cycle of various genotypes within the species group *E. granulosus sensu lato* (see below) is present. However, the lack of accurate case recording currently prevents any precise mapping of the true epidemiological picture; a European Registry of CE (the Heracles project) has been launched to improve this situation (19).

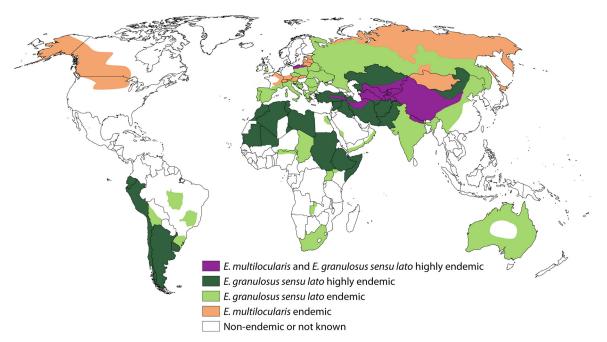


FIG 3 Global distribution of *Echinococcus granulosus sensu lato*, responsible for cystic echinococcosis (CE), and *Echinococcus multilocularis*, responsible for alveolar echinococcosis (AE). The map is based on recent epidemiological studies (1, 13, 19, 247) as far as the current situation has been studied in a given area. The different colors represent a proxy for human prevalence and infection in animal hosts in a given area (to take autochthonous human cases into account only). For AE, the represented disease density is based mainly on the presence of autochthonous AE cases in humans, *E. multilocularis* metacestodes in small mammals, and *E. multilocularis* adult worms in foxes and dogs. For CE, the represented disease density is based mainly on the presence of autochthonous human cases of CE and of *E. granulosus sensu lato* metacestodes (irrespective of species or genotype) in intermediate hosts, including sheep, cattle, equids, and camels. For more accurate and detailed data and maps, see a recent comprehensive review paper by Deplazes et al. (13).

AE has been a public health concern in northern Japan for the past 40 years (20–22). Mass screening with ultrasound (US) and serology in China have confirmed a high incidence of AE on the Tibetan plateau (in Qinghai, Sichuan, and Tibet [23]) and show that AE prevalence, especially in children, is higher than that of CE in several areas (24). Among the 18,235 estimated new AE cases per year globally, 91% occur in China (7), with human prevalence of >3% in some areas (25). AE is also endemic in Central Asia, with high endemicity of both *E. multilocularis* and *E. granulosus* in Kazakhstan and Kyrgyzstan (26–29). In Europe, the prevalence of *E. multilocularis* in definitive and intermediate hosts increased markedly within the first 15 years of this century, and the geographic distribution of fox infections is far broader than earlier reported; urban foxes may be involved in transmission (13, 17, 30–33). Human cases have been found in European countries previously considered to be free of AE, and the situation in the Baltic region has become worrisome; in addition, AE incidence has doubled in the previously recognized areas of endemicity of France, Switzerland, Germany, and Austria (13, 34–36).

In regard to North America, the north-central United States, northwestern Alaska, and northwestern Canada have long been areas of *E. multilocularis* endemicity, but the parasite's geographic range appears to be expanding due, at least in part, to increased and improved sampling efforts and the targeting of definitive hosts other than foxes (such as coyotes [*Canis latrans*]) (13). AE had not been considered a mainstream human health issue in North America other than in Alaska until recently, and *E. multilocularis* has not been reported from Mexico or the southern United States (13). However, human cases were reported in Alberta, Canada, in the past decade (37) as well as in Quebec and Manitoba (unpublished reports to a WHO Collaborating Centre); molecular analysis of *E. multilocularis* in Alberta suggests that coyotes are important definitive hosts and that a European strain is involved, perhaps through carnivores imported from Europe, and not the local endemic "Alaskan" strains (38, 39).

TABLE 1 Current recognized species within the genus Echinococcus and their preferential hosts and geographic distribution

Species	Definitive host(s)	Intermediate host(s)	Human cases	Distribution
Echinococcus granulosus sensu stricto	Domestic dog, wolf, dingo, jackal, other canids	Sheep, goat, cattle, pig, camel, buffalo, horse, wild ungulates, marsupials, etc.	Yes	Cosmopolitan
Echinococcus canadensis	Domestic dog, wolf	Pig, camel, cervids	Yes	Eurasia, Africa, North and South America
Echinococcus ortleppi	Domestic dog	Cattle	Yes	Eurasia, Africa
Echinococcus felidis	Lion	Hyena, warthog, zebra, wildebeest, bush pig, buffalo, various antelopes, giraffe, hippopotamus	Not reported	Africa
Echinococcus equinus	Domestic dog	Horse, other equids, cervids	Not reported	Eurasia, Africa
Echinococcus multilocularis	All fox species, wolf, raccoon dog, domestic dog, cat	Arvicoline and microtine rodents and small herbivorous mammals, including lagomorphs (e.g., pika); pigs, boars, horses, cattle, nutrias, nonhuman primates, and dogs are accidental hosts	Yes	Eurasia, North America
Echinococcus oligarthra	Wild felids (e.g., <i>Puma concolor</i> [puma])	Dasyprocta azarae (agouti), Didelphis marsupialis (opossum)	Yes	Central and South America
Echinococcus vogeli	Bush dog, domestic dog	Cuniculus paca Linnaeus, 1766 (paca)	Yes	Central and South America
Echinococcus shiquicus	Tibetan fox	Ochotona curzoniae (Tibetan plateau pika)	Not reported	Tibetan Plateau

The distribution of "neotropical echinococcosis," i.e., echinococcosis due to Echinococcus vogeli and Echinococcus oligarthra (see comment below on its correct taxonomic spelling), remains limited to South America (40); newly recognized human cases of E. vogeli infection in new areas, such as French Guyana in eastern South America (41, 42), are likely the result of improved diagnosis and molecular identification of the disease (43).

Genetics and Genetic Epidemiology

A major change in the epidemiological picture for CE has come about as a result of the redefinition of the Echinococcus spp. causing the disease. Until relatively recently, E. granulosus was considered a single species, but it is now recognized as having extensive genetic diversity, with distinct strains/genotypes exhibiting differences in pathology and differing responses to drugs and the defined recombinant vaccine EG95 (see "Vaccination of intermediate hosts" below) for ovine CE (3). The application of mitochondrial DNA sequencing has resulted in the recognition of 10 genotypes (G1 to G10) and their accurate identification in molecular epidemiological surveys of CE in different geographical settings and host assemblages (44). Accordingly, the 10 strains/ genotypes of E. granulosus sensu lato have been demarcated into 5 species, including E. granulosus sensu stricto (the former "sheep strain," G1 to G3), Echinococcus equinus (horse strain, G4), Echinococcus ortleppi (cattle strain, G5), Echinococcus canadensis (camel strain, G6; pig strain, G7; G9, probably a variant of the pig strain; and cervid strains, G8 and G10), and Echinococcus felidis ("lion strain") (12, 45-48). Currently, 9 species, listed in Table 1, are recognized in the genus Echinococcus (49); the life cycles of some of these are shown in Fig. 1. E. granulosus sensu stricto is the most widely distributed (Fig. 3), with other species being focal.

The use of mitochondrial DNA and/or DNA microsatellites, such as the EmsB marker (50), has made discrimination between distinct genotypes of E. multilocularis possible. The influence of these genetic differences on increased prevalence or severity of AE in humans is unknown, but such genetic analysis is useful for tracking the transmission of a particular genotype from one area to another (51-54). The Alaskan origin of E. multilocularis found in the Norwegian Svalbard islands and the European origin of E. multilocularis found in Alberta, Canada, are examples where molecular markers have proved useful (37, 55). Echinococcus shiquicus, which is transmitted between Tibetan foxes/dogs (56) and the plateau pika (Ochotona curzoniae), is a new species found only in the Tibetan region, with no human cases thus far recorded (56-59). The less common E. vogeli and E. oligarthra (replacing the previously used incorrect taxonomic spelling "E.

oligarthrus" ["arthra" being the plural of the Greek noun "arthron," which means "joints" {i.e., proglottids}, and not an adjective subject to gender agreement with "Echinococcus" {48}]) are restricted to Central and South America (42, 60, 61). Analysis of nuclear and mitochondrial markers revealed that populations of E. vogeli (Brazilian Amazon) and E. oligarthra (Argentina) are genotypically variant (60, 62).

CLINICAL FEATURES

The clinical features of echinococcosis have been comprehensively described (1, 63). In CE these are associated with damage or dysfunction of target organs, particularly the liver (70%) and lungs (20%), with the remainder including the brain, spleen, kidney, and heart. Almost all primary AE lesions are in the liver. Clinically, most AE and CE patients present late at clinics or hospitals. Population screening has shown that CE liver cysts in humans grow very slowly, with more than half of cysts showing no change in size in 10 years and one-third growing less than 3 cm; mean cyst growth in cases with a prolonged follow-up was 0.7 cm (8). The early stages of CE and AE do not cause symptoms, and CE cysts and AE lesions can remain asymptomatic for 10 to 15 years; consequently, children comprise only a small percentage of echinococcosis patients. Clinical symptoms usually appear when a cyst reaches more than 10 cm in diameter in the liver or when more than 70% of the organ volume is occupied by a cyst or cysts, resulting in physical compression or damage to bile ducts, hepatic veins, the portal vein, or the hepatic artery. Various symptoms may be due to compression or damage to bronchia in the lungs or various structures of the brain, which can also result in life-threatening complications. In any organ, compression of vital structures may be symptomatic even with small or medium-sized cysts.

Symptomatic CE patients with liver cysts most often present with upper abdominal discomfort and poor appetite; compression of bile ducts may lead to jaundice. On palpation, a tumor-like mass, hepatomegaly, or abdominal distension may be found. Chest pain, cough, or hemoptysis can be indicative of cysts in the lung, and cyst rupture into the bronchi may result in the expulsion of hydatid materials. Any neurological symptom (signs of intracranial hypertension, epilepsy, all types of paralysis, etc.) may be observed in patients with brain cysts. In any organ, cyst rupture can induce fever, urticaria, eosinophilia, and anaphylactic shock. Potentially lethal allergic reactions due to cyst rupture and even minor fissures have long contraindicated any puncture of a CE cyst. Antigen leakage associated with such fissures may reveal the usual development of specific IgE antibodies, common for E. granulosus sensu lato and E. multilocularis, which is part of the predominant Th2-type immune response in echinococcosis, with high levels of interleukin-5 (IL-5) in both AE and CE (64). Despite the presence of IgE antibodies and demonstrated possible basophil activation (65), eosinophilia and allergic reactions are very uncommon in AE because of the different structure of the parasitic lesions, with dense fibrosis preventing vesicle fluid leakage; these reactions may be observed in rare cases of blood dissemination of lesion fragments (64). Routine imaging or population mass ultrasound (US) screening of the liver may identify asymptomatic cysts (66), and the procedure is extremely important for finding earlystage AE patients in areas of endemicity (23).

Faster growth of cysts in CE patients with AIDS suggests that immune suppression may play a role in CE progression (67, 68). Conversely, the concept of an enhancing effect of CE on the occurrence of cancer in the population, because of a defect in immune surveillance linked to an *Echinococcus*-induced tolerance state, has been raised recently (69, 70), although this hypothesis has not been rigorously evaluated. In areas where CE is endemic, the simultaneous occurrence of two frequent diseases cannot be ruled out, and preliminary assessment from hospital medical information systems of the occurrence of cancer in 2,350 patients with CE compared with patients without CE showed no difference (Bo Ran, First Affiliated Hospital of Xinjiang Medical University, personal communication). However, the promoting effect of cancer or its treatment on parasite growth seems to interfere far less with the occurrence and/or disease course in CE than in AE (71).

Genetic variation of the human leukocyte antigen (HLA) system is associated with the occurrence and/or progression of AE lesions in humans (72, 73); patients with the HLA-DR3 DQ2 haplotype were shown to have more severe disease and a more pronounced Th2-type immune response, associated with a deeper tolerance status (74, 75). The effects of immune suppression on *E. multilocularis* growth are well known in animal models (76). They were first reported in human cases after liver transplantation performed to treat AE and in patients with AIDS. Early recurrence of AE was observed in transplanted patients and was found associated with the level of immune suppression resulting from the treatment used to prevent liver rejection (77); unusually rapid progression of lesions and the corrective effect of antiretroviral therapy were observed in AIDS patients, including children (67, 78).

After AE occurrence was reported in transplant patients, other than those undergoing liver transplantation for AE, and in patients treated for malignant or chronic inflammatory diseases (79-81), a systematic study based on the French National Registry of AE (FrancEchino) confirmed the significant and recent increase (from the beginning of the 21st century) in the occurrence of AE in such patients (71). Acquired therapeutic immunosuppression (which combined chemotherapy, corticosteroids, and biotherapy such as anti-tumor necrosis factor [anti-TNF] agents) appears to be the main factor for AE occurrence and its fast progression. Unusual presenting symptoms, such as bacterial abscess-like acute clinical symptoms, and misleading imaging findings, such as abscess-like, metastasis-like, or hemangioma-like aspects on computed tomography (CT) scans, contributed to delayed diagnosis. This resulted in incorrect therapeutic management of a number of these patients, such as those undergoing cancer treatment intensification, which further enhanced metacestode growth, or ineffectual radiofrequency ablation attempts on the presumed liver "tumor" or "metastasis" (71). Negative serology, likely due to the patient's immune suppression status, is also an issue and makes pathological and/or molecular identification of the metacestode often necessary before diagnosis can be confirmed (71). Whether the patients were actually infected with E. multilocularis eggs during the period of therapeutic immune suppression or the symptoms were due to a reactivated, dormant metacestode resulting from a previous infection remains to be established (71).

Diagnosis

General comments. Imaging techniques are essential for diagnosis, with the relatively inexpensive and portable ultrasound (US) widely used to diagnose CE or AE liver lesions; X-ray is used for lung cysts. Both techniques are used for diagnosis and population screening and for follow-up (82–84). Serology, i.e., detection of specific antibodies against *Echinococcus* sp. antigens, is a confirmatory step with various levels of sensitivity/specificity correlating with the involved species, lesion location, or antigen used (63) (Fig. 4). Mass population screening of CE and AE in areas of endemicity using US is considered the best method for early diagnosis. In addition to organized mass screening, routine health checks and systematic follow-up of associated diseases, including US examination, are now a major approach in echinococcosis diagnosis. This has contributed to the diagnosis of asymptomatic cases in the general population and to changes in the presentation of AE cases, especially in Europe (85). Systematic follow-up of patients with malignant diseases and a variety of chronic diseases, by using US, CT scan, and fluorodeoxyglucose-positron emission tomography (FDG-PET), may also have contributed to the early diagnosis of AE in such individuals (71).

Imaging and classification of CE and AE lesions. Based on US imaging, the World Health Organization Informal Working Group on Echinococcosis (WHO-IWGE) has classified hepatic CE cysts into five types, CE1 to CE5, and AE lesions into different PNM (parasite lesion, neighbor organs, metastases) types (1, 86–88), which provides basic information for clinicians to make treatment decisions (Fig. 5; Table 2). Harmonization of disease assessment at the international level is crucial to progress toward an evidence-based and stage-specific strategy for treatment (89). New classification of US images and the associated classification of computed tomography (CT) images in AE

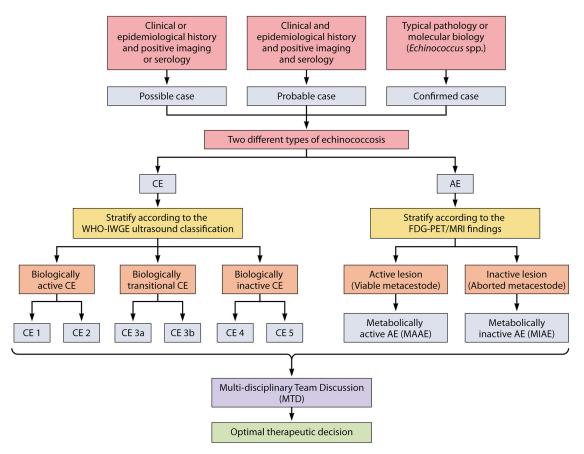


FIG 4 Algorithm for the diagnosis of cystic echinococcosis (CE) and alveolar echinococcosis (AE). Definitions of "possible," "probable," and "confirmed" cases refer to the "Expert consensus for diagnosis and treatment of echinococcosis in humans" (86). CE1 to CE5 refer to the "WHO-IWGE [World Health Organization Informal Working Group on Echinococcosis] international classification of ultrasound images in cystic echinococcosis for application in clinical and field epidemiological settings" (86) and Fig. 5. FDG-PET, fluorodeoxyglucose-positron emission tomography (increased uptake of FDG by the periparasitic immune response is the currently accepted evidence for AE lesion metabolic activity) (94). MRI, magnetic resonance imaging (identification of typical microcysts on T2-weighted images at MRI is a surrogate marker for AE lesion metabolic activity) (98).

have been proposed (90, 91) and are currently being tested on patients at European and Chinese centers in order to evaluate their usefulness for diagnosis and follow-up.

The challenge in imaging diagnosis of echinococcosis is detecting small cysts/ lesions (<2 cm in diameter). Contrast-enhanced ultrasonography (CEUS) may be used for detecting small AE lesions and differentiating them from abscesses and tumors based on pulsating blood flow imaging (92–94). Fluorodeoxyglucose (FDG) uptake surrounding AE lesions is higher than in other areas, and FDG-positron emission tomography (FDG-PET) has become the favored reference tool to evaluate their metabolic activity (Fig. 4 and 6) (95–98). Color and pulsed doppler US, dual-energy CT or spectral CT, and diffusion-weighted magnetic resonance imaging (MRI) might also be useful in detecting blood supply and the metabolism of lesions (97, 99) but they cannot be recommended without further evaluation (97). In CE, MRI appears to be of better diagnostic value than CT scanning (100), and both procedures are complementary for AE and should be performed to provide sufficient information for therapeutic decision-making (Fig. 4) (97). However, MRI T2-weighted microcystic images are pathognomonic of AE lesions (Fig. 6), and in difficult cases US-guided core-needle biopsy is reliable and effective in combination with DNA diagnostic testing (101) or specific immunostaining (102).

Serology. The sensitivities and specificities of serological tests for CE and AE have been comprehensively reviewed (103). Hydatid fluid (HF) is the major antigenic source for echinococcosis immunodiagnosis, with the HF lipoproteins antigen B (AgB) and

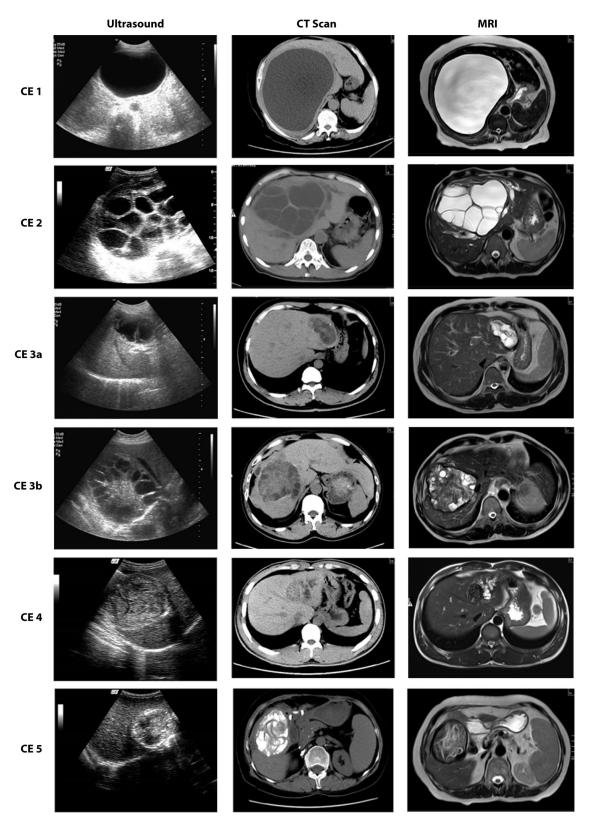


FIG 5 Imaging of cystic echinococcosis. The description of ultrasound images is according to the WHO Informal Working Group on Echinococcosis (WHO-IWGE) international classification (86) and corresponding images were obtained from plain computed tomography (CT) scanning and T2-weighted magnetic resonance imaging (MRI) in representative cases. In the international classification, types CE1 and CE2 correspond to "active stages," types CE3a and b to the "transitional stage," and types CE4 and CE5 to "degenerating stages" (CT and MRI images provided by Liu Wenya, Department of Radiology, First Affiliated Hospital of Xinjiang Medical University, Urumqi, People's Republic of China).

TABLE 2 WHO Informal Working Group on Echinococcosis PNM classification and staging of alveolar echinococcosis^a

Classification	Description
P	Hepatic localization of the parasite
PX	Primary AE lesion cannot be assessed
P0	No detectable AE lesion in the liver
P1	Peripheral lesion(s) without proximal vascular and/or biliary involvement
P2	Central AE lesion(s) with proximal vascular and/or biliary involvement of one lobe ^b
P3	Central lesion(s) with hilum vascular or biliary involvement of both lobes and/or with involvement of two hepatic veins
P4	Any liver lesion with extension along the vessels ^c and the biliary tree
N	Extrahepatic involvement of neighboring organs (diaphragm, lung, pleura, pericardium, heart, gastric and duodenal wall, adrenal glands, peritoneum, retroperitoneum, parietal wall [muscles, skin, bone], pancreas, regional lymph nodes, liver ligaments, kidney)
NX	Not evaluable
N0	No regional involvement ^d
N1	Regional involvement of contiguous organs or tissues
М	Absence or presence of distant metastases (lung, distant lymph nodes, spleen, central nervous system, orbital, bone, skin muscle, kidney, distant peritoneum, and retroperitoneum)
MX	Not completely evaluated
M0	No metastasis ^e
M1	Metastasis
PNM stages	
1	P1 N0 M0
II	P2 N0 M0
Illa	P3 N0 M0
IIIb	P1–P3 N1 M0, P4 N0 M0
IV	P4 N1 M0, any P any N and/or M1

^aAccording to reference 88.

antigen 5 widely used in serological assays for CE (1). Although the Casoni intradermal test exhibits low specificity and sensitivity (104), it may be used for confirming the results of US or other physical imaging methods (Xinyu Peng, personal communication). Poor standardization and ethical issues regarding reagents from animal origin injected into humans have, however, considerably limited the use of skin tests for echinococcosis diagnosis. Reported sensitivities and specificities of serological methods for testing CE patients confirmed by surgical resection vary from 60% to 90%. Use of enriched antigen 5 (105) and recombinant antigens based on repeated tandem E. granulosus AgB (2B2t antigen) and recombinant Ag5 (106) increases diagnostic value. A large number of other novel antigens, including the tegumental protein EgTeg (107), alkaline phosphatase (EgAP) (108), and EpC1 (109), exhibited greater than 90% sensitivity and specificity on selected serum samples. However, their performance has never been evaluated on a large scale, and none of the reported antigens are sufficiently sensitive or specific to be used as a first-intent tool for diagnosis or mass population screening (110, 111). One major issue is the lack of appropriate antigens with the required sensitivity for the serological detection of small CE cysts in the liver and cysts of any size in lungs. Encystment of the metacestode, preventing the stimulation of antibody-producing cells and thus causing the absence of measurable levels of antibodies generated against Echinococcus sp. antigens, can explain many negative results.

AE serology is more reliable. Em2 and Em492, which represent constituents of the excretory/secretory (ES) fraction of intact metacestodes, as well as EmAP and EmP2, are specific for *E. multilocularis* infection (76). EM10, or its derivatives EmII/3 and Em18, which are encoded by part of the EM10 gene sequence, show high diagnostic performance for confirming AE (112). A commercialized Em2^{plus} enzyme-linked immunosorbent assay (ELISA) (Bordier, Crissier, Switzerland) has been used extensively for clinical diagnosis of AE (113), with sensitivity and specificity exceeding 90% (114). Nevertheless,

^bFor classification, the plane projecting between the bed of the gall bladder and the inferior vena cava divides the liver in two lobes.

^cVessels mean inferior vena cava, portal vein, and arteries

dIncluding a negative chest X ray or thoracic CT result.

^eIncluding a negative chest X ray or thoracic CT result as well as a negative brain CT result.

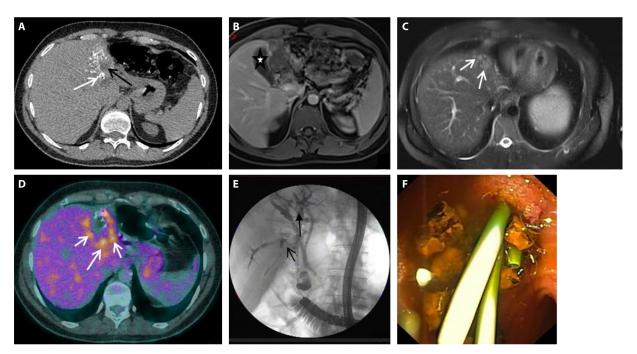


FIG 6 Complementarity of imaging techniques for the diagnosis and preoperative assessment of alveolar echinococcosis (AE) lesions in a patient with portal vein and bile duct invasion by the parasitic lesions. (A) On computed tomography (CT) scanning, the lesion shows the characteristic heterogeneous aspect of AE, with hyperdense calcifications (white arrow) and hypodense area (central necrosis) (black arrow). (B) Fat-suppressed T1-weighted magnetic resonance (MR) image after gadolinium injection at the portal venous phase shows an atrophy of the left liver and invasion of the left portal vein and of the left intrahepatic biliary tree. The lesion is at the contact of the gallbladder (white star) which is not invaded. (C) T2-weighted MR images show the presence of hyperintense microcysts (white arrows), pathognomonic of AE, but also a solid component (Kodama type II). (D) Fluorodeoxyglucose (FDG) uptake in positron emission tomography (PET) is markedly increased at the periphery of the lesion (white arrows). (E) Assessment of biliary tree involvement and treatment by perendoscopic stenting in a patient with late postoperative biliary complications of AE. Endoscopic retrograde cholangiopancreatography (ERCP) was performed (with a colonoscope because of previous gastrectomy), showing dilation of the extrahepatic and intrahepatic biliary tree with several defects (black arrows) because of biliary stones due to chronic biliary obstruction and bacterial superinfection. (F) Perendoscopic stenting via ERCP. After balloon dilation followed by extensive lavage with isotonic saline and stone extraction, 3 plastic stents (size, 7 and 10 French) are inserted in the stenosis of the bile duct (endoscopic view).

serology cannot be used as a first-intent diagnostic tool for mass screening in areas of endemicity, where a proportion of the human population exhibits positive serology without AE lesions (115). Serology has also been shown to be frequently negative in immunosuppressed individuals with AE, and thus, it should not be used on its own as an argument against a diagnosis in such patients (71).

For both CE and AE, serology is thus now used only to confirm imaging results; it may also provide some insight into the infection pressure on a given population (e.g., children) in a particular geographic area. Serology results are included in the definition of "possible" and "probable" cases by the Expert Consensus of the WHO-IWGE (86). The uses of imaging, serology, and molecular identification of the metacestode in the diagnostic strategy are shown in the algorithm proposed in Fig. 4.

Protein biomarkers. *Echinococcus* spp. can survive in humans for a long time through active regulation of the host immune response by the secretion of proteins at the interface of parasite and host tissues. Profiling HF protein composition and excretory/secretory (ES) products provides valuable information on parasite survival strategies and the molecular mechanisms of parasite-host interaction. In addition, analysis of the protein profiles can help in identifying potential molecular markers for developing diagnostic and follow-up tools. Proteomic analysis of the composition of CE (116, 117) and AE (118) cyst/vesicular fluids has identified hundreds of proteins from both *Echinococcus* spp. and the host that may help differentiate subpopulations of patients. Characterization of ES proteins from *E. granulosus* adult worms (119) and protoscoleces (120, 121) and *E. multilocularis* protoscoleces (122) also shows promise for identification of potential diagnostic markers.

Application of proteomics to patient care management is in its infancy. Specific

immunodominant epitopes of *E. granulosus* HF change as the disease progresses (e.g., from CE1 to CE2) (123), and the HF protein composition is different in different organ locations of the cysts (124); this could explain the well-known differences in the host antibody response correlating with the stage and location of cysts. Immunoreactive proteins from *E. multilocularis* vesicular fluid have been recently identified and quantified, and comparative proteomics revealed 9 proteins (actin modulator protein, fucosidase alpha L1 tissue, prosaposin a preprotein, glutathione S-transferase, beta-galactosidase, NiemannPick C2 protein, elongation factor 2, cathepsin b, and H17g protein tegumental antigen) that were more abundant in immunoprecipitation eluates from albendazole (ABZ)-nonresponders than in those from ABZ-responder AE patients, suggesting that detection of antibodies against these proteins by ELISA could be helpful in monitoring the course of AE under ABZ treatment (118).

DNA detection. Recently developed DNA-based methods, such as quantitative and/or nested PCR assays, are highly sensitive, reasonably specific, and able to distinguish Echinococcus species from each other and from other cestodes; they can, as discussed above, discriminate the various genotypes of E. granulosus, including following clinical biopsy of a suspected CE or AE case, and identify infected mammalian host species (125-130). Other examples include the identification of E. vogeli in patients infected in an area not previously considered to be an area of endemicity (42), in revealing the reemergence of E. ortleppi in France (131), in the retrospective identification of an E. multilocularis strain in a historical human case of AE in the United States (132), and in the detection of E. multilocularis infection in primates (133). DNA identification methods are now routinely used on biopsy or fine-needle cytology specimens for the diagnosis of AE in patients with unusual imaging aspects and/or with negative serology, typified by immunosuppressed patients (71), and they form part of the definition of "confirmed cases" of the WHO-IWGE Expert Consensus (86) (Fig. 4). As is discussed further below, molecular diagnosis, including loop-mediated isothermal amplification (LAMP), can be used as a first-line screen for Echinococcus spp. in the field (134-137) and to detect Echinococcus sp. egg DNA in environmental samples as an important step for identifying high-risk contaminated areas and for defining the actual routes of human infection (138-142).

CARE MANAGEMENT

Based on image classification and following a stage-specific approach, various options are possible, alone or combined, for the treatment of both CE and AE, including (i) surgery, (ii) nonsurgical interventions, (iii) anti-infective benzimidazole drug treatment, and (iv) a "watch-and-wait" approach (Fig. 7). The current recommendations for echinococcosis management take the cancer-like nature of the disease, which is prone to recurrence, into account and thus use the model of cancer care management which promotes a multidisciplinary approach, with interdisciplinary team consultations for therapeutic decisions, the combination of surgical and drug treatments, a long-term follow-up of patients, the establishment of international recommendations, and the creation of reference centers (86).

Treatment of CE

With CE, treatment centers on cyst type according to the WHO-IWGE US classification (1) (Fig. 5), size, location, and presence/absence of complications, as well as available medical expertise and equipment (86). Curative treatment is achieved by the complete removal of the cyst, regardless of location. If the cyst with all its layers (including adventitia) cannot be removed totally, which is the case with sub-total cystectomy and all types of partial cystectomy and with the percutaneous "PAIR" (puncture, aspiration, injection, and reaspiration) technique, the therapeutic procedure should be complemented with the use of protoscolecidal agents. Intraoperative dissemination of protoscolex-rich fluid during surgery and insufficient killing of protoscoleces and germinal membrane during the percutaneous procedures are major causes of

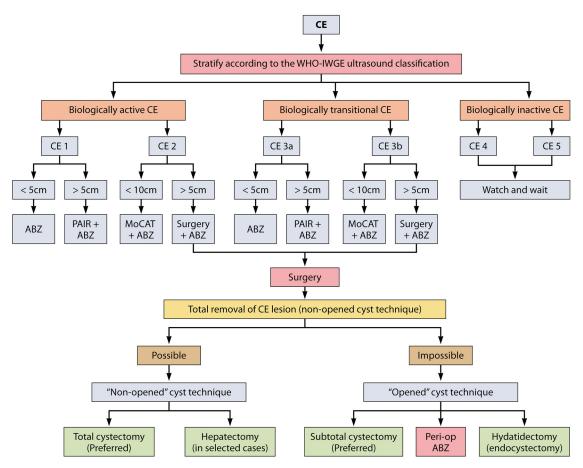


FIG 7 Algorithm for the treatment of cystic echinococcosis (CE), based on "WHO-IWGE international classification of ultrasound images in cystic echinococcosis for application in clinical and field epidemiological settings" (86) and Fig. 4 and 5. WHO-IWGE, World Health Organization Informal Working Group on Echinococcosis; ABZ, albendazole; PAIR, puncture-aspiration-injection-reaspiration (nonsurgical percutaneous interventional technique for treatment of CE cysts).

CE recurrence (86). An algorithm that describes the recommended therapeutic strategy for CE is given in Fig. 7.

Use of protoscolecides during CE surgery. The intraoperative dissemination of protoscolex-rich hydatid fluid during surgery or the PAIR procedure for CE is a major cause of cyst recurrence (86). Injection of a protoscolecide into CE cysts to reduce the risk of spillage of viable protoscoleces and possible recurrence is an integral part of the surgical technique employed by many surgeons around the world (143). A very broad spectrum of protoscolecidal agents, from warm water (144) to the highly toxic formalin (145), have been tested over the past 50 years, but the feasibility, safety, and efficacy of many of these compounds have generally not been determined. Details of the concentrations and modes of administration of currently available and tested protoscolecides are provided in Table 3. Serious complications have limited the use of some of these; formalin and betadine should never be used under any circumstances. However, the majority, albeit less harmful than these two compounds, resulted in serious biliary, gas embolism, renal, and toxic complications that limited their use (146) (Table 3); toxicity to bile duct mucosa explains why communication between the CE cyst and the bile ducts must be carefully checked before the use of any protoscolecidal agent. There have been considerable efforts made to discover additional potential protoscolecides, including plant extracts and the use of physical methods in vitro, but details of their clinical application are lacking. Currently, the WHO-IWGE recommends 20% hypertonic saline as the preferred protoscolecide in surgery and 20% hypertonic saline or 95% alcohol in PAIR (86), but rigorous, high-quality comparative studies on these protoscolecidal agents are still awaited.

TABLE 3 Protoscolecides tested for use in CE surgery or PAIR $^{\!\scriptscriptstyle \alpha}$

		Use:					
Category	Tested agents (references)	In surgery	In PAIR	In vitro	Limitations	Recommendation	Cautions
Chemical compounds	Albendazole (147), alcohol (148), betadine (149), cetrimide-chlorhexidine solution (150), chendeosycholic acid (151), cyclosporin A (152), formalin (145), FBG (153), hydrogen peroxide(154), octenidine dihydrochloride (155), pyridinyl imidazole derivative (156), silver nitrate (05%) (157), synthesized silver nanoparticles (158), praziquantel (159), taurolidine (160), thymol (161)	Alcohol, betadine, cetrimide- chlorhexidine solution, formaldehyde solution, hydrogen peroxide, silver nitrate (0.5%)	Albendazole, alcohol, betadine, hyperronic saline, sliver nitrate	Chenodeoxycholic acid, cydosporin A, FBG, pyridinyl imidazole derivative, synthesized silver nanoparticles, praziquantel, taurolidine, thymol	The efficacy of these agents is concentration dependent and thus demonstrated toxic effects on bile duct mucosa, leading to sclerosing cholangitis	For surgery (20% hypertonic saline), careful monitoring of patients' electrolytes and appropriate protection (with isotonic 0.9% saline-moistened laparotomy towels) to prevent hyperosmolar damage to the peritoneum are recommended; for PAIR, (hypertonic saline and >95% alcohol, communication of the cyst with bile ducts must be checked (and repaired, if any found, at surgery) before the use of any protoscolecide	Formaldehyde solution and betadine must not be used; gas embolism has been reported with the use of hydrogen peroxide, and this protosolecide should not be used in deep highly vascularized cavities
Natural extracts	Allium sativum extract (162), Berberis vulgaris aqueous extract (163), Foeniculum vulgare mill extract (164), Myrus communis oli extract(165), Nigella sativa oil (166), Pistacia vera oil extract (167), P. khinjuk methanolic extract (168), P. atlantica hydroalcoholic extracts (169), Peganum seed extracts (170), Salvadora persica root extracts (171), Salvadora persica methanolic extract (171), Zalvadora multiflora methanolic extract (1712).	No reports	No reports	These agents showed strong protoscolecidal effects (100% killing) at different concentrations and incubation times	Lack of reports of clinical application in surgery and PAIR	Further experimental testing and clinical trials strongly advocated to assess their practical application	
Other agents	Warm water (144), honey (173), propolis (174)	No reports	No reports	These agents showed strong protoscolecidal effects (100% killing) at different concentrations and incubation times	Lack of reports of clinical application in surgery and PAIR	Further experimental testing and clinical trials strongly advocated to assess their practical application	
Physical protoscolecidals	nsPEF (175), RFA (176)	RFA tested in human Not assessed and animal surgery	Not assessed	nsPEF exhibited a good protoscolecidal effect	Low availability and high cost	Further assessment strongly recommended	

Abbreviations: PAIR, puncture, aspiration, injection, and reaspiration of the cyst content; FBG, fluoride-containing bioactive glass; nsPEF, nanosecond pulsed electric field; RFA, radiofrequency thermal ablation.

Anti-infective treatment. Systemic anti-infective treatment relies on continuous administration of 2 benzimidazole carbamates, ABZ and mebendazole, which are the only anti-infective drugs clinically efficient to interrupt larval growth of Echinococcus spp. (177, 178). Mebendazole was the first benzimidazole that was proven efficient for the treatment of echinococcosis (179, 180). Because of its increased bioavailability and easier administration to patients, ABZ was then preferred as the anti-infective treatment of choice for echinococcosis, at an average dosage of 15 mg/kg/day (181). Currently, mebendazole is only an alternative drug for those patients who have experienced severe hepatic adverse effects with ABZ. Most of these patients experience similar adverse effects with both drugs; however, some individuals may tolerate mebendazole, which is critical when the patients cannot be operated on and their survival totally depends on the anti-infective treatment, a situation more frequent in AE than in CE (182). In CE, anti-infective treatment alone is reserved for small or medium-sized isolated cysts or, alternatively, multiple and inoperable cysts in the liver and/or in multiple organs. A combination of interventional techniques with ABZ is recommended and is used routinely with PAIR and derived techniques; it is less widely used with surgery (86, 89, 183, 184). However, criteria for curtailing anti-infective treatment are clearly missing and deserve prospective studies to be undertaken, and treatment length and schedule are still a matter of debate; a prospective study on a limited number of patients showed that 3 months of AE treatment was no better than 1 month of administration after PAIR (63). Based on pharmacological evidence and the relatively low and slow efficacy of ABZ to kill protoscoleces, a reasonable compromise would be to administer ABZ from 1 week before to 2 months after the interventional procedure (surgery or PAIR) whenever the cyst has been opened; however, firm recommendations should await the results of real studies, especially for the association with surgery. The "watch-and-wait" strategy is recommended for asymptomatic and small CE1 cysts, obviously degenerating CE4 cysts, and all CE5 type cysts (89).

A more systematic use of total cystectomy (also known as periadventitial cystectomy or, incorrectly "pericystectomy"), modified by the Chinese surgeon Peng Xinyu (Fig. 8) (89), has increased over the past 15 years (185, 186), and the attitude of surgical teams regarding total cystectomy is currently changing. To prevent recurrence, total cystectomy, which avoids cyst opening, is the technique of choice (Fig. 7). When the cyst is adjacent to major vessels, sub-total cystectomy, which avoids dissection of these vessels, is encouraged. If both techniques are not feasible, hydatidectomy (also called "endocystectomy" or "partial cystectomy"), after cyst opening, may be used together with obsessional prevention of protoscolex spillage, the major cause of recurrence, during surgery and with perioperative ABZ administration.

Since the first report of perlaparoscopic treatment of a CE patient in 1992 (187), robotically assisted (188) and single-incision laparoscopic total cystectomy and hepatic resection (189) have put laparoscopy into CE surgical practice (190). When laparoscopic surgery is performed, there must be no compromise to the principle of avoiding cyst content spillage and respecting cyst wall integrity; however, the influence of the laparoscopic approach on recurrence is controversial (191). For those CE patients with obvious biliary communication and unsuited for total cystectomy, "double drainage" of the fistula and cystic duct (i.e., drainage of the main bile duct with a Kehr T-tube through the cystic bile duct and drainage of the fistula or of the remaining cavity after partial cystectomy) is now preferred to reduce postoperative biliary leakage. In case of postoperative biliary leakage, a perendoscopic drainage (after endoscopic retrograde cholangiopancreatography [ERCP]) should be considered before any reoperation (192, 193). Liver transplantation for CE could be the last option in selected cases (194).

PAIR has definitely become part of the interventional therapeutic options in CE for midsized CE1 and CE3a cysts (63) (Fig. 7). A recent improvement in PAIR is the "modified catheterization technique" (MoCAT), a procedure appropriate for cysts up to 10 cm in diameter that includes aspiration of the parasitic membranes in addition to the cyst content and with a catheter left in place for the postintervention period of time. (195). When used by experienced operators, this technique may be an alternative to surgery

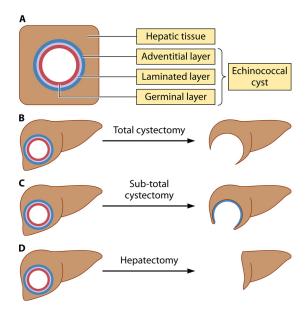


FIG 8 Schematic structure of the echinococcal cyst and different approaches for surgical removal. (A) The echinococcal cyst is made up of the adventitial layer, laminated layer, and germinal layer (from outside to inside). (B) Total cystectomy involves resection of the entire adventitial layer ("subadventitial resection"), the laminated layer, and the germinal layer. (C) Sub-total cystectomy involves partial resection of the adventitial layer and total resection of the laminated layer and the germinal layer, leaving parts of the adventitial layer in place whenever the operation is difficult because of the proximity of large vessels and/or adhesions. (D) Hepatectomy involves the *en bloc* resection of the echinococcal cyst along with part of the normal liver parenchyma. Partial cystectomy, which requires opening of the cyst, may leave all or part of the laminated layer and germinal layer and relies on the efficacy of a protoscolecide to destroy the metacestode; it should generally not be considered because of the potential for recurrence.

for noncomplicated CE2 and CE3b cysts (Fig. 5 and 7). Recent reviews have confirmed the efficacy and safety of PAIR and its variants if a stage-related strategy and technical recommendations are strictly followed (86, 89, 185).

Treatment of AE

With AE, therapeutic decision is based on the possibility of complete resection of liver lesions, after multidisciplinary assessment involving liver imaging, the general status of the patient, and the technical capabilities of the surgical team (86, 178, 182). As AE lesions are most often located in the right liver lobe and in advanced cases have invaded the major bile ducts and vessels (portal veins, hepatic veins, and vena cava), major hepatic surgery is often required, with significant morbidity and mortality resulting because of uncontrolled bleeding or liver failure. A number of cases cannot be safely operated on, even by highly experienced hepatic surgeons; only left hepatectomy in the less-frequent cases, where lesions are located in the left lobe, is accessible to less-experienced, nonspecialized surgeons. This explains why only one-third of patients with AE may benefit from curative liver resection, and the number is even lower in communities where AE is endemic and patients live in remote places and are late in seeking care (63). Palliative operations have been shown to be a source of complications without improving patient survival; it is the reason why they are now not recommended (63, 86). Objectives for the treatment of AE thus include the following: (i) totally removing the parasitic lesion, which is achieved through "curative" surgery combined with 2 years of ABZ treatment at the same dosage and with the same precautions as for CE treatment; (ii) if this is not possible, reducing the proliferating potential of the E. multilocularis metacestode by continuous administration of ABZ; and (iii) alleviating complications, especially bile duct obstruction and cholangitis and bacterial infection of the necrotic cavity that develops in the centers of advanced lesions (Fig. 6). Lesions which are massively calcified and/or negative by FDG-PET may

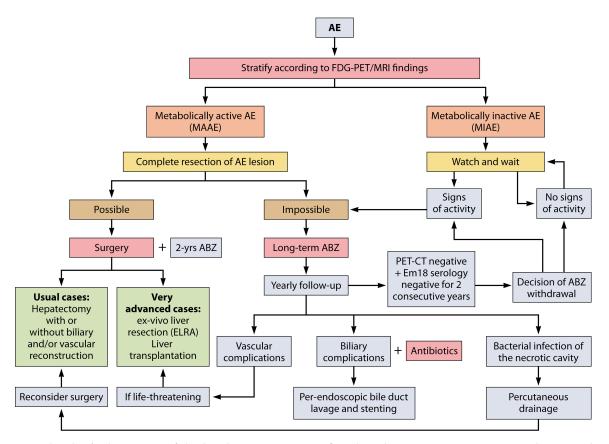


FIG 9 Algorithm for the treatment of alveolar echinococcosis. FDG-PET, fluorodeoxyglucose-positron emission tomography (increased uptake of FDG by the periparasitic immune response is the currently accepted evidence for AE lesion metabolic activity) (94). MRI, magnetic resonance imaging (identification of typical microcysts on T2-weighted images at MRI is a surrogate marker for AE lesion metabolic activity) (98). ABZ, albendazole; ELRA, *ex vivo* liver resection with autotransplantation.

benefit from a "watch-and-wait" approach. An algorithm that describes the recommended therapeutic strategy for AE is given in Fig. 9.

In immunocompromised patients, reducing immunosuppressive treatment must be considered when this is possible. In the French cohort, ABZ efficacy was shown to be fast and excellent but with more adverse effects than in nonimmunocompromised patients treated in the same centers. Whenever possible, and depending on the prognosis of the associated disease, curative liver resection should be performed as early as is realistic because of the fast growth of the metacestode in this situation and in order to facilitate the care management of the associated disease. Liver allotransplantation (77) is still used in advanced cases, especially when hepatic veins and the vena cava are included in the parasitic lesions, in case life-threatening complications result, but the shortage of donors and life-long immunosuppressant administration, which is followed by higher susceptibility to disease recurrence, have discouraged application of this approach (182). The high rate of postoperative morbidity and mortality (30% within the first 6 months after transplantation), as well as the recurrence rate (10% locally and 20% for distant metastases), in a recent report from Turkey even raises an ethical question, especially when the livers are from living donors (196). Ex vivo liver resection followed by autotransplantation is a surgical procedure for excising lesions following removal of the liver from the patient; the remaining lesion-free liver is then reinserted, similar to a liver transplantation (197) (Fig. 10). The procedure was initially developed to treat conventionally "unresectable" tumors as it does not require an organ donor and postoperative immunosuppressive treatment (198, 199); it was first applied to patients with advanced AE in 2011 (197, 200, 201). AE patients often present with hypertrophy of the liver lobe not invaded by the parasitic lesion (because of

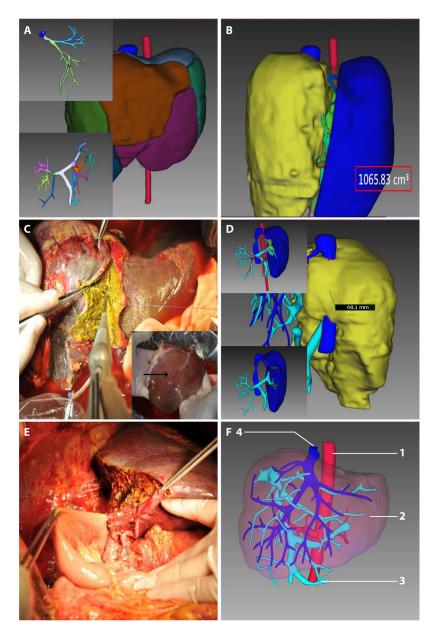


FIG 10 Three-dimensional reconstruction in the application of *ex vivo* liver resection and autotransplantation (ELRA). (A) Portal veins, hepatic veins, and segments of liver are visualized by three-dimensional reconstruction. (B) Calculation of remnant hepatic parenchymal volume after three-dimensional reconstruction and virtual resection. The volume of remnant liver is 1,065.83 cm³ (yellow, giant AE lesion; blue, normal parenchyma). (C) Precise resection of giant AE lesion on bench (black arrow, normal liver parenchyma after resection). (D) The length (68.1 mm) of obliterated retro-hepatic vena cava is calculated through three-dimensional reconstruction (yellow, giant AE lesion; blue, retro-hepatic vena cava). (E) Hepaticojejunostomy is performed on the table (black arrow, anastomosis). (F) Postoperative follow-up demonstrating liver remnant and vasculatures (1, aorta and hepatic artery; 2, liver; 3, portal vein; 4, hepatic vein).

chronic long-term portal vein obstruction and/or a specific influence of the immune response to the parasite which favors hepatic regeneration). This is one of the reasons for a relatively favorable outcome of the procedure (and of major hepatectomies in general) compared with cancers which develop rapidly and do not promote hypertrophy of the remaining liver lobe (202) (Fig. 10). Midterm results of such operations seem acceptable in comparison to conventional major hepatectomy with complex bile duct and vessel reconstruction or to liver transplantation (203, 204). With an average follow-up of 22.5 months (range, 14 to 89 months) in 69 patients, overall mortality was

12%, complications higher than IIIa (according to the Clavien classification) were observed in 10 patients, and there was no recurrence (204). However, long-term results and comparison with nonsurgical care management, including long-term ABZ and perendoscopic treatment of biliary complications, are not yet available.

European data indicate a marked trend toward a reduction in the percentage of surgical operations for AE, whatever their type (complete or only partial resection of the lesions, or any surgical procedure such as bile duct derivation, or simple diagnostic laparotomy) but an increase in the percentage of radical/curative operations, i.e., liver resections capable of totally removing the metacestode tissue from the liver and/or other organs (178, 205, 206). However, the percentage of patients undergoing surgery has remained high in China (207). The differences are mainly due to recruitment of patients (either symptomatic with advanced lesions, most often in China, or asymptomatic with less-developed lesions, most often in Europe) and also, whatever the continent, to the specialization, experience, and boldness of the surgical teams and appreciation of the "safety margin" necessary to perform an R0 resection (according to the grading followed in cancer surgery). Evaluation of the influence of this safety margin suggests that at least a 1-mm distance is important (thus, being less than in cancer) combined with ABZ therapy (208). Difficulties in the strict follow-up of patients, because of their residence in remote areas, in measurement of ABZ sulfoxide (ABZ-SO) to monitor adherence to treatment (which is crucial for the success of an anti-infective drug approach) and the origin of adverse effects, and differences in the organization of health care management, may also be among the reasons that make Chinese teams more prone to favor a surgical approach.

Percutaneous puncture for treating AE patients with a necrotic cavity inside liver lesions and bacterial superinfection has been used for more than 30 years, and, in combination with antibiotics, the procedure may save patients and allows a reassessment of the resectability of the lesion (83, 178, 192). Injection of protoscolecidal agents should never be used in AE: in fact, the central cavity often observed in advanced cases of AE is due to the necrosis of the lesions, including the multiple degenerating microcysts of the metacestode and associated immune infiltrate and fibrosis; the still active microcysts are at the periphery of this cavity, as is well shown by FDG-PET images and T2-weighted MRI (98). These microcysts are embedded in the immune and fibrotic reaction and are not accessible to a protoscolecidal agent; in addition, at this stage, communication of this central cavity with bile ducts is the rule, and such an injection would be not only useless but also harmful (178). Early and/or late biliary complications clearly heavily impact the immediate prognosis of the disease, at presentation and within the first year of follow-up, and thus the final outcome of AE (85, 209); they represent a negative turning point in the course of the disease (210). Percutaneous dilation of the bile ducts obstructed by the progression of the metacestode was widely performed until the end of the 20th century, instead of palliative surgical bile diversion (83, 209, 210).

A European survey of perendoscopic procedures (through ERCP) to treat biliary complications of AE in 18 clinical centers showed that such procedures are now used routinely and are generally successful in alleviating symptoms and in maintaining long-term permeability of the biliary strictures; to achieve good results, extensive saline lavage of the bile ducts, which removes necrotic debris and intrahepatic biliary stones that are common in such patients, and the use of multiple plastic stents are recommended (211) (Fig. 6). Perendoscopic bile duct stenting is currently nearly totally replacing surgical palliative operations and percutaneous biliary drainage to treat biliary complications in AE patients (Fig. 6). Although there have been no specific studies assessing the quality of life of patients receiving such treatment, we may anticipate that this has markedly contributed to the improved quality of life of those patients with chronic biliary obstruction and multiple cholangitis episodes; in the past, more than 10 reoperations in a single patient were not uncommon, and most patients had a very uncomfortable external biliary drainage tube for life (211).

In Europe, all retrospective evaluations of survival after AE diagnosis have indicated that there have been major improvements in the 21st century compared with the prior 30-year experience; improvement was already noted in the 1990s compared with the 1970s, being mostly due to earlier diagnosis, the introduction of anti-infective treatment, and the progressive abandonment of palliative surgery (85, 212). The situation remains worrisome, however, in countries/regions with limited medical facilities and where diagnosis is performed only at an advanced stage of disease, as well as for those patients who experience severe side effects of benzimidazoles, as there is no alternative. Until now, the numerous attempts at finding new drugs or at converting drugs already used in other parasitic diseases for their application to the echinococcoses have essentially failed (213). Drugs that showed some promise in vitro and in experimental animals, and were effectively tested in humans or domestic animals, include other benzimidazole compounds such as flubendazole and oxfendazole, as well as nitazoxanide (182). Lists of those drugs that were tested in vitro and found not suitable for use in humans after being tested in vivo in experimental animals are available in morespecialized reviews (182, 213). There is some hope from in vivo experiments with mefloquine (214-218) and artemisinin (219, 220) derivatives, and very recently, derivatives of carbazole aminoalcohols have been shown effective against cysts of E. granulosus both in vitro and in vivo (221). However, none of these compounds have, as yet, been subjected to pilot clinical trials.

CE and AE Disease Follow-Up

Long-term imaging follow-up after initiation of treatment (more than 10 years) has long been stressed for AE (86, 222). It is now accepted that a close follow-up of patients with CE should also be undertaken for at least 5 years because of the high rates of relapse after surgery and the uncertainty of complete cure after drug treatment and/or percutaneous puncture. Regular tests of blood counts and serum transaminases are necessary to assess the safety of management within the first 6 months after initiation of anti-infective treatment, since hepatic toxicity and leukopenia are the most severe adverse effects and may prevent ABZ use in some patients. ABZ-SO or mebendazole measurements are extremely useful to assess the patient's observance and to adjust drug dosages. ELISAs using HF antigens and/or purified AgB and Ag5 for CE and EM2-plus or Em18 ELISA, if available, for AE exhibit high performance in detecting disease recurrence after surgical resection of the cyst/lesion, although they are less accurate if all or part of the cyst/lesion remains in the infected organ (182, 223). FDG-PET is currently considered the "gold standard" for the evaluation of the metabolic activity of AE lesions and for decisions about anti-infective treatment interruption; however, its predictive value is far from perfect, despite technical improvements (e.g., delayed image acquisition 3 h after FDG injection [224, 225]), and all other imaging techniques deserve more evaluation (97).

There has been recent active searching for parasite viability/disease progression markers in the sera of AE patients (226), with antibodies against recEm18 showing promise for long-term monitoring (227-229); however, when used alone, the test is not sufficiently discriminant to make a decision on treatment withdrawal. Double-negative results for FDG-PET and anti-recEm18 antibodies currently represent the best marker to consider treatment interruption (228). In all patients, recommended follow-up is at 1, 4, and 12 weeks for the first 3 months after diagnosis and initiation of ABZ, to check for ABZ adverse effects by measuring blood cell count and transaminases, and whenever possible, adjusting the treatment based on ABZ-SO measurement. The patient then is asked to present every 3 months for the first year and every 6 months until the end of the second year; this follow-up includes US, blood count, transaminases, and serology, and ABZ-SO is also measured 2 to 4 weeks after each dose adjustment, if necessary (182). In all patients FDG-PET is performed at the end of the second year. In patients with curative surgery, ABZ treatment is withdrawn 2 years after surgery if there is no recurrence as assessed by US, FDG-PET, and serology; yearly follow-up using US and serology is then recommended until 10 years after surgery. In patients without curative

surgery, after the second year, yearly follow-up includes US, blood count, transaminases, and gamma-glutamyl transferase measurement, and serology, with FDG-PET-CT being performed every 2 years. A decision on anti-infective treatment withdrawal is made after at least 2 consecutive negative FDG-PET and Em18 serology assessments.

Detection of circulating serum or plasma *Echinococcus* sp. antigens (CAg) may be an alternative approach to serology. Early studies showed that *E. granulosus sensu lato*-specific circulating antigens, positive in 75% sera of antibody-negative CE patients, were associated with the growth dynamics and activity of cysts (230). Sensitivity of CAg detection, however, varied between 21% and 85%, mostly owing to the formation of circulating immune complexes. To our knowledge, only preliminary studies have been performed (231) to determine whether antigen detection may be a useful approach for assessing the efficacy of treatment, especially after removal of the cyst, and no studies using circulating antigens for long-term disease follow-up on a significant number of patients with CE or AE are available (36).

Assessing circulating cell-free DNA (cfDNA) could also be an option, as its detection as a biomarker has proved useful in cancer, and it has shown some promise in parasite diagnosis; for example, infections with all the three major human schistosomes (*Schistosoma mansoni, Schistosoma haematobium*, and *Schistosoma japonicum*) have been identified with PCR-based cfDNA assays using both species- and genus-specific target genes in animal models and patients (232). However, with the currently available detection methods, the approach does not seem sensitive enough for use in clinical practice, at least for AE, due to the low levels of cfDNA detectable in patient serum (233).

PREVENTION AND CONTROL

Current prevention and control of CE relies on the provision of safe animal slaughtering conditions (offal destruction and preventing dogs from feeding on infected organs of ungulates) and on dosing dogs with praziquantel (234). With notable exceptions, such as New Zealand, Tasmania, Iceland, Cyprus (at least temporarily), Chile, and some provinces in Argentina (3), other attempts at control have been generally disappointing. In countries where very strict slaughtering measures have been implemented, leading to the near disappearance of human CE cases, such as mainland Australia, persistence of a wild cycle of E. granulosus sensu stricto makes a reappearance of the disease always possible (39). Evaluation of control programs shows that (i) success is more readily achieved on islands, (ii) a combined multidisciplinary and multi-institution, and often multicountry, effort is necessary, and (iii) a One-Health approach is required (235, 236). An ambitious, well-financed, and state-driven control program, including community US screening, care management of diagnosed patients, and monthly dog dosing with praziguantel, is in operation across western China. The monthly dosing of dogs is suitable for village-based communities (237) but is far less effective in seminomadic or pastoral areas (238).

Vaccination of sheep with the EG95 vaccine has been promoted as a complementary intervention to eliminate CE transmission (14), and there have been trials of this approach in China and South America. For most countries where CE is endemic, however, the logistics and costs of vaccinating sufficient numbers of animals may preclude widespread application of the vaccine. Dog vaccination would be an effective complementary intervention for controlling echinococcosis transmission (239), although recent progress has been slow (240). Nevertheless, development of a single vaccine, effective against both *E. granulosus* and *E. multilocularis* in canines, may be feasible and would be practical given that the two species are sympatric in many countries of the Northern Hemisphere. Besides the red fox, the main definitive host of *E. multilocularis* (241–243), other carnivores such as the raccoon dog and domestic dog also act as definitive hosts (32, 244, 245). The role of dogs in AE transmission is especially important in western China and in central Asia (53, 246) and may be more relevant in Europe than previously considered; conversely, a wildlife cycle may also be of concern for CE, especially in Africa (13, 247). Baits impregnated with praziquantel

have been applied against *E. multilocularis* (248, 249), and a bait-delivered vaccine, when available, could be used to interrupt the parasite's transmission cycle in wildlife, notably foxes, in cities and parks (250) and in selected rural areas (31, 251).

A successful control campaign should focus on the most at-risk areas and on those animal hosts mainly involved in transmission, with progress constantly monitored (10, 17, 252, 253). Close surveillance of the prevalence of *Echinococcus* spp. in dogs/foxes is extremely important for evaluating the progress of a control program (254). Detecting and quantifying *Echinococcus* sp. eggs/proglottids in canine fecal samples is recommended as an alternative to necropsy. Major improvement in parasite DNA detection in feces and environmental samples has occurred in the last 15 years, and this is currently preferred to antigen-based diagnosis (126, 140, 255–259). As indicated above, LAMP-based assays are useful as a first-line screen for *Echinococcus* spp. in the field (134–137), and PCR methods allow combined identification of definitive host species and *Echinococcus* sp. infection status using feces collected in the field (260). Adaptation of the available molecular methods to detect *Echinococcus* sp. egg DNA in environmental samples (e.g., in soil, water, and sewage and on vegetables) is an important step to better identify high-risk areas and the actual routes of human infection (138–140), leading to more effective control.

RECENT APPLICATIONS OF OMICS TECHNOLOGIES

Improving Understanding of the Complexity of *Echinococcus* Species Life Cycles and Unravelling Species-Specific Phenotypic Differences

Gene transcript analysis of representative CE life stages (protoscoleces, cyst germinal cells and membranes, adult worms, and oncospheres) has allowed exploration of different aspects of tapeworm biology and parasitism (261). Further, the recent publication of the complete genomes of *E. granulosus* (261) and *E. multilocularis* (262) has revealed other key features associated with parasitism, including a description of a number of domain families gained during the course of evolution. Other important genes identified include those associated with strobilization and reproduction, signaling pathways, and neuroendocrine and nervous systems and others involved in evasion of immune recognition and regulation of host immunological responses. The genome and transcriptome data therefore provide a critical basis for more-detailed understanding of cestode biology, differentiation, development, evolution, and pathogenesis and other host-parasite interactions.

The complex life cycles of E. granulosus sensu lato and E. multilocularis provide a platform for addressing the functions of the expressed products of novel genes. Up- or downregulation of gene expression likely underpins the phenotypic changes associated with the different life cycle stages and the respective modulation of the immune response that each species determines. In-depth transcription analysis is critical for searching for key genes associated with these changes and for identification of their specific functions. One of the unique characteristic physiological features of E. granulosus sensu lato and E. multilocularis is the remarkable ability of the protoscolex to differentiate into an adult worm or to dedifferentiate into a cystic stage. Specific host stimuli (bile acids) govern the direction of development (263-265), and relevant parasite-expressed receptors and transporters likely stimulate the relevant developmental pathways. Gene function prediction analysis was unable to attribute a function to 3,900 of the 11,325 genes predicted to be present in E. granulosus sensu stricto; among these, 361 genes were transcribed in adult worms, of which 21 were highly expressed and may be associated with adult worm development (261). The morphology of the adult worms of Echinococcus spp., though following the typical taeniid cestode pattern, has only up to 5 immature, mature (with reproductive organs), and gravid proglottids sequentially present, which are replicated through strobilization. The gravid proglottid contains eggs which are released into the environment to infect intermediate and human hosts. Of the 361 genes shown by mRNA transcript analysis to be highly expressed in adult E. granulosus, 55 were specifically expressed compared with the oncosphere and cyst stages, in which these genes are silenced. The metaces-

tode stage is associated with unlimited asexual development, whereas adult worms have limited, sexual development; of 8,361 genes expressed in the two stages, 498 and 502 genes were highly expressed in the adult worm and in the metacestode, respectively (261). Future work aimed at posttranscriptional suppression of these genes through RNA interference (RNAi) and gene knockout techniques may unravel their functional characteristics.

A comprehensive comparison of the genomes and transcriptomes of E. granulosus and E. multilocularis will also be central to our understanding of the biological/ pathological differences between the two species. A major difference between the two is the morphology of the metacestode. E. granulosus has a unique cyst formation, with a shell-like adventitia which clearly separates the cyst from the surrounding (liver, lung, and brain) parenchyma. Conversely, the E. multilocularis metacestode is an infiltrating lesion composed of aggregated microvesicles, cells of the intermediate host's immune response, fibrosis and necrosis, with no clear edge to the lesion, which continuously progresses eccentrically and damages the liver or other target organs. Comparative analysis of divergent and convergent gene pairs and their pattern of expression using microarray technology or RNA sequencing (RNA-Seq) is a relatively recent approach that can be used to identify patterns that are shared by more than one species or are unique to a particular species, with the capacity to reveal differences in biological phenotype. Although in its infancy for studying Echinococcus spp., such gene pair analysis has revealed that E. granulosus and E. multilocularis have 10,018 genes with high sequence similarity, with 5,418 being identical. The next stage will be to identify and characterize nonsimilar/unique genes so as to shed light on the inherent differences in morphology or pathology between the two tapeworms.

Improving Diagnosis and Drug Treatment of Echinococcosis

The currently available rich genomic and transcriptomic data may be useful for developing new public health interventions against echinococcosis; these include improved diagnostic tests and the identification of new drug targets. BLAST sequence analysis of the E. granulosus sensu stricto genome indicated that one-third (n = 3,903)of the genes present have no gene homologues or orthologues in other taxa, suggesting that these genes are probably Echinococcus specific, likely underpinning the unique features and biological characteristics of E. granulosus sensu lato. The products of these genes may also be of value as new candidates for diagnosis and as novel drug targets for the treatment and control of echinococcosis. Some proteins likely serve as messengers for cross talk between E. granulosus and its hosts and may prove useful as chemotherapeutic targets as well as for improved immunodiagnosis or immunotherapy (261, 262). Potentially "druggable" proteins (i.e., polypeptides which might be the targets of new or existing drugs) expressed by genes in the germinal layer of the metacestode include G-protein-coupled receptors (GPCRs), serine proteases, ion channels, and neuropeptides (266) and components of the mitogen-activated protein kinase (MAPK) pathway (267-270).

Hormone- and cytokine-activated pathways have been identified in both *E. granulosus sensu stricto* and *E. multilocularis* metacestodes (271–273), and their activation/inactivation by host components is highly suggested (267, 269, 270, 273–281). Importantly, comparison of the *E. granulosus* and *E. multilocularis* genomes indicates a high level of gene sequence similarity, suggesting that the two parasites may share many common molecules that can be targeted for developing new interventions. MAPK inhibitors are currently being actively studied for their killing effects on the metacestode and/or protoscoleces. An ATP-competitive pyridinyl imidazole inhibitor (ML3403), targeting the P38-like MAPK from *E. granulosus sensu stricto*, effectively suppressed Egp38 activity, which led to significant protoscolex death within 5 days *in vitro* (267). Similar results were obtained with *E. multilocularis*; ML3403, in particular, and SB202190, another pyridinyl imidazole, tested on metacestode vesicles cultured *in vitro* led to dephosphorylation of the parasite's EmMPK2 and subsequent killing of the parasite vesicles at concentrations that did not affect cultivated mammalian cells (270).

As a direct result of the unveiling of the complete genomes of the Echinococcus spp., several metabolic pathways have been explored, and other inhibitors are currently being studied (177, 282-287). Nilotinib, an ABL (Abelson murine leukemia viral oncogene homologue)-tyrosine kinase inhibitor, and everolimus, a serine/threonine kinase inhibitor, caused alterations of E. multilocularis metacestode vesicles in vitro; however, neither of these compounds resulted in any reduction of parasite growth in E. multilocularis-infected mice, and combined application of the kinase inhibitors with ABZ did not lead to synergistic or additive treatment efficacy (282). BI2536, a Polo-like kinase (a kinase containing Polo box domains) inhibitor that has been tested in clinical trials against cancer, was shown to inhibit EmPlk1 activity and to block the formation of metacestode vesicles from cultivated E. multilocularis germinal cells; furthermore, it eliminated the germinal cell population from mature metacestode vesicles in vitro, yielding parasite tissue that was no longer capable of proliferation (286). Similarly, imatinib, another ABL tyrosine kinase inhibitor used to treat cancer, has been shown to interact with the ABL-like kinases present in E. multilocularis and to be highly effective in killing Echinococcus stem cells, metacestode vesicles, and protoscoleces in vitro (287). However, the potential of these kinase inhibitors to treat AE in vivo is as yet unknown.

Improving Understanding of Immunological Mechanisms of Host-Parasite Interactions To Develop Immunotherapy

Although the extreme susceptibility of *Echinococcus* spp., and especially of *E. multilocularis*, to the cellular immune response of the host has been well recognized since the 1980s (288), much of the comprehensive knowledge of the immunological mechanisms at work in the subtle balance between host protection and parasite growth has been gained in the 21st century (76, 289). In this field, advances in genomics have been of help by suggesting additional molecular mechanisms/pathways and new therapeutic targets. In particular, studies of the transcriptional profiles observed in the livers of *E. multilocularis*-infected mice and the use of mouse models with specific gene deletions have been crucial (290–293). Recently obtained information suggests strongly that immunotherapy could complement the anti-infective drug approach to treat echinococcosis. Conversely, a better understanding of the immunological profiles of intermediate hosts infected with *Echinococcus* spp. may add new tools to the therapeutic arsenal targeting chronic inflammatory diseases.

The predominance of a T helper 2 (Th2) profile, including interleukin-5 (IL-5)- and IgE-dependent reactions, and high levels of IL-10 cytokine at the chronic stage of both AE and CE in humans have been known for some time (64). In CE, a dominant Th2/T regulatory (Treg) cytokine profile is rapidly established after the formation of the adventitial fibrous barrier (293, 294). In AE, the immune response follows a 3-stage course characterized by a mixed Th1/Th2 profile at the early stage, a dominant Th2/Treg profile, including IL-10 and transforming growth factor β $(TGF-\beta)$ regulatory cytokines, at the chronic middle stage, and a T-cell exhaustion status at the final stage of infection (289, 295). Clinical studies on CE have shown the response to anti-infective therapy (or of "inactive" cyst) is associated with a Th1 profile, whereas on the other hand, resistance to treatment (or of "active" cyst) is associated with a Th2 profile and elevated IL-10 levels (289, 293). Clinical studies in AE suggest that a combination of Th2-related cytokine serum levels, such as those of IL-23 and IL-5, could be used as a surrogate marker of AE metabolic activity in humans (294). Some proteins likely serve as messengers for cross talk between E. granulosus and its hosts and may also prove useful as targets for improved immunodiagnosis or patient follow-up (261, 262).

The composition and type of the periparasitic immune response elicited by *Echinococcus* sp. infection causatively influence the outcome and progression of disease, ranging from resistance (self-cure) to rapidly evolving host fatality (high susceptibility) (291). In *E. multilocularis*, the parasite load, which can be quantitatively assessed in an experimental model involving infection by intraportal injection of protoscoleces, significantly influences this periparasitic response as well as the systemic cell and cytokine

profile (295). Recent experimental studies show that Th1/Th17 polarization is a pivotal factor for resistance, while FoxP3+ Tregs are key players in the immune regulatory processes favoring *E. multilocularis* metacestode survival (296). *In vivo* treatment of mice by a single intravenous injection of 200 μ l recombinant IL-17A at the optimal concentration of 125 pg/ml 2 weeks after *E. granulosus sensu stricto* infection decreased the infectivity rate by 2/3 and reduced metacestode growth by more than 90% (297). FoxP3 Treg depletion after infection using the DEREG (depletion of regulatory T cell) model in mice (296) and also genetic inhibition of the synthesis of fibrinogen-like protein 2 (FGL-2), a CD4+ CD25+ Treg effector molecule (298), are able to control *E. multilocularis* secondary infection.

In the per-oral model of infection, which better mimics AE in humans, the potential of FoxpP3 as a good target for application in immunotherapy has been confirmed (299). Another promising candidate is the programmed death-1 (PD-1)/PD-ligand 1 (PD-L1) signaling pathway, which plays a critical role in the induction of Foxp3+ CD25+ CD4+ Tregs, positively influences IL-10 and TGF- β secretion, inhibits effector T-cell proliferation and activation, and prevents Th1 cytokine production (300). Elevated soluble PD-L1 levels (301) and increased number of PD-1-expressing follicular helper T cells were observed in patients with CE compared with healthy controls (302); in experimental AE, percentages of PD-1+ Tregs and PD-L1+ dendritic cells (DCs) increased significantly together with levels of Foxp3, IL-10, and TGF- β during the chronic middle stage of infection (303). Preliminary experiments with a PD-1/PD-L1 engagement blockade are promising (304). Several PD-1/PD-L1 inhibitors already used in cancer treatment (305) are available to clinicians for pilot immunotherapeutic trials in AE. Combined anti-infective and immune therapy could also help clinicians in the management of severe, multiorgan cases of CE.

The specific immunological profile of the chronic stage of Echinococcus spp. infections has attracted attention as an established tolerance state that could be used to alleviate deleterious effects of inflammatory reactions in a variety of clinical conditions. Concomitant E. multilocularis infection in the rat delays rejection of a liver allograft (306), whereas concomitant E. granulosus sensu lato infection also reduces ovalbumin-induced airway inflammation of mice (307); in both situations, the effects were associated with raised IL-10 levels in the experimental animals. Although the potential efficacy of Echinococcus sp. components to treat rheumatoid arthritis was also evoked, it does not seem to have been demonstrated yet (308). The strongest evidence for the immunoregulatory role of established Echinococcus sp. infection in its murine intermediate host on an inflammatory disease has come from observations in experimental colitis. Both E. granulosus sensu stricto infection (309) and E. multilocularis infection (310) are able to reduce the development of dextran sulfate sodium (DSS)-induced colitis in mice. The possible use of noninfective Echinococcus sp. extracts is supported by observations made after treating mice daily, starting 3 days before colitis induction, with extracts from E. granulosus sensu stricto laminated layer; the treatment significantly improved the clinical symptoms and intestinal histological scores and maintained mucus production by goblet cells, while causing a significant decrease in gamma interferon (IFN- γ) and TNF- α and an increase in IL-10 production (311).

The immunomodulatory properties of *Echinococcus* sp. laminated layer have been well studied by reference to the shift they may induce in the immune response of the host to enhance metacestode growth and thus reduce host protection (76, 312). A variety of immunomodulating molecules produced by *Echinococcus* spp. have been identified, including antigen B (AgB) subclasses, Eg2 heat shock protein (Hsp) 70, and EgTeg from *E. granulosus sensu lato* (313) and Em2 (G11), EmAP, and *E. multilocularis* activin-like (EmACT) from *E. multilocularis* (290); they should certainly be reconsidered with a positive view to obtain the best combination of immunomodulating components to be used in inflammatory bowel diseases and, more generally, in all clinical situations that require tolerance induction.

Improving Vaccine Development

Vaccination of intermediate hosts. Vaccination of intermediate hosts of *E. granulosus* with the EG95 antigen has resulted in remarkable protective efficacy in pilot and field trials and is currently being used in areas of endemicity in China and South America (17, 314–317). The *Echinococcus* oncosphere is the infective stage for humans and intermediate hosts. Products of other genes differentially expressed by this stage likely represent potential additional vaccine candidates given that the protein expressed by the oncosphere-specific *eg95* gene induces a high level of protection against egg challenge infection in sheep and cattle (317, 318). Gene transcript analysis revealed that *eg95* is highly expressed in oncospheres (261), and recent studies show that *eg95* comprises a family of 7 distinct genes. Gene transcript analysis also showed that 340 (out of 3,811) genes were highly up-regulated in oncospheres compared with those in the adult and cyst stages of *E. granulosus* (261). Of the oncosphere-expressed genes, 2% (74/3,811) encode secreted proteins which likely play a key role in the penetration of the hatched oncosphere through the mammalian intestinal wall and in subsequent oncospheral development.

Vaccination of definitive hosts. A dog vaccine effective against adult Echinococcus sp. infection would be highly desirable as an intervention in integrated echinococcosis control. Such a vaccine is not currently available. The protoscolex is the stage which develops into an adult worm in the canine intestine. Proteins of genes highly expressed in the protoscolex or in the adult may provide suitable vaccine candidates against adult worms in the definitive host. Products of a novel, highly expressed eqM gene family (eqM4, eqM9, and eqM123) in mature adult worms, which may be associated with adult worm maturation and/or egg development, showed encouraging protective efficacy against adult worm infection in vaccine trials where dogs were vaccinated and necropsied 45 days after challenge infection (319, 320). Adult Echinococcus worms are localized to the middle of the small intestines of their definitive hosts, where abundant nutrients, especially amino acids, are present together with high levels of trypsin and trypsin-related enzymes. The worms secrete serine protease inhibitors (serpins) which counteract the potentially lethal effects of these host intestinal proteases and thereby likely play a key protective role in preventing proteolytic enzyme attack, ensuring survival of E. granulosus within its canine hosts. These, along with molecular chaperones, neurotransmitter receptors and transporters, and other protease inhibitors, specifically expressed in adult worms, likely represent additional vaccine candidates that warrant future study (261, 285, 321-323).

CONCLUSIONS AND FUTURE PERSPECTIVES

Awareness of clinicians and medical researchers of the public health importance of echinococcosis, even in areas where it is not endemic, is crucial, and an improved knowledge of echinococcosis imaging is essential for diagnosis and a prerequisite for multidisciplinary decisions on treatment strategy. For diagnostic confirmation, standardization and quality control of the currently available serological tests, both for diagnosis and for disease monitoring, is more of a priority than an everlasting quest for the "perfect antigen"; an easier recourse to molecular identification of the parasites ensures a more rapid and reliable species diagnosis.

For the care management of both CE and AE, a new concept, akin to that considered for cancer patients, has emerged, and the issue of recurrence, and thus of prolonged patient follow-up, is now taken seriously in CE. New surgical techniques make the complete resection of CE cysts easier and complete resection of AE lesions possible even in very advanced cases. The currently available techniques of nonsurgical interventional treatments have improved the quality of life of patients. However, prospective studies with prolonged follow-up are still needed to base echinococcosis therapeutic strategy on evidence. In addition, more than 30 years after the first trials of mebendazole and ABZ, there is no available alternative to these two drugs as anti-infective therapy, a situation unique in the field of infectious diseases. Fortunately, new biological or immunological therapeutic targets may now be more easily identified

because of new proteomics information, the complete sequencing of the *E. granulosus* and *E. multilocularis* genomes, and increased understanding of the host-parasite interactions in both AE and CE.

Controlling the transmission of *Echinococcus* spp. continues to be a considerable obstacle, but precise identification of the infecting species/genotypes may help public health institutions better focus and optimize the effectiveness of control programs. Involvement of wild animals in the life cycle of all *Echinococcus* species makes disease control dependent on landscape and climate changes and is, consequently, more challenging now than hitherto. Important improvements in molecular biology-based tests to detect *Echinococcus* spp. in definitive hosts and in the environment, however, make control program monitoring potentially easier. One challenge in the control of AE and CE in coming years will be to define optimum targets for developing vaccines effective in definitive canine hosts to interrupt the chain of transmission to humans if echinococcosis elimination, slated for 2050 by WHO, is to be achieved.

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REFERENCES

- McManus DP, Gray DJ, Zhang W, Yang Y. 2012. Diagnosis, treatment, and management of echinococcosis. BMJ 344:e3866. https://doi.org/10 .1136/bmi.e3866.
- Craig PS, Larrieu E. 2006. Control of cystic echinococcosis/hydatidosis: 1863-2002. Adv Parasitol 61:443–508. https://doi.org/10.1016/S0065 -308X(05)61011-1.
- Craig PS, McManus DP, Lightowlers MW, Chabalgoity JA, Garcia HH, Gavidia CM, Gilman RH, Gonzalez AE, Lorca M, Naquira C, Nieto A, Schantz PM. 2007. Prevention and control of cystic echinococcosis. Lancet Infect Dis 7:385–394. https://doi.org/10.1016/S1473-3099 (07)70134-2.
- 4. Schweiger A, Ammann RW, Candinas D, Clavien PA, Eckert J, Gottstein B, Halkic N, Muellhaupt B, Prinz BM, Reichen J, Tarr PE, Torgerson PR, Deplazes P. 2007. Human alveolar echinococcosis after fox population increase, Switzerland. Emerg Infect Dis 13:878–882. https://doi.org/10.3201/eid1306.061074.
- Budke CM, Carabin H, Ndimubanzi PC, Nguyen H, Rainwater E, Dickey M, Bhattarai R, Zeziulin O, Qian MB. 2013. A systematic review of the literature on cystic echinococcosis frequency worldwide and its associated clinical manifestations. Am J Trop Med Hyg 88:1011–1027. https://doi.org/10.4269/ajtmh.12-0692.
- Budke CM, Deplazes P, Torgerson PR. 2006. Global socioeconomic impact of cystic echinococcosis. Emerg Infect Dis 12:296–303. https:// doi.org/10.3201/eid1202.050499.
- Torgerson PR, Keller K, Magnotta M, Ragland N. 2010. The global burden of alveolar echinococcosis. PLoS Negl Trop Dis 4:e722. https:// doi.org/10.1371/journal.pntd.0000722.
- Frider B, Larrieu E, Odriozola M. 1999. Long-term outcome of asymptomatic liver hydatidosis. J Hepatol 30:228–231. https://doi.org/10.1016/S0168-8278(99)80066-X.
- 9. Wang Y, He T, Wen X, Li T, Waili A, Zhang W, Xu X, Vuitton DA, Rogan MT, Wen H, Craig PS. 2006. Post-survey follow-up for human cystic echinococcosis in northwest China. Acta Trop 98:43–51. https://doi.org/10.1016/j.actatropica.2006.01.009.
- Zhang W, Zhang Z, Wu W, Shi B, Li J, Zhou X, Wen H, McManus DP. 2015. Epidemiology and control of echinococcosis in central Asia, with particular reference to the People's Republic of China. Acta Trop 141: 235–243. https://doi.org/10.1016/j.actatropica.2014.03.014.
- Craig PS, Li T, Qiu J, Zhen R, Wang Q, Giraudoux P, Ito A, Heath D, Warnock B, Schantz P, Yang W. 2008. Echinococcosis and Tibetan communities. Emerg Infect Dis 14:1674–1675. https://doi.org/10.3201/ eid1410.071636.

- 12. Alvarez Rojas CA, Romig T, Lightowlers MW. 2014. *Echinococcus granulosus* sensu lato genotypes infecting humans—review of current knowledge. Int J Parasitol 44:9–18. https://doi.org/10.1016/j.ijpara.2013 .08.008.
- Deplazes P, Rinaldi L, Alvarez Rojas CA, Torgerson PR, Harandi MF, Romig T, Antolova D, Schurer JM, Lahmar S, Cringoli G, Magambo J, Thompson RC, Jenkins EJ. 2017. Global distribution of alveolar and cystic echinococcosis. Adv Parasitol 95:315–493. https://doi.org/10 .1016/bs.apar.2016.11.001.
- Larrieu E, Zanini F. 2012. Critical analysis of cystic echinococcosis control programs and praziquantel use in South America, 1974-2010. Rev Panam Salud Publica 31:81–87. https://doi.org/10.1590/S1020 -49892012000100012.
- Pavletic CF, Larrieu E, Guarnera EA, Casas N, Irabedra P, Ferreira C, Sayes J, Gavidia CM, Caldas E, Lise MLZ, Maxwell M, Arezo M, Navarro AM, Vigilato MAN, Cosivi O, Espinal M, Vilas V. 2017. Cystic echinococcosis in South America: a call for action. Rev Panam Salud Publica 41:e42.
- Cucher MA, Macchiaroli N, Baldi G, Camicia F, Prada L, Maldonado L, Avila HG, Fox A, Gutierrez A, Negro P, Lopez R, Jensen O, Rosenzvit M, Kamenetzky L. 2016. Cystic echinococcosis in South America: systematic review of species and genotypes of *Echinococcus granulosus* sensu lato in humans and natural domestic hosts. Trop Med Int Health 21:166–175. https://doi.org/10.1111/tmi.12647.
- Craig PS, Hegglin D, Lightowlers MW, Torgerson PR, Wang Q. 2017.
 Echinococcosis: control and prevention. Adv Parasitol 96:55–158.
 https://doi.org/10.1016/bs.apar.2016.09.002.
- Jenkins DJ, Lievaart JJ, Boufana B, Lett WS, Bradshaw H, Armua-Fernandez MT. 2014. Echinococcus granulosus and other intestinal helminths: current status of prevalence and management in rural dogs of eastern Australia. Aust Vet J 92:292–298. https://doi.org/10.1111/avj .12218.
- Rossi P, Tamarozzi F, Galati F, Pozio E, Akhan O, Cretu CM, Vutova K, Siles-Lucas M, Brunetti E, Casulli A. 2016. The first meeting of the European Register of Cystic Echinococcosis (ERCE). Parasit Vectors 9:243. https://doi.org/10.1186/s13071-016-1532-3.
- Kawamura N, Kamiyama T, Sato N, Nakanishi K, Yokoo H, Kamachi H, Tahara M, Yamaga S, Matsushita M, Todo S. 2011. Long-term results of hepatectomy for patients with alveolar echinococcosis: a single-center experience. J Am Coll Surg 212:804–812. https://doi.org/10.1016/j .jamcollsurg.2011.02.007.
- Irie T, Mukai T, Yagi K. 2018. Echinococcus multilocularis surveillance using copro-DNA and egg examination of shelter dogs from an en-

demic area in Hokkaido, Japan. Vector Borne Zoonotic Dis 18:390–392. https://doi.org/10.1089/vbz.2017.2245.

- Vuitton DA, Wang Q, Zhou HX, Raoul F, Knapp J, Bresson-Hadni S, Wen H, Giraudoux P. 2011. A historical view of alveolar echinococcosis, 160 years after the discovery of the first case in humans. 1. What have we learnt on the distribution of the disease and on its parasitic agent? Chin Med J (Engl) 124:2943–2953.
- Feng X, Qi X, Yang L, Duan X, Fang B, Gongsang Q, Bartholomot B, Vuitton DA, Wen H, Craig PS. 2015. Human cystic and alveolar echinococcosis in the Tibet Autonomous Region (TAR), China. J Helminthol 89:671–679. https://doi.org/10.1017/S0022149X15000656.
- Cai H, Guan Y, Ma X, Wang L, Wang H, Su G, Zhang X, Han X, Ma J, Liu YF, Li J, Zhang J, Wang Y, Wang W, Du R, Lei W, Wu W. 2017. Epidemiology of echinococcosis among schoolchildren in Golog Tibetan Autonomous Prefecture, Qinghai, China. Am J Trop Med Hyg 96:674–679. https://doi.org/10.4269/ajtmh.16-0479.
- Craig P. 2003. Echinococcus multilocularis. Curr Opin Infect Dis 16: 437–444. https://doi.org/10.1097/01.qco.0000092815.64370.39.
- Abdybekova A, Sultanov A, Karatayev B, Zhumabayeva A, Shapiyeva Z, Yeshmuratov T, Toksanbayev D, Shalkeev R, Torgerson PR. 2015. Epidemiology of echinococcosis in Kazakhstan: an update. J Helminthol 89:647–650. https://doi.org/10.1017/S0022149X15000425.
- Torgerson PR. 2013. The emergence of echinococcosis in central Asia. Parasitology 140:1667–1673. https://doi.org/10.1017/S0031 182013000516.
- Counotte MJ, Minbaeva G, Usubalieva J, Abdykerimov K, Torgerson PR. 2016. The burden of zoonoses in Kyrgyzstan: a systematic review. PLoS Negl Trop Dis 10:e0004831. https://doi.org/10.1371/journal.pntd.0004831.
- Usubalieva J, Minbaeva G, Ziadinov I, Deplazes P, Torgerson PR. 2013.
 Human alveolar echinococcosis in Kyrgyzstan. Emerg Infect Dis 19: 1095–1097. https://doi.org/10.3201/eid1907.121405.
- Beerli O, Guerra D, Baltrunaite L, Deplazes P, Hegglin D. 2017. Microtus arvalis and Arvicola scherman: key players in the Echinococcus multilocularis life cycle. Front Vet Sci 4:216. https://doi.org/10.3389/fvets .2017.00216.
- Conraths FJ, Deplazes P. 2015. Echinococcus multilocularis: Epidemiology, surveillance and state-of-the-art diagnostics from a veterinary public health perspective. Vet Parasitol 213:149–161. https://doi.org/10.1016/j.vetpar.2015.07.027.
- Bagrade G, Deksne G, Ozolina Z, Howlett SJ, Interisano M, Casulli A, Pozio E. 2016. Echinococcus multilocularis in foxes and raccoon dogs: an increasing concern for Baltic countries. Parasit Vectors 9:615. https:// doi.org/10.1186/s13071-016-1891-9.
- Berke O, Romig T, von Keyserlingk M. 2008. Emergence of *Echinococcus multilocularis* among red foxes in northern Germany, 1991-2005. Vet Parasitol 155:319–322. https://doi.org/10.1016/j.vetpar.2008.05.017.
- Vuitton DA, Demonmerot F, Knapp J, Richou C, Grenouillet F, Chauchet A, Vuitton L, Bresson-Hadni S, Millon L. 2015. Clinical epidemiology of human AE in Europe. Vet Parasitol 213:110–120. https://doi.org/10 .1016/j.vetpar.2015.07.036.
- Marcinkutė A, Šarkūnas M, Moks E, Saarma U, Jokelainen P, Bagrade G, Laivacuma S, Strupas K, Sokolovas V, Deplazes P. 2015. Echinococcus infections in the Baltic region. Vet Parasitol 213:121–131. https://doi.org/10.1016/j.vetpar.2015.07.032.
- Gottstein B, Stojkovic M, Vuitton DA, Millon L, Marcinkute A, Deplazes P. 2015. Threat of alveolar echinococcosis to public health—a challenge for Europe. Trends Parasitol 31:407–412. https://doi.org/10.1016/j.pt.2015.06.001.
- Massolo A, Liccioli S, Budke C, Klein C. 2014. Echinococcus multilocularis in North America: the great unknown. Parasite 21:73. https://doi.org/ 10.1051/parasite/2014069.
- 38. Catalano S, Lejeune M, Liccioli S, Verocai GG, Gesy KM, Jenkins EJ, Kutz SJ, Fuentealba C, Duignan PJ, Massolo A. 2012. *Echinococcus multilocularis* in urban coyotes, Alberta, Canada. Emerg Infect Dis 18:1625–1628. https://doi.org/10.3201/eid.1810.120119.
- Romig T, Deplazes P, Jenkins D, Giraudoux P, Massolo A, Craig PS, Wassermann M, Takahashi K, de la Rue M. 2017. Ecology and life cycle patterns of *Echinococcus* species. Adv Parasitol 95:213–314. https://doi.org/10.1016/bs.apar.2016.11.002.
- Eckert J, Thompson RC. 2017. Historical aspects of echinococcosis. Adv Parasitol 95:1–64. https://doi.org/10.1016/bs.apar.2016.07.003.
- Debourgogne A, Blanchet D, Fior A, Umhang G, Simon S, Aznar C. 2017.
 Neotropical echinococcosis caused by Echinococcus vogeli in a 6-year-old

- child: the second case report in humans in French Guiana. Paediatr Int Child Health 37:63–65. https://doi.org/10.1179/2046905515Y.0000000054.
- 42. Knapp J, Chirica M, Simonnet C, Grenouillet F, Bart JM, Sako Y, Itoh S, Nakao M, Ito A, Millon L. 2009. *Echinococcus vogeli* infection in a hunter, French Guiana. Emerg Infect Dis 15:2029–2031. https://doi.org/10.3201/eid1512.090940.
- Siles-Lucas M, Casulli A, Conraths FJ, Müller N. 2017. Laboratory diagnosis of *Echinococcus* spp. in human patients and infected animals. Adv Parasitol 96:159–257. https://doi.org/10.1016/bs.apar.2016.09.003.
- 44. McManus DP. 2006. Molecular discrimination of taeniid cestodes. Parasitol Int 55:S31-37. https://doi.org/10.1016/j.parint.2005.11.004.
- Nakao M, McManus DP, Schantz PM, Craig PS, Ito A. 2007. A molecular phylogeny of the genus *Echinococcus* inferred from complete mitochondrial genomes. Parasitology 134:713–722. https://doi.org/10.1017/ S0031182006001934.
- Thompson RC. 2008. The taxonomy, phylogeny and transmission of Echinococcus. Exp Parasitol 119:439 – 446. https://doi.org/10.1016/j .exppara.2008.04.016.
- Romig T, Dinkel A, Mackenstedt U. 2006. The present situation of echinococcosis in Europe. Parasitol Int 55:S187–S191. https://doi.org/ 10.1016/j.parint.2005.11.028.
- 48. Nakao M, Lavikainen A, Yanagida T, Ito A. 2013. Phylogenetic systematics of the genus *Echinococcus* (Cestoda: Taeniidae). Int J Parasitol 43:1017–1029. https://doi.org/10.1016/j.ijpara.2013.06.002.
- 49. Romig T, Ebi D, Wassermann M. 2015. Taxonomy and molecular epidemiology of *Echinococcus granulosus* sensu lato. Vet Parasitol 213: 76–84. https://doi.org/10.1016/j.vetpar.2015.07.035.
- Valot B, Knapp J, Umhang G, Grenouillet F, Millon L. 2015. Genomic characterization of EmsB microsatellite loci in *Echinococcus multilocularis*. Infect Genet Evol 32:338–341. https://doi.org/10.1016/j.meegid 2015.03.040
- Wu C, Zhang W, Ran B, Fan H, Wang H, Guo B, Zhou C, Shao Y, Zhang W, Giraudoux P, Knapp J, Wen H, Kuang L, Li J. 2017. Genetic variation of mitochondrial genes among *Echinococcus multilocularis* isolates collected in western China. Parasit Vectors 10:265. https://doi.org/10.1186/s13071-017-2172-v.
- Knapp J, Gottstein B, Saarma U, Millon L. 2015. Taxonomy, phylogeny and molecular epidemiology of *Echinococcus multilocularis*: from fundamental knowledge to health ecology. Vet Parasitol 213:85–91. https://doi.org/10.1016/j.vetpar.2015.07.030.
- 53. Afonso E, Knapp J, Tete N, Umhang G, Rieffel D, van Kesteren F, Ziadinov I, Craig PS, Torgerson PR, Giraudoux P. 2015. Echinococcus multilocularis in Kyrgyzstan: similarity in the Asian EmsB genotypic profiles from village populations of Eastern mole voles (Ellobius tancrei) and dogs in the Alay valley. J Helminthol 89:664–670. https://doi.org/10.1017/S0022149X15000474.
- 54. Umhang G, Knapp J, Hormaz V, Raoul F, Boue F. 2014. Using the genetics of *Echinococcus multilocularis* to trace the history of expansion from an endemic area. Infect Genet Evol 22:142–149. https://doi.org/10.1016/j.meegid.2014.01.018.
- Knapp J, Staebler S, Bart JM, Stien A, Yoccoz NG, Drogemuller C, Gottstein B, Deplazes P. 2012. Echinococcus multilocularis in Svalbard, Norway: microsatellite genotyping to investigate the origin of a highly focal contamination. Infect Genet Evol 12:1270–1274. https://doi.org/ 10.1016/j.meegid.2012.03.008.
- Boufana B, Qiu J, Chen X, Budke CM, Campos-Ponce M, Craig PS. 2013.
 First report of *Echinococcus shiquicus* in dogs from eastern Qinghai-Tibet plateau region, China. Acta Trop 127:21–24. https://doi.org/10.1016/j.actatropica.2013.02.019.
- 57. Xiao N, Qiu J, Nakao M, Li T, Yang W, Chen X, Schantz PM, Craig PS, Ito A. 2005. *Echinococcus shiquicus* n. sp., a taeniid cestode from Tibetan fox and plateau pika in China. Int J Parasitol 35:693–701. https://doi.org/10.1016/j.ijpara.2005.01.003.
- Carmena D, Cardona GA. 2014. Echinococcosis in wild carnivorous species: epidemiology, genotypic diversity, and implications for veterinary public health. Vet Parasitol 202:69–94. https://doi.org/10.1016/j .vetpar.2014.03.009.
- Fan YL, Lou ZZ, Li L, Yan HB, Liu QY, Zhan F, Li JQ, Liu CN, Cai JZ, Lei MT, Shi WG, Yang YR, McManus DP, Jia WZ. 2016. Genetic diversity in *Echinococcus shiquicus* from the plateau pika (*Ochotona curzoniae*) in Darlag County, Qinghai, China. Infect Genet Evol 45:408–414. https://doi.org/10.1016/j.meegid.2016.06.016.
- Santos GB, Soares MDCP, Brito EMDF, Rodrigues AL, Siqueira NG, Gomes-Gouvêa MS, Alves MM, Carneiro LA, Malheiros AP, Póvoa MM,

Zaha A, Haag KL. 2012. Mitochondrial and nuclear sequence polymorphisms reveal geographic structuring in Amazonian populations of *Echinococcus vogeli* (Cestoda: Taeniidae). Int J Parasitol 42:1115–1118. https://doi.org/10.1016/j.ijpara.2012.10.010.

- 61. Soares MC, Rodrigues AL, Moreira Silva CA, Brito EM, Gomes-Gouvêa MS, Corrêa IR, Pinho JR, Malheiros AP, Nunes HM, Póvoa MM. 2013. Anatomo-clinical and molecular description of liver neotropical echinococcosis caused by *Echinococcus oligarthrus* in human host. Acta Trop 125:110–114. https://doi.org/10.1016/j.actatropica.2012.09.004.
- Arrabal JP, Avila HG, Rivero MR, Camicia F, Salas MM, Costa SA, Nocera CG, Rosenzvit MC, Kamenetzky L. 2017. *Echinococcus oligarthrus* in the subtropical region of Argentina: first integration of morphological and molecular analyses determines two distinct populations. Vet Parasitol 240:60 – 67. https://doi.org/10.1016/j.vetpar.2017.03.019.
- Kern P, Menezes da Silva A, Akhan O, Mullhaupt B, Vizcaychipi KA, Budke C, Vuitton DA. 2017. The echinococcoses: diagnosis, clinical management and burden of disease. Adv Parasitol 96:259–369. https:// doi.org/10.1016/bs.apar.2016.09.006.
- 64. Vuitton DA. 2004. Echinococcosis and allergy. Clin Rev Allergy Immunol 26:93–104. https://doi.org/10.1007/s12016-004-0004-2.
- 65. Vuitton DA, Bresson-Hadni S, Lenys D, Flausse F, Liance M, Wattre P, Miguet JP, Capron A. 1988. IgE-dependent humoral immune response in *Echinococcus multilocularis* infection: circulating and basophil-bound specific IgE against *Echinococcus* antigens in patients with alveolar echinococcosis. Clin Exp Immunol 71:247–252.
- Kilimcioğlu AA, Girginkardeşler N, Korkmaz M, Özkol M, Düzgün F, Östan I, Pabuşcu Y, Dinç G, Ok UZ. 2013. A mass screening survey of cystic echinococcosis by ultrasonography, Western blotting, and ELISA among university students in Manisa, Turkey. Acta Trop 128:578–583. https://doi.org/10.1016/j.actatropica.2013.08.010.
- Ran B, Shao Y, Guo Y, Yimiti Y, Aji T, Jia J, Shayiding P, Jiang T, Cheng L, Li J, McManus DP, Zhang W, Wen H. 2016. Surgical treatment of hepatic cystic echinococcosis in patients co-infected with HIV/AIDS. J Helminthol 90:125–128. https://doi.org/10.1017/S0022149X15000188.
- 68. Erayman I, Kalkan E, Erdi F, Kerimoglu U, Esen H. 2011. Primary spinal hydatid cyst in a patient with acquired immunodeficiency syndrome. Eur Spine J 20:235. https://doi.org/10.1007/s00586-010-1614-4.
- Oikonomopoulou K, Yu H, Wang Z, Vasiliou SK, Brinc D, Christofi G, Theodorou M, Pavlou P, Hadjisavvas A, Demetriou CA, Kyriacou K, Diamandis EP. 2016. Association between *Echinococcus granulosus* infection and cancer risk—a pilot study in Cyprus. Clin Chem Lab Med 54:1955–1961.
- Turhan N, Esendagli G, Ozkayar O, Tunali G, Sokmensuer C, Abbasoglu O. 2015. Co-existence of *Echinococcus granulosus* infection and cancer metastasis in the liver correlates with reduced Th1 immune responses. Parasite Immunol 37:16–22. https://doi.org/10.1111/pim.12152.
- Chauchet A, Grenouillet F, Knapp J, Richou C, Delabrousse E, Dentan C, Millon L, Di Martino V, Contreras R, Deconinck E, Blagosklonov O, Vuitton DA, Bresson-Hadni S. 2014. Increased incidence and characteristics of alveolar echinococcosis in patients with immunosuppressionassociated conditions. Clin Infect Dis 59:1095–1104. https://doi.org/10 .1093/cid/ciu520.
- 72. Eiermann TH, Bettens F, Tiberghien P, Schmitz K, Beurton I, Bresson-Hadni S, Ammann RW, Goldmann SF, Vuitton DA, Gottstein B, Kern P. 1998. HLA and alveolar echinococcosis. Tissue Antigens 52:124–129. https://doi.org/10.1111/j.1399-0039.1998.tb02275.x.
- Zhang S, Penfornis A, Harraga S, Chabod J, Beurton I, Bresson-Hadni S, Tiberghien P, Kern P, Vuitton DA. 2003. Polymorphisms of the TAP1 and TAP2 genes in human alveolar echinococcosis. Eur J Immunogenet 30:133–139. https://doi.org/10.1046/j.1365-2370.2003.00375.x.
- Vuitton DA, Zhang SL, Yang Y, Godot V, Beurton I, Mantion G, Bresson-Hadni S. 2006. Survival strategy of *Echinococcus multilocularis* in the human host. Parasitol Int 55:S51-55. https://doi.org/10.1016/j.parint.2005.11.007.
- Godot V, Harraga S, Beurton I, Tiberghien P, Sarciron E, Gottstein B, Vuitton DA. 2000. Resistance/susceptibility to *Echinococcus multilocularis* infection and cytokine profile in humans. II. Influence of the HLA B8, DR3, DQ2 haplotype. Clin Exp Immunol 121:491–498. https://doi.org/10.1046/j.1365-2249.2000.01309.x.
- Vuitton DA, Gottstein B. 2010. Echinococcus multilocularis and its intermediate host: a model of parasite-host interplay. J Biomed Biotechnol 2010:923193. https://doi.org/10.1155/2010/923193.
- 77. Koch S, Bresson-Hadni S, Miguet JP, Crumbach JP, Gillet M, Mantion GA, Heyd B, Vuitton DA, Minello A, Kurtz S. 2003. Experience of liver

- transplantation for incurable alveolar echinococcosis: a 45-case European collaborative report. Transplantation 75:856–863. https://doi.org/10.1097/01.TP.0000054230.63568.79.
- Sailer M, Soelder B, Allerberger F, Zaknun D, Feichtinger H, Gottstein B. 1997. Alveolar echinococcosis of the liver in a six-year-old girl with acquired immunodeficiency syndrome. J Pediatr 130:320–323. https:// doi.org/10.1016/S0022-3476(97)70364-0.
- Geyer M, Wilpert J, Wiech T, Theilacker C, Stubanus M, Kramer-Zucker A, Fischer KG, Drognitz O, Frydrychowicz A, Kern W, Walz G, Pisarski P. 2011. Rapidly progressive hepatic alveolar echinococcosis in an ABOincompatible renal transplant recipient. Transpl Infect Dis 13:278–284. https://doi.org/10.1111/j.1399-3062.2010.00583.x.
- Kern P, Gruner B, Wahlers K. 2011. Diagnosis and course of echinococcocal diseases in the transplant setting. Transpl Infect Dis 13:217–221. https://doi.org/10.1111/j.1399-3062.2011.00643.x.
- 81. Dentan C, Mazet R, Gilson M, Marchou-Lopez S, Gaudin P. 2012. Rheumatoid arthritis, alveolar echinococcosis, and rituximab: a case report. Joint Bone Spine 79:325–327. https://doi.org/10.1016/j.jbspin.2011.10.014.
- 82. Tamarozzi F, Akhan O, Cretu CM, Vutova K, Akinci D, Chipeva R, Ciftci T, Constantin CM, Fabiani M, Golemanov B, Janta D, Mihailescu P, Muhtarov M, Orsten S, Petrutescu M, Pezzotti P, Popa AC, Popa LG, Popa MI, Velev V, Siles-Lucas M, Brunetti E, Casulli A. 2018. Prevalence of abdominal cystic echinococcosis in rural Bulgaria, Romania, and Turkey: a cross-sectional, ultrasound-based, population study from the HERACLES project. Lancet Infect Dis 18:769–778. https://doi.org/10.1016/S1473-3099(18)30221-4.
- Bresson-Hadni S, Delabrousse E, Blagosklonov O, Bartholomot B, Koch S, Miguet J-P, André Mantion G, Angèle Vuitton D. 2006. Imaging aspects and non-surgical interventional treatment in human alveolar echinococcosis. Parasitol Int 55:S267. https://doi.org/10.1016/j.parint .2005.11.053.
- Solomon N, Zeyhle E, Subramanian K, Fields PJ, Romig T, Kern P, Carter JY, Wachira J, Mengiste A, Macpherson CNL. 2018. Cystic echinococcosis in Turkana, Kenya: 30 years of imaging in an endemic region. Acta Trop 178:182–189. https://doi.org/10.1016/j.actatropica.2017.11.006.
- Piarroux M, Piarroux R, Giorgi R, Knapp J, Bardonnet K, Sudre B, Watelet J, Dumortier J, Gerard A, Beytout J, Abergel A, Mantion G, Vuitton DA, Bresson-Hadni S. 2011. Clinical features and evolution of alveolar echinococcosis in France from 1982 to 2007: results of a survey in 387 patients. J Hepatol 55:1025–1033. https://doi.org/10.1016/j.jhep.2011 .02.018.
- Brunetti E, Kern P, Vuitton DA. 2010. Expert consensus for the diagnosis and treatment of cystic and alveolar echinococcosis in humans. Acta Trop 114:1–16. https://doi.org/10.1016/j.actatropica.2009.11.001.
- Kern P. 2010. Clinical features and treatment of alveolar echinococcosis. Curr Opin Infect Dis 23:505–512. https://doi.org/10.1097/QCO.0b013e32833d7516.
- 88. Kern P, Wen H, Sato N, Vuitton DA, Gruener B, Shao Y, Delabrousse E, Kratzer W, Bresson-Hadni S. 2006. WHO classification of alveolar echinococcosis: principles and application. Parasitol Int 55:S283–S287. https://doi.org/10.1016/j.parint.2005.11.041.
- 89. Junghanss T, da Silva AM, Horton J, Chiodini PL, Brunetti E. 2008. Clinical management of cystic echinococcosis: state of the art, problems, and perspectives. Am J Trop Med Hyg 79:301–311. https://doi.org/10.4269/ajtmh.2008.79.301.
- Kratzer W, Gruener B, Kaltenbach TE, Ansari-Bitzenberger S, Kern P, Fuchs M, Mason RA, Barth TF, Haenle MM, Hillenbrand A, Oeztuerk S, Graeter T. 2015. Proposal of an ultrasonographic classification for hepatic alveolar echinococcosis: Echinococcosis multilocularis Ulm classification-ultrasound. World J Gastroenterol 21:12392–12402. https://doi.org/10.3748/wjg.v21.i43.12392.
- Graeter T, Kratzer W, Oeztuerk S, Haenle MM, Mason RA, Hillenbrand A, Kull T, Barth TF, Kern P, Gruener B. 2016. Proposal of a computed tomography classification for hepatic alveolar echinococcosis. World J Gastroenterol 22:3621–3631. https://doi.org/10.3748/wjg.v22.i13.3621.
- 92. Song T, Qin Z, Hao W, Yongquan L, Lanhui Y, Lei Y. 2011. Usefulness of gray-scale contrast-enhanced ultrasonography (SonoVue(R)) in diagnosing hepatic alveolar echinococcosis. Ultrasound Med Biol 37: 1024–1028. https://doi.org/10.1016/j.ultrasmedbio.2011.04.014.
- Cai DM, Wang HY, Wang XL, Jiang Y, Luo Y, Li YZ. 2017. Ultrasonographic findings of small lesion of hepatic alveolar echinococcosis. Acta Trop 174:165–170. https://doi.org/10.1016/j.actatropica.2016.01.030.
- 94. Ehrhardt AR, Reuter S, Buck AK, Haenle MM, Mason RA, Gabelmann A,

Kern P, Kratzer W. 2007. Assessment of disease activity in alveolar echinococcosis: a comparison of contrast enhanced ultrasound, three-phase helical CT and [(18)F] fluorodeoxyglucose positron emission tomography. Abdom Imaging 32:730–736. https://doi.org/10.1007/s00261-007-9177-5.

- 95. Qin Y, Li X, Zhang Q, Xie B, Ji X, Li Y, Yiblayan A, Wen H. 2016. Analysis of the clinical value of 18F-FDG PET/CT in hepatic alveolar echinococcosis before and after autologous liver transplantation. Exp Ther Med 11:43–48. https://doi.org/10.3892/etm.2015.2857.
- Rolle AM, Soboslay PT, Reischl G, Hoffmann WH, Pichler BJ, Wiehr S. 2015. Evaluation of the metabolic activity of *Echinococcus multilocularis* in rodents using positron emission tomography tracers. Mol Imaging Biol 17:512–520. https://doi.org/10.1007/s11307-014-0815-3.
- Liu W, Delabrousse É, Blagosklonov O, Wang J, Zeng H, Jiang Y, Wang J, Qin Y, Vuitton DA, Wen H. 2014. Innovation in hepatic alveolar echinococcosis imaging: best use of old tools, and necessary evaluation of new ones. Parasite 21:74. https://doi.org/10.1051/parasite/2014072.
- Azizi A, Blagosklonov O, Lounis A, Berthet L, Vuitton DA, Bresson-Hadni S, Delabrousse E. 2015. Alveolar echinococcosis: correlation between hepatic MRI findings and FDG-PET/CT metabolic activity. Abdom Imaging 40:56–63. https://doi.org/10.1007/s00261-014-0183-0.
- Kantarci M, Pirimoglu B, Ogul H, Bayraktutan U, Eren S, Aydinli B, Ozturk G, Karaca L. 2014. Can biliary-cyst communication be predicted by Gd-EOB-DTPA-enhanced MR cholangiography before treatment for hepatic hydatid disease? Clin Radiol 69:52–58. https://doi.org/10.1016/j .crad.2013.08.005.
- Stojkovic M, Junghanss T, Veeser M, Weber TF, Sauer P. 2016. Endoscopic treatment of biliary stenosis in patients with alveolar echinococcosis—report of 7 consecutive patients with serial ERC approach. PLoS Negl Trop Dis 10:e0004278. https://doi.org/10.1371/journal.pntd .0004278.
- 101. Bulakci M, Ilhan M, Bademler S, Yilmaz E, Gulluoglu M, Bayraktar A, Asik M, Guloglu R. 2016. Efficacy of ultrasound-guided core-needle biopsy in the diagnosis of hepatic alveolar echinococcosis: a retrospective analysis. Parasite 23:19. https://doi.org/10.1051/parasite/2016019.
- 102. Barth TFE, Herrmann TS, Tappe D, Stark L, Grüner B, Buttenschoen K, Hillenbrand A, Juchems M, Henne-Bruns D, Kern P, Seitz HM, Möller P, Rausch RL, Kern P, Deplazes P. 2012. Sensitive and specific immuno-histochemical diagnosis of human alveolar echinococcosis with the monoclonal antibody Em2G11. PLoS Negl Trop Dis 6:e1877. https://doi.org/10.1371/journal.pntd.0001877.
- 103. Zhang W, Li J, McManus DP. 2003. Concepts in immunology and diagnosis of hydatid disease. Clin Microbiol Rev 16:18–36. https://doi.org/10.1128/CMR.16.1.18-36.2003.
- 104. Ray R, De PK, Karak K. 2002. Combined role of Casoni test and indirect haemagglutination test in the diagnosis of hydatid disease. Indian J Med Microbiol 20:79–82.
- 105. Pagnozzi D, Addis MF, Biosa G, Roggio AM, Tedde V, Mariconti M, Tamarozzi F, Meroni V, Masu G, Masala G, Brunetti E, Uzzau S. 2016. Diagnostic accuracy of antigen 5-based ELISAs for human cystic echinococcosis. PLoS Negl Trop Dis 10:e0004585. https://doi.org/10.1371/journal.pntd.0004585.
- 106. Hernández-González A, Santivañez S, García HH, Rodríguez S, Muñoz S, Ramos G, Orduña A, Siles-Lucas M. 2012. Improved serodiagnosis of cystic echinococcosis using the new recombinant 2B2t antigen. PLoS Negl Trop Dis 6:e1714. https://doi.org/10.1371/journal.pntd.0001714.
- 107. Ortona E, Margutti P, Delunardo F, Nobili V, Profumo E, Rigano R, Buttari B, Carulli G, Azzara A, Teggi A, Bruschi F, Siracusano A. 2005. Screening of an *Echinococcus granulosus* cDNA library with IgG4 from patients with cystic echinococcosis identifies a new tegumental protein involved in the immune escape. Clin Exp Immunol 142:528–538. https://doi.org/10.1111/j.1365-2249.2005.02939.x.
- 108. Mahmoud MS, Abou Gamra MM. 2004. Alkaline phosphatase from *Echinococcus granulosus* metacestodes for immunodiagnosis of human cystic echinococcosis. J Egypt Soc Parasitol 34:865–879.
- 109. Fathi S, Jalousian F, Hosseini SH, Parsa H, Kordafshari S. 2016. A study of cross-reactivity between recombinant EPC1 antigen of *Echinococcus granulosus* in serum from patients with confirmed cystic echinococcosis infection and other parasitic infections. Am J Trop Med Hyg 94: 1313–1317. https://doi.org/10.4269/ajtmh.15-0680.
- Zhang W, Wen H, Li J, Lin R, McManus DP. 2012. Immunology and immunodiagnosis of cystic echinococcosis: an update. Clin Dev Immunol 2012:101895. https://doi.org/10.1155/2012/101895.
- 111. Gavidia CM, Gonzalez AE, Zhang W, McManus DP, Lopera L, Ninaquispe

- B, Garcia HH, Rodriguez S, Verastegui M, Calderon C, Pan WK, Gilman RH. 2008. Diagnosis of cystic echinococcosis, central Peruvian highlands. Emerg Infect Dis 14:260–266. https://doi.org/10.3201/eid1402.061101.
- 112. Tappe D, Frosch M, Sako Y, Itoh S, Gruner B, Reuter S, Nakao M, Ito A, Kern P. 2009. Close relationship between clinical regression and specific serology in the follow-up of patients with alveolar echinococcosis in different clinical stages. Am J Trop Med Hyg 80:792–797. https://doi.org/10.4269/ajtmh.2009.80.792.
- 113. Knapp J, Sako Y, Grenouillet F, Bresson-Hadni S, Richou C, Gbaguidi-Haore H, Ito A, Millon L. 2014. Comparison of the serological tests ICT and ELISA for the diagnosis of alveolar echinococcosis in France. Parasite 21:34. https://doi.org/10.1051/parasite/2014037.
- 114. Cisak E, Sroka J, Wójcik-Fatla A, Zając V, Dutkiewicz J. 2015. Evaluation of reactivity to *Echinococcus* spp. among rural inhabitants in Poland. Acta Parasitol 60:525–529. https://doi.org/10.1515/ap-2015-0074.
- 115. Bartholomot G, Vuitton DA, Harraga S, Shi da Z, Giraudoux P, Barnish G, Wang YH, MacPherson CN, Craig PS. 2002. Combined ultrasound and serologic screening for hepatic alveolar echinococcosis in central China. Am J Trop Med Hyg 66:23–29. https://doi.org/10.4269/ajtmh.2002.66.23.
- 116. Aziz A, Zhang W, Li J, Loukas A, McManus DP, Mulvenna J. 2011. Proteomic characterisation of *Echinococcus granulosus* hydatid cyst fluid from sheep, cattle and humans. J Proteomics 74:1560–1572. https://doi.org/10.1016/j.jprot.2011.02.021.
- 117. Santos GB, Monteiro KM, da Silva ED, Battistella ME, Ferreira HB, Zaha A. 2016. Excretory/secretory products in the *Echinococcus granulosus* metacestode: is the intermediate host complacent with infection caused by the larval form of the parasite? Int J Parasitol 46:843–856. https://doi.org/10.1016/j.ijpara.2016.07.009.
- 118. Valot B, Rognon B, Prenel A, Baraquin A, Knapp J, Anelli M, Richou C, Bresson-Hadni S, Grenouillet F, Wang J, Vuitton DA, Gottstein B, Millon L. 2017. Screening of antigenic vesicular fluid proteins of *Echinococcus multilocularis* as potential viability biomarkers to monitor drug response in alveolar echinococcosis patients. Proteomics Clin Appl 11: 11–12. https://doi.org/10.1002/prca.201700010.
- 119. Wang Y, Xiao D, Shen Y, Han X, Zhao F, Li X, Wu W, Zhou H, Zhang J, Cao J. 2015. Proteomic analysis of the excretory/secretory products and antigenic proteins of *Echinococcus granulosus* adult worms from infected dogs. BMC Vet Res 11:119. https://doi.org/10.1186/s12917-015-0423-8.
- Hidalgo C, Garcia MP, Stoore C, Ramirez JP, Monteiro KM, Hellman U, Zaha A, Ferreira HB, Galanti N, Landerer E, Paredes R. 2016. Proteomics analysis of *Echinococcus granulosus* protoscolex stage. Vet Parasitol 218:43–45. https://doi.org/10.1016/j.vetpar.2015.12.026.
- Virginio VG, Monteiro KM, Drumond F, de Carvalho MO, Vargas DM, Zaha A, Ferreira HB. 2012. Excretory/secretory products from in vitrocultured *Echinococcus granulosus* protoscoleces. Mol Biochem Parasitol 183:15–22. https://doi.org/10.1016/j.molbiopara.2012.01.001.
- 122. Kouguchi H, Matsumoto J, Katoh Y, Suzuki T, Oku Y, Yagi K. 2010. Echinococcus multilocularis: two-dimensional Western blotting method for the identification and expression analysis of immunogenic proteins in infected dogs. Exp Parasitol 124:238–243. https://doi.org/10.1016/j.exppara.2009.09.016.
- 123. Ahn CS, Han X, Bae YA, Ma X, Kim JT, Cai H, Yang HJ, Kang I, Wang H, Kong Y. 2015. Alteration of immunoproteome profile of *Echinococcus granulosus* hydatid fluid with progression of cystic echinococcosis. Parasit Vectors 8:10. https://doi.org/10.1186/s13071-014-0610-7.
- 124. Zeghir-Bouteldja R, Polome A, Bousbata S, Touil-Boukoffa C. 2017. Comparative proteome profiling of hydatid fluid from Algerian patients reveals cyst location-related variation in *Echinococcus granulosus*. Acta Trop 171:199–206. https://doi.org/10.1016/j.actatropica.2017.03.034.
- 125. Knapp J, Umhang G, Poulle ML, Millon L. 2016. Development of a real-time PCR for a sensitive one-step coprodiagnosis allowing both the identification of carnivore feces and the detection of *Toxocara* spp. and *Echinococcus multilocularis*. Appl Environ Microbiol 82:2950–2958. https://doi.org/10.1128/AEM.03467-15.
- 126. Knapp J, Millon L, Mouzon L, Umhang G, Raoul F, Ali ZS, Combes B, Comte S, Gbaguidi-Haore H, Grenouillet F, Giraudoux P. 2014. Real time PCR to detect the environmental faecal contamination by *Echinococcus multilocularis* from red fox stools. Vet Parasitol 201:40–47. https://doi.org/10.1016/j.vetpar.2013.12.023.
- Maas M, van Roon A, Dam-Deisz C, Opsteegh M, Massolo A, Deksne G, Teunis P, van der Giessen J. 2016. Evaluation by latent class analysis of

a magnetic capture based DNA extraction followed by real-time qPCR as a new diagnostic method for detection of *Echinococcus multilocularis* in definitive hosts. Vet Parasitol 230:20–24. https://doi.org/10.1016/j.vetpar.2016.10.016.

- Dinkel A, Kern S, Brinker A, Oehme R, Vaniscotte A, Giraudoux P, Mackenstedt U, Romig T. 2011. A real-time multiplex-nested PCR system for coprological diagnosis of *Echinococcus multilocularis* and host species. Parasitol Res 109:493–498. https://doi.org/10.1007/s00436-011-22772-0
- 129. Boufana BS, Campos-Ponce M, Naidich A, Buishi I, Lahmar S, Zeyhle E, Jenkins DJ, Combes B, Wen H, Xiao N, Nakao M, Ito A, Qiu J, Craig PS. 2008. Evaluation of three PCR assays for the identification of the sheep strain (genotype 1) of *Echinococcus granulosus* in canid feces and parasite tissues. Am J Trop Med Hyg 78:777–783. https://doi.org/10.4269/ajtmh.2008.78.777.
- 130. Boufana B, Umhang G, Qiu J, Chen X, Lahmar S, Boue F, Jenkins D, Craig P. 2013. Development of three PCR assays for the differentiation between *Echinococcus shiquicus*, *E. granulosus* (G1 genotype), and *E. multilocularis* DNA in the co-endemic region of Qinghai-Tibet plateau, China. Am J Trop Med Hyg 88:795–802. https://doi.org/10.4269/ajtmh.12-0331.
- 131. Grenouillet F, Umhang G, Arbez-Gindre F, Mantion G, Delabrousse E, Millon L, Boué F. 2014. *Echinococcus ortleppi* infections in humans and cattle, France. Emerg Infect Dis 20:2100–2102. https://doi.org/10.3201/eid2012.140641.
- 132. Klein C, Massolo A. 2015. Demonstration that a case of human alveolar echinococcosis in Minnesota in 1977 was caused by the N2 strain. Am J Trop Med Hyg 92:477–478. https://doi.org/10.4269/ajtmh.14-0484.
- 133. Yamano K, Kouguchi H, Uraguchi K, Mukai T, Shibata C, Yamamoto H, Takaesu N, Ito M, Makino Y, Takiguchi M, Yagi K. 2014. First detection of *Echinococcus multilocularis* infection in two species of nonhuman primates raised in a zoo: a fatal case in Cercopithecus diana and a strongly suspected case of spontaneous recovery in Macaca nigra. Parasitol Int 63:621–626. https://doi.org/10.1016/j.parint.2014.04.006.
- 134. Wassermann M, Mackenstedt U, Romig T. 2014. A loop-mediated isothermal amplification (LAMP) method for the identification of species within the *Echinococcus granulosus* complex. Vet Parasitol 200:97–103. https://doi.org/10.1016/j.vetpar.2013.12.012.
- 135. Ni XW, McManus DP, Lou ZZ, Yang JF, Yan HB, Li L, Li HM, Liu QY, Li CH, Shi WG, Fan YL, Liu X, Cai JZ, Lei MT, Fu BQ, Yang YR, Jia WZ. 2014. A comparison of loop-mediated isothermal amplification (LAMP) with other surveillance tools for *Echinococcus granulosus* diagnosis in canine definitive hosts. PLoS One 9:e100877. https://doi.org/10.1371/journal.pone.0100877.
- 136. Ni X, McManus DP, Yan H, Yang J, Lou Z, Li H, Li L, Lei M, Cai J, Fan Y, Li C, Liu Q, Shi W, Liu X, Zheng Y, Fu B, Yang Y, Jia W. 2014. Loop-mediated isothermal amplification (LAMP) assay for the identification of *Echinococcus multilocularis* infections in canine definitive hosts. Parasit Vectors 7:254. https://doi.org/10.1186/1756-3305-7-254.
- 137. Salant H, Abbasi I, Hamburger J. 2012. The development of a loop-mediated isothermal amplification method (LAMP) for *Echinococcus* granulosis coprodetection. Am J Trop Med Hyg 87:883–887. https://doi.org/10.4269/ajtmh.2012.12-0184.
- 138. Gurler AT, Gori F, Bolukbas CS, Umur S, Acici M, Deplazes P. 2018. Investigation of *Echinococcus multilocularis* in environmental definitive host feces in the Asian and the European parts of Turkey. Front Vet Sci 5:48. https://doi.org/10.3389/fvets.2018.00048.
- 139. Knapp J, Giraudoux P, Combes B, Umhang G, Boué F, Said-Ali Z, Aknouche S, Garcia C, Vacheyrou M, Laboissière A, Raton V, Comte S, Favier S, Demerson JM, Caillot C, Millon L, Raoul F. 2018. Rural and urban distribution of wild and domestic carnivore stools in the context of *Echinococcus multilocularis* environmental exposure. Int J Parasitol 48:937–946. https://doi.org/10.1016/j.ijpara.2018.05.007.
- 140. Federer K, Armua-Fernandez MT, Gori F, Hoby S, Wenker C, Deplazes P. 2016. Detection of taeniid (*Taenia* spp., *Echinococcus* spp.) eggs contaminating vegetables and fruits sold in European markets and the risk for metacestode infections in captive primates. Int J Parasitol Parasites Wildl 5:249–253. https://doi.org/10.1016/j.ijppaw.2016.07.002.
- 141. Hidalgo A, Melo A, Romero F, Hidalgo V, Villanueva J, Fonseca-Salamanca F. 2018. DNA extraction in *Echinococcus granulosus* and *Taenia* spp. eggs in dogs stool samples applying thermal shock. Exp Parasitol 186:10–16. https://doi.org/10.1016/j.exppara.2018.01.016.
- 142. Hegglin D, Bontadina F, Deplazes P. 2015. Human-wildlife interactions

- and zoonotic transmission of *Echinococcus multilocularis*. Trends Parasitol 31:167–173. https://doi.org/10.1016/j.pt.2014.12.004.
- 143. Cooney RM, Flanagan KP, Zehyle E. 2004. Review of surgical management of cystic hydatid disease in a resource limited setting: Turkana, Kenya. Eur J Gastroenterol Hepatol 16:1233–1236. https://doi.org/10.1097/00042737-200411000-00024.
- 144. Moazeni M, Alipour-Chaharmahali MR. 2011. *Echinococcus granulosus*: in vitro effectiveness of warm water on protoscolices. Exp Parasitol 127:14–17. https://doi.org/10.1016/j.exppara.2010.06.021.
- Aggarwal AR, Garg RL. 1983. Formalin toxicity in hydatid liver disease.
 Anaesthesia 38:662–665. https://doi.org/10.1111/j.1365-2044.1983.tb12159.x.
- Besim H, Karayalcin K, Hamamci O, Gungor C, Korkmaz A. 1998. Scolicidal agents in hydatid cyst surgery. HPB Surg 10:347–351. https://doi .org/10.1155/1998/78170.
- 147. Paksoy Y, Odev K, Sahin M, Arslan A, Koc O. 2005. Percutaneous treatment of liver hydatid cysts: comparison of direct injection of albendazole and hypertonic saline solution. Am J Roentgenol 185: 727–734. https://doi.org/10.2214/ajr.185.3.01850727.
- 148. Filice C, Pirola F, Brunetti E, Dughetti S, Strosselli M, Foglieni CS. 1990. A new therapeutic approach for hydatid liver cysts. Aspiration and alcohol injection under sonographic guidance. Gastroenterology 98: 1366–1368. https://doi.org/10.1016/0016-5085(90)90358-8.
- 149. Ozçelik C, Inci I, Toprak M, Eren N, Ozgen G, Yaşar T. 1994. Surgical treatment of pulmonary hydatidosis in children: experience in 92 patients. J Pediatr Surg 29:392–395. https://doi.org/10.1016/0022-3468(94)90575-4.
- Langer JC, Rose DB, Keystone JS, Taylor BR, Langer B. 1984. Diagnosis and management of hydatid disease of the liver. A 15-year North American experience. Ann Surg 199:412–417. https://doi.org/10.1097/ 00000658-198404000-00007.
- Shi H, Lei Y, Wang B, Wang Z, Xing G, Lv H, Jiang Y. 2016. Protoscolicidal effects of chenodeoxycholic acid on protoscoleces of *Echinococcus* granulosus. Exp Parasitol 167:76–82. https://doi.org/10.1016/j.exppara .2016.05.004.
- 152. Colebrook AL, Jenkins DJ, Jones MK, Tatarczuch L, Lightowlers MW. 2004. Effect of cyclosporin A on the survival and ultrastructure of *Echinococcus granulosus* protoscoleces in vitro. Parasitology 129: 497–504. https://doi.org/10.1017/S0031182004005773.
- 153. Rostami A, Mozafari M, Gholipourmalekabadi M, Caicedo HH, Lasjerdi Z, Sameni M, Samadikuchaksaraei A. 2015. Optimization of fluoride-containing bioactive glasses as a novel scolicidal agent adjunct to hydatid surgery. Acta Trop 148:105–114. https://doi.org/10.1016/j.actatropica.2015.04.021.
- Djilali G, Mahrour A, Oussedik T, Abad M, Bouguerra T, Nekrouf G, Belkaid M, Souilamas F. 1983. Hydrogen peroxide in the surgery of hydatid cyst. Presse Med 12:235–237.
- 155. Ciftci IH, Esme H, Sahin DA, Solak O, Sezer M, Dilek ON. 2007. Effect of octenidine dihydrochloride on viability of protoscoleces in hepatic and pulmonary hydatid diseases. J Natl Med Assoc 99:674–677.
- 156. Lv H, Li S, Zhang J, Liang W, Mu X, Jiang Y. 2013. In vitro effects of SB202190 on *Echinococcus granulosus*. Korean J Parasitol 51:255–258. https://doi.org/10.3347/kjp.2013.51.2.255.
- Saidi F. 1977. A new approach to the surgical treatment of hydatid cyst.
 Ann R Coll Surg Engl 59:115–118.
- Rahimi MT, Ahmadpour E, Rahimi Esboei B, Spotin A, Kohansal Koshki MH, Alizadeh A, Honary S, Barabadi H, Ali Mohammadi M. 2015. Scolicidal activity of biosynthesized silver nanoparticles against *Echinococcus granulosus* protoscolices. Int J Surg 19:128–133. https://doi.org/10 .1016/j.ijsu.2015.05.043.
- Morris DL, Richards KS, Chinnery JB. 1986. Protoscolicidal effect of praziquantel—in-vitro and electron microscopical studies on *Echino-coccus granulosus*. J Antimicrob Chemother 18:687–691. https://doi.org/10.1093/jac/18.6.687.
- Aydin I, Teksoz S, Aytac E, Polat E, Ramazanoglu R, Ozcan M. 2012.
 Scolicidal activity of taurolidine for the treatment of hydatid disease.
 Bratisl Lek Listy 113:648–651.
- Elissondo MC, Pensel PE, Denegri GM. 2013. Could thymol have effectiveness on scolices and germinal layer of hydatid cysts? Acta Trop 125:251–257. https://doi.org/10.1016/j.actatropica.2012.12.007.
- 162. Rahimi-Esboei B, Ebrahimzadeh MA, Fathi H, Rezaei Anzahaei F. 2016. Scolicidal effect of Allium sativum flowers on hydatid cyst protoscolices. Eur Rev Med Pharmacol Sci 20:129–132.
- 163. Rouhani S, Salehi N, Kamalinejad M, Zayeri F. 2013. Efficacy of Berberis

vulgaris aqueous extract on viability of *Echinococcus granulosus* protoscolices. J Invest Surg 26:347–351. https://doi.org/10.3109/08941939.2013.818746.

- 164. Lashkarizadeh MR, Asgaripour K, Saedi Dezaki E, Fasihi Harandi M. 2015. Comparison of scolicidal effects of amphotricin B, silver nanoparticles, and Foeniculum vulgare mill on hydatid cysts protoscoleces. Iran J Parasitol 10:206–212.
- Mahmoudvand H, Fallahi S, Mahmoudvand H, Shakibaie M, Harandi MF, Dezaki ES. 2016. Efficacy of *Myrtus communis* L. to inactivate the hydatid cyst protoscoleces. J InvestSurg 29:137–143. https://doi.org/10 .3109/08941939.2015.1088601.
- 166. Mahmoudvand H, Dezaki ES, Kheirandish F, Ezatpour B, Jahanbakhsh S, Harandi MF. 2014. Scolicidal effects of black cumin seed (*Nigella sativa*) essential oil on hydatid cysts. Korean J Parasitol 52:653–659. https://doi.org/10.3347/kjp.2014.52.6.653.
- 167. Mahmoudvand H, Kheirandish F, Dezaki ES, Shamsaddini S, Harandi MF. 2016. Chemical composition, efficacy and safety of *Pistacia vera* (var. Fandoghi) to inactivate protoscoleces during hydatid cyst surgery. Biomed Pharmacother 82:393–398. https://doi.org/10.1016/j.biopha 2016.05.012
- 168. Mahmoudvand H, Mirbadie SR, Ghasemi Kia M, Badparva E, Shamsadini Lori S, Fasihi Harandi M. 2016. Efficacy of *Pistacia khinjuk* fruits on viability of hydatid cyst protoscoleces and its acute toxicity in mice model. Iran J Parasitol 11:383–388.
- Zibaei M, Rostamipour R, Nayebzadeh H. 2016. Effect of *Pistacia atlantica* fruit and leaf extracts on hydatid cyst protoscolices. Recent Pat Antiinfect Drug Discov 11:53–58. https://doi.org/10.2174/1574891X 10666151029113334.
- 170. Zhao J, Yang W, Ma X. 2004. Experimental study on combination chemotherapy of *Echinococcus granulosus* in mice with *Peganum harmala* L, harmine and albendazole. J Xinjiang Med Univ 24:125–127.
- 171. Abdel-Baki AA, Almalki E, Mansour L, Al-Quarishy S. 2016. In vitro scolicidal effects of *Salvadora persica* root extract against protoscolices of *Echinococcus granulosus*. Korean J Parasitol 54:61–66. https://doi.org/10.3347/kjp.2016.54.1.61.
- 172. Jahanbakhsh S, Azadpour M, Tavakoli Kareshk A, Keyhani A, Mahmoudvand H. 2016. *Zataria multiflora* Bioss: lethal effects of methanolic extract against protoscoleces of *Echinococcus granulosus*. J Parasit Dis 40:1289–1292. https://doi.org/10.1007/s12639-015-0670-4.
- 173. Kilicoglu B, Kismet K, Koru O, Tanyuksel M, Oruc MT, Sorkun K, Akkus MA. 2006. The scolicidal effects of honey. Adv Ther 23:1077–1083. https://doi.org/10.1007/BF02850228.
- 174. Kismet K, Kilicoglu B, Koru O, Tanyuksel M, Oruc MT, Sorkun K, Salih B, Akkus MA. 2006. Evaluation on scolicidal efficacy of propolis. Eur Surg Res 38:476–481. https://doi.org/10.1159/000096006.
- 175. Zhang R, Aji T, Shao Y, Jiang T, Yang L, Lv W, Chen Y, Chen X, Wen H. 2017. Nanosecond pulsed electric field (nsPEF) disrupts the structure and metabolism of human *Echinococcus granulosus* protoscolex *in vitro* with a dose effect. Parasitol Res 116:1345–1351. https://doi.org/10.1007/s00436-017-5412-3.
- 176. Lamonaca V, Virga A, Minervini MI, Di Stefano R, Provenzani A, Tagliareni P, Fleres G, Luca A, Vizzini G, Palazzo U, Gridelli B. 2009. Cystic echinococcosis of the liver and lung treated by radiofrequency thermal ablation: an ex-vivo pilot experimental study in animal models. World J Gastroenterol 15:3232–3239. https://doi.org/10.3748/wjg.15.3232.
- 177. Siles-Lucas M, Casulli A, Cirilli R, Carmena D. 2018. Progress in the pharmacological treatment of human cystic and alveolar echinococcosis: compounds and therapeutic targets. PLoS Negl Trop Dis 12:e0006422. https://doi.org/10.1371/journal.pntd.0006422.
- 178. Vuitton DA, Azizi A, Richou C, Vuitton L, Blagosklonov O, Delabrousse E, Mantion GA, Bresson-Hadni S. 2016. Current interventional strategy for the treatment of hepatic alveolar echinococcosis. Expert Rev anti Infect Ther 14:1179–1194. https://doi.org/10.1080/14787210.2016.1240030.
- Davis A, Dixon H, Pawlowski ZS. 1989. Multicentre clinical trials of benzimidazole-carbamates in human cystic echinococcosis (phase 2). Bull World Health Organ 67:503–508.
- Ammann R, Tschudi K, Ziegler M, Meister F, Cotting J, Eckert J, Witassek F, Freiburghaus A. 1988. The long-term course of 60 patients with alveolar echinococcosis in continuous therapy with mebendazole (1976-85)]. Klin Wochenschr 66:1060–1073. https://doi.org/10.1007/BF01711918.
- 181. Horton J. 2003. Albendazole for the treatment of echinococcosis. Fundam Clin Pharmacol 17:205–212.
- 182. Vuitton D, Bresson-Hadni S. 2014. Alveolar echinococcosis: evaluation

- of therapeutic strategies. Expert Opin Orphan Drugs 2:67–86. https://doi.org/10.1517/21678707.2014.870033.
- 183. Nazligul Y, Kucukazman M, Akbulut S. 2015. Role of chemotherapeutic agents in the management of cystic echinococcosis. Int Surg 100: 112–114. https://doi.org/10.9738/INTSURG-D-14-00068.1.
- 184. Li T, Ito A, Pengcuo R, Sako Y, Chen X, Qiu D, Xiao N, Craig PS. 2011. Post-treatment follow-up study of abdominal cystic echinococcosis in Tibetan communities of northwest Sichuan Province, China. PLoS Negl Trop Dis 5:e1364. https://doi.org/10.1371/journal.pntd.0001364.
- 185. Gupta N, Javed A, Puri S, Jain S, Singh S, Agarwal AK. 2011. Hepatic hydatid: PAIR, drain or resect? J Gastrointest Surg 15:1829–1836. https://doi.org/10.1007/s11605-011-1649-9.
- Tagliacozzo S, Miccini M, Amore Bonapasta S, Gregori M, Tocchi A.
 Surgical treatment of hydatid disease of the liver: 25 years of experience. Am J Surg 201:797–804. https://doi.org/10.1016/j.amjsurg.2010.02.011.
- 187. Katkhouda N, Fabiani P, Benizri E, Mouiel J. 1992. Laser resection of a liver hydatid cyst under videolaparoscopy. Br J Surg 79:560–561. https://doi.org/10.1002/bjs.1800790628.
- 188. Casciola L, Patriti A, Ceccarelli G, Bartoli A, Ceribelli C, Spaziani A. 2011. Robot-assisted parenchymal-sparing liver surgery including lesions located in the posterosuperior segments. Surg Endosc 25:3815–3824. https://doi.org/10.1007/s00464-011-1796-9.
- Soni HN, Nagpal AP, Zumkhawala BR, Haribhakti SP. 2011. Single-incision laparoscopic percutaneous hydatid cystectomy: a novel curative approach for segment 7 hydatid cyst of liver. Surg Laparosc Endosc Percutan Tech 21:e253. https://doi.org/10.1097/SLE.0b013e318230f732.
- Chen W, Xusheng L. 2007. Laparoscopic surgical techniques in patients with hepatic hydatid cyst. Am J Surg 194:243–247. https://doi.org/10 .1016/j.amjsurg.2006.11.033.
- Tuxun T, Zhang JH, Zhao JM, Tai QW, Abudurexti M, Ma HZ, Wen H. 2014. World review of laparoscopic treatment of liver cystic echinococcosis—914 patients. Int J Infect Dis 24C:43–50. https://doi.org/10.1016/j.ijid.2014.01.012.
- 192. Tamarozzi F, Vuitton L, Brunetti E, Vuitton DA, Koch S. 2014. Nonsurgical and non-chemical attempts to treat echinococcosis: do they work? Parasite 21:75. https://doi.org/10.1051/parasite/2014071.
- Akaydin M, Erozgen F, Ersoy YE, Birol S, Kaplan R. 2012. Treatment of hepatic hydatid disease complications using endoscopic retrograde cholangiopancreatography procedures. Can J Surg 55:244–248. https:// doi.org/10.1503/cjs.036010.
- 194. Sakçak I, Eriş C, Ölmez A, Kayaalp C, Yılmaz S. 2012. Replacement of the vena cava with aortic graft for living donor liver transplantation in Budd-Chiari syndrome associated with hydatid cyst surgery: a case report. Transplant Proc 44:1757–1758. https://doi.org/10.1016/j.transproceed .2012.04.023.
- Akhan O, Gumus B, Akinci D, Karcaaltincaba M, Ozmen M. 2007. Diagnosis and percutaneous treatment of soft-tissue hydatid cysts. Cardiovasc Intervent Radiol 30:419–425. https://doi.org/10.1007/s00270-006-0153-1.
- Ozdemir F, Ince V, Barut B, Onur A, Kayaalp C, Yilmaz S. 2015. Living donor liver transplantation for *Echinococcus* Alveolaris: single-center experience. Liver Transpl 21:1091–1095. https://doi.org/10.1002/lt .24170.
- Mantion GA, Vuitton DA. 2011. Auto-versus allo-transplantation of the liver for end-stage alveolar echinococcosis? Chin Med J (Engl) 124: 2803–2805.
- 198. Ringe B, Pichlmayr R, Burdelski M. 1988. A new technique of hepatic vein reconstruction in partial liver transplantation. Transplant Int 1:30–35. https://doi.org/10.1111/j.1432-2277.1988.tb01776.x.
- 199. Pichlmayr R, Bretschneider HJ, Kirchner E, Ringe B, Lamesch P, Gubernatis G, Hauss J, Niehaus KJ, Kaukemuller J. 1988. Ex situ operation on the liver. A new possibility in liver surgery. Langenbecks Arch Chir 373:122–126. https://doi.org/10.1007/BF01262775.
- 200. Wen H, Dong JH, Zhang JH, Zhao JM, Shao YM, Duan WD, Liang YR, Ji XW, Tai QW, Aji T, Li T. 2011. Ex vivo liver resection followed by autotransplantation for end-stage hepatic alveolar echinococcosis. Chin Med J (Engl) 124:2813–2817.
- 201. Jianyong L, Jingcheng H, Wentao W, Lunan Y, Jichun Z, Bing H, Ding Y. 2015. Ex vivo liver resection followed by autotransplantation to a patient with advanced alveolar echinococcosis with a replacement of the retrohepatic inferior vena cava using autogenous vein grafting: a case report and literature review. Medicine (Baltimore, MD) 94:e514. https://doi.org/10.1097/MD.000000000000514.
- 202. Wen H, Dong JH, Zhang JH, Duan WD, Zhao JM, Liang YR, Shao YM,

- Ji XW, Tai QW, Li T, Gu H, Tuxun T, He YB, Huang JF. 2016. Ex vivo liver resection and autotransplantation for end-stage alveolar echinococcosis: a case series. Am J Transplant 16:615–624. https://doi.org/10.1111/ajt.13465.
- 203. Aji T, Dong JH, Shao YM, Zhao JM, Li T, Shalayiadang P, Ran B, Zhang TM, Zhang RQ, Tuxun T, He YB, Huang JF, Wen H. 2017. Autologous liver transplantation in end-stage hepatic alveolar echinoccossis with 51 patients' experience—where do we stand? J Hepatol 66:S112. https://doi.org/10.1016/S0168-8278(17)30484-1.
- 204. Aji T, Dong JH, Shao YM, Zhao JM, Li T, Tuxun T, Shalayiadang P, Ran B, Jiang TM, Zhang RQ, He YB, Huang JF, Wen H. 2018. Ex vivo liver resection and autotransplantation as alternative to allotransplantation for end-stage hepatic alveolar echinococcosis. J Hepatol 69:1037–1046. https://doi.org/10.1016/j.jhep.2018.07.006.
- 205. Buttenschoen K, Carli Buttenschoen D, Gruener B, Kern P, Beger HG, Henne-Bruns D, Reuter S. 2009. Long-term experience on surgical treatment of alveolar echinococcosis. Langenbecks Arch Surg 394: 689–698. https://doi.org/10.1007/s00423-008-0392-5.
- 206. Kadry Z, Renner EC, Bachmann LM, Attigah N, Renner EL, Ammann RW, Clavien PA. 2005. Evaluation of treatment and long-term follow-up in patients with hepatic alveolar echinococcosis. Br J Surg 92:1110–1116. https://doi.org/10.1002/bjs.4998.
- Ayifuhan A, Tuerganaili A, Jun C, Ying-Mei S, Xiang-Wei L, Hao W. 2012.
 Surgical treatment for hepatic alveolar echinococcosis: report of 50 cases. Hepatogastroenterology 59:790–793. https://doi.org/10.5754/hge10545.
- 208. Hillenbrand A, Gruener B, Kratzer W, Kern P, Graeter T, Barth TF, Buttenschoen K, Henne-Bruns D. 2017. Impact of safe distance on long-term outcome after surgical therapy of alveolar echinococcosis. World J Surg 41:1012–1018. https://doi.org/10.1007/s00268-016-3813-6.
- Graeter T, Ehing F, Oeztuerk S, Mason RA, Haenle MM, Kratzer W, Seufferlein T, Gruener B. 2015. Hepatobiliary complications of alveolar echinococcosis: a long-term follow-up study. World J Gastroenterol 21:4925–4932. https://doi.org/10.3748/wjg.v21.i16.4925.
- 210. Frei P, Misselwitz B, Prakash MK, Schoepfer AM, Prinz Vavricka BM, Mullhaupt B, Fried M, Lehmann K, Ammann RW, Vavricka SR. 2014. Late biliary complications in human alveolar echinococcosis are associated with high mortality. World J Gastroenterol 20:5881–5888. https://doi.org/10.3748/wjg.v20.i19.5881.
- 211. Ambregna S, Koch S, Sulz MC, Gruner B, Ozturk S, Chevaux JB, Sulima M, de Gottardi A, Napoleon B, Abergel A, Bichard P, Boytchev I, Deprez P, Dumortier J, Frossard JL, Kull E, Meny B, Moradpour D, Prat F, Vanbiervliet G, Thevenot T, Vuitton DA, Bresson-Hadni S, Vuitton L. 2017. A European survey of perendoscopic treatment of biliary complications in patients with alveolar echinococcosis. Expert Rev Anti Infect Ther 15:79–88. https://doi.org/10.1080/14787210.2017.1252260.
- 212. Torgerson PR, Schweiger A, Deplazes P, Pohar M, Reichen J, Ammann RW, Tarr PE, Halkik N, Mullhaupt B. 2008. Alveolar echinococcosis: from a deadly disease to a well-controlled infection. Relative survival and economic analysis in Switzerland over the last 35 years. J Hepatol 49:72–77. https://doi.org/10.1016/j.jhep.2008.03.023.
- Hemphill A, Stadelmann B, Rufener R, Spiliotis M, Boubaker G, Muller J, Muller N, Gorgas D, Gottstein B. 2014. Treatment of echinococcosis: albendazole and mebendazole–what else? Parasite 21:70. https://doi.org/10.1051/parasite/2014073.
- 214. Rufener R, Ritler D, Zielinski J, Dick L, da Silva ET, da Silva Araujo A, Joekel DE, Czock D, Goepfert C, Moraes AM, de Souza MVN, Müller J, Mevissen M, Hemphill A, Lundström-Stadelmann B. 2018. Activity of mefloquine and mefloquine derivatives against *Echinococcus multilocularis*. Int J Parasitol Drugs Drug Resist 8:331–340. https://doi.org/10.1016/j.ijpddr.2018.06.004.
- 215. Liu C, Yin J, Xue J, Tao Y, Hu W, Zhang H. 2018. In vitro effects of amino alcohols on *Echinococcus granulosus*. Acta Trop 182:285–290. https://doi.org/10.1016/j.actatropica.2017.08.031.
- Liu C, Zhang H, Yin J, Hu W. 2015. In vivo and in vitro efficacies of mebendazole, mefloquine and nitazoxanide against cyst echinococcosis. Parasitol Res 114:2213–2222. https://doi.org/10.1007/s00436-015 -4412-4
- 217. Kuster T, Stadelmann B, Rufener R, Risch C, Muller J, Hemphill A. 2015. Oral treatments of *Echinococcus multilocularis*-infected mice with the antimalarial drug mefloquine that potentially interacts with parasite ferritin and cystatin. Int J Antimicrob Agents 46:546–551. https://doi.org/10.1016/j.ijantimicag.2015.07.016.
- 218. Stadelmann B, Kuster T, Scholl S, Barna F, Kropf C, Keiser J, Boykin DW,

- Stephens CE, Hemphill A. 2011. In vitro efficacy of dicationic compounds and mefloquine enantiomers against *Echinococcus multilocularis* metacestodes. Antimicrob Agents Chemother 55:4866–4872. https://doi.org/10.1128/AAC.00478-11.
- 219. Lü G, Zhang W, Wang J, Xiao Y, Zhao J, Zhao J, Sun Y, Zhang C, Wang J, Lin R, Liu H, Zhang F, Wen H. 2014. Application of a cDNA microarray for profiling the gene expression of *Echinococcus granulosus* protoscoleces treated with albendazole and artemisinin. Mol Biochem Parasitol 198:59–65. https://doi.org/10.1016/j.molbiopara.2014.12.002.
- 220. Spicher M, Roethlisberger C, Lany C, Stadelmann B, Keiser J, Ortega-Mora LM, Gottstein B, Hemphill A. 2008. In vitro and in vivo treatment of *Echinococcus* protoscoleces and metacestodes with artemisinin and artemisinin-derivatives. Antimicrob Agents Chemother 52:3447–3450. https://doi.org/10.1128/AAC.00553-08.
- 221. Wang W, Li J, Yao J, Wang T, Li S, Zheng X, Duan L, Zhang W. 2017. In vitro and in vivo efficacies of novel carbazole aminoalcohols in the treatment of cystic echinococcosis. J Antimicrob Chemother 72: 3122–3130. https://doi.org/10.1093/jac/dkx250.
- 222. WHO Informal Working Group on Echinococcosis. 1996. Guidelines for treatment of cystic and alveolar echinococcosis in humans. Bull World Health Organ 74:231–242.
- Gottstein B, Wang J, Blagosklonov O, Grenouillet F, Millon L, Vuitton DA, Muller N. 2014. *Echinococcus* metacestode: in search of viability markers. Parasite 21:63. https://doi.org/10.1051/parasite/2014063.
- 224. Bresson-Hadni S, Blagosklonov O, Knapp J, Grenouillet F, Sako Y, Delabrousse E, Brientini MP, Richou C, Minello A, Antonino AT, Gillet M, Ito A, Mantion GA, Vuitton DA. 2011. Should possible recurrence of disease contraindicate liver transplantation in patients with end-stage alveolar echinococcosis? A 20-year follow-up study. Liver Transpl 17:855–865. https://doi.org/10.1002/lt.22299.
- 225. Caoduro C, Porot C, Vuitton DA, Bresson-Hadni S, Grenouillet F, Richou C, Boulahdour H, Blagosklonov O. 2013. The role of delayed 18F-FDG PET imaging in the follow-up of patients with alveolar echinococcosis. J Nucl Med 54:358–363. https://doi.org/10.2967/jnumed.112.109942.
- Matsumoto J, Muller N, Hemphill A, Oku Y, Kamiya M, Gottstein B. 2006.
 14-3-3- and Il/3-10-gene expression as molecular markers to address viability and growth activity of *Echinococcus multilocularis* metacestodes.
 Parasitology 132:83–94. https://doi.org/10.1017/S0031182005008632.
- 227. Li T, Ito A, Chen X, Sako Y, Qiu J, Xiao N, Qiu D, Nakao M, Yanagida T, Craig PS. 2010. Specific IgG responses to recombinant antigen B and em18 in cystic and alveolar echinococcosis in china. Clin Vaccine Immunol 17:470–475. https://doi.org/10.1128/CVI.00466-09.
- 228. Ammann RW, Stumpe KD, Grimm F, Deplazes P, Huber S, Bertogg K, Fischer DR, Mullhaupt B. 2015. Outcome after discontinuing long-term benzimidazole treatment in 11 patients with non-resectable alveolar echinococcosis with negative FDG-PET/CT and anti-Emll/3-10 serology. PLoS Negl Trop Dis 9:e0003964. https://doi.org/10.1371/journal.pntd.0003964.
- 229. Sako Y, Tappe D, Fukuda K, Kobayashi Y, Itoh S, Frosch M, Gruner B, Kern P, Ito A. 2011. Immunochromatographic test with recombinant Em18 antigen for the follow-up study of alveolar echinococcosis. Clin Vaccine Immunol 18:1302–1305. https://doi.org/10.1128/CVI.05156-11.
- 230. Ferragut G, Ljungstrom I, Nieto A. 1998. Relevance of circulating antigen detection to follow-up experimental and human cystic hydatid infections. Parasite Immunol 20:541–549. https://doi.org/10.1046/j.1365-3024.1998.00177.x.
- 231. Bauomi IR, El-Amir AM, Fahmy AM, Zalat RS, Diab TM. 2015. Evaluation of purified 27.5 kDa protoscolex antigen-based ELISA for the detection of circulating antigens and antibodies in sheep and human hydatidosis. J Helminthol 89:577–583. https://doi.org/10.1017/S0022149X14000479.
- 232. Weerakoon KG, McManus DP. 2016. Cell-free DNA as a diagnostic tool for human parasitic infections. Trends Parasitol 32:378–391. https://doi.org/10.1016/j.pt.2016.01.006.
- 233. Baraquin A, Hervouet E, Richou C, Flori P, Peixoto P, Azizi A, Delabrousse E, Blagosklonov O, Umhang G, Bresson-Hadni S, Valot B, Grenouillet F, Felix S, Heyd B, Mantion G, Di Martino V, Montange D, Vanlemmens C, Vuitton DA, Weil-Verhoeven D, Chavanet P, Dalle F, Gohier S, Minello A, Piroth L, Dumortier J, Mabrut J-Y, Wallon M, Frentiu E, Machouart M, Watelet J, Chemla C, Feron T, Heurge-Berlot A, Sommacale D, Thiefin G, Abou-Bacar A, Brunet J, Candolfi E, Hansmann Y, Lefebvre N. 2018. Circulating cell-free DNA in patients with alveolar echinococcosis. Mol Biochem Parasitol 222:14–20. https://doi.org/10.1016/j.molbiopara.2018.04.004.
- 234. Cvejic D, Schneider C, Fourie J, de Vos C, Bonneau S, Bernachon N,

Hellmann K. 2016. Efficacy of a single dose of milbemycin oxime/praziquantel combination tablets, Milpro((R)), against adult *Echinococcus multilocularis* in dogs and both adult and immature *E. multilocularis* in young cats. Parasitol Res 115:1195–1202. https://doi.org/10.1007/s00436-015-4855-7.

- 235. Del Carpio M, Hugo Mercapide C, Salvitti JC, Uchiumi L, Sustercic J, Panomarenko H, Moguilensky J, Herrero E, Talmon G, Volpe M, Araya D, Mujica G, Calabro A, Mancini S, Chiosso C, Luis Labanchi J, Saad R, Goblirsch S, Brunetti E, Larrieu E. 2012. Early diagnosis, treatment and follow-up of cystic echinococcosis in remote rural areas in Patagonia: impact of ultrasound training of non-specialists. PLoS Negl Trop Dis 6:e1444. https://doi.org/10.1371/journal.pntd.0001444.
- 236. Merino V, Westgard CM, Bayer AM, Garcia PJ. 2017. Knowledge, attitudes, and practices regarding cystic echinococcosis and sheep herding in Peru: a mixed-methods approach. BMC Vet Res 13:213. https://doi.org/10.1186/s12917-017-1130-4.
- 237. Zhang W, Zhang Z, Yimit T, Shi B, Aili H, Tulson G, You H, Li J, Gray DJ, McManus DP, Wang J. 2009. A pilot study for control of hyperendemic cystic hydatid disease in China. PLoS Negl Trop Dis 3:e534. https://doi.org/10.1371/journal.pntd.0000534.
- van Kesteren F, Qi X, Tao J, Feng X, Mastin A, Craig PS, Vuitton DA, Duan X, Chu X, Zhu J, Wen H. 2015. Independent evaluation of a canine echinococcosis control programme in Hobukesar County, Xinjiang, China. Acta Trop 145:1–7. https://doi.org/10.1016/j.actatropica.2015.01.009.
- 239. Zhang W, McManus DP. 2008. Vaccination of dogs against *Echinococcus granulosus*: a means to control hydatid disease? Trends Parasitol 24: 419–424. https://doi.org/10.1016/j.pt.2008.05.008.
- 240. Petavy AF, Hormaeche C, Lahmar S, Ouhelli H, Chabalgoity A, Marchal T, Azzouz S, Schreiber F, Alvite G, Sarciron ME, Maskell D, Esteves A, Bosquet G. 2008. An oral recombinant vaccine in dogs against *Echinococcus granulosus*, the causative agent of human hydatid disease: a pilot study. PLoS Negl Trop Dis 2:e125. https://doi.org/10.1371/journal.pptd.0000125
- 241. Umhang G, Comte S, Raton V, Hormaz V, Boucher JM, Favier S, Combes B, Boue F. 2014. *Echinococcus multilocularis* infections in dogs from urban and peri-urban areas in France. Parasitol Res 113:2219–2222. https://doi.org/10.1007/s00436-014-3875-z.
- Robardet E, Giraudoux P, Caillot C, Boue F, Cliquet F, Augot D, Barrat J.
 Robardet E, Giraudoux P, Caillot C, Boue F, Cliquet F, Augot D, Barrat J.
 Robardet E, Giraudoux P, Caillot C, Boue F, Cliquet F, Augot D, Barrat J.
 Robardet E, Giraudoux P, Caillot C, Boue F, Cliquet F, Augot D, Barrat J.
 Robardet E, Giraudoux P, Caillot C, Boue F, Cliquet F, Augot D, Barrat J.
 Robardet E, Giraudoux P, Caillot C, Boue F, Cliquet F, Augot D, Barrat J.
 Robardet E, Giraudoux P, Caillot C, Boue F, Cliquet F, Augot D, Barrat J.
 Robardet E, Giraudoux P, Caillot C, Boue F, Cliquet F, Augot D, Barrat J.
 Robardet E, Giraudoux P, Caillot C, Boue F, Cliquet F, Augot D, Barrat J.
 Robardet E, Giraudoux P, Caillot C, Boue F, Cliquet F, Augot D, Barrat J.
 Robardet E, Giraudoux P, Caillot C, Boue F, Cliquet F, Augot D, Barrat J.
 Robardet E, Giraudoux P, Caillot C, Boue F, Cliquet F, Augot D, Barrat J.
 Robardet E, Giraudoux P, Caillot C, Boue F, Cliquet F, Augot D, Barrat J.
 Robardet E, Giraudoux P, Caillot C, Boue F, Cliquet F, Augot D, Barrat J.
 Robardet E, Giraudoux P, Caillot C, Boue F, Cliquet F, Augot D, Barrat J.
 Robardet E, Giraudoux P, Caillot C, Boue F, Cliquet F, Augot D, Barrat J.
 Robardet E, Giraudoux P, Caillot C, Boue F, Cliquet F, Augot D, Barrat J.
 Robardet E, Giraudoux P, Caillot C, Boue F, Cliquet F, Augot D, Barrat J.
 Robardet E, Giraudoux P, Caillot C, Boue F, Cliquet F, Augot D, Barrat J.
 Robardet E, Giraudoux P, Caillot C, Boue F, Cliquet F, Augot D, Barrat J.
 Robardet E, Giraudoux P, Caillot C, Boue F, Cliquet F, Augot D, Barrat J.
 Robardet E, Caillot C, Barrat D, Barrat J.
 Robardet E, Giraudoux P, Caillot C, Barrat D, Barr
- 243. Isaksson M, Hagstrom A, Armua-Fernandez MT, Wahlstrom H, Agren EO, Miller A, Holmberg A, Lukacs M, Casulli A, Deplazes P, Juremalm M. 2014. A semi-automated magnetic capture probe based DNA extraction and real-time PCR method applied in the Swedish surveillance of *Echinococcus multilocularis* in red fox (*Vulpes vulpes*) faecal samples. Parasit Vectors 7:583. https://doi.org/10.1186/s13071-014-0583-6.
- 244. Duscher T, Hodzic A, Glawischnig W, Duscher GG. 2017. The raccoon dog (*Nyctereutes procyonoides*) and the raccoon (*Procyon lotor*)—their role and impact of maintaining and transmitting zoonotic diseases in Austria, Central Europe. Parasitol Res 116:1411–1416. https://doi.org/10.1007/s00436-017-5405-2.
- 245. Oksanen A, Siles-Lucas M, Karamon J, Possenti A, Conraths FJ, Romig T, Wysocki P, Mannocci A, Mipatrini D, La Torre G, Boufana B, Casulli A. 2016. The geographical distribution and prevalence of *Echinococcus multilocularis* in animals in the European Union and adjacent countries: a systematic review and meta-analysis. Parasit Vectors 9:519. https://doi.org/10.1186/s13071-016-1746-4.
- 246. Mastin A, van Kesteren F, Torgerson PR, Ziadinov I, Mytynova B, Rogan MT, Tursunov T, Craig PS. 2015. Risk factors for *Echinococcus* coproantigen positivity in dogs from the Alay valley, Kyrgyzstan. J Helminthol 89:655–663. https://doi.org/10.1017/S0022149X15000590.
- 247. Romig T, Omer RA, Zeyhle E, Huttner M, Dinkel A, Siefert L, Elmahdi IE, Magambo J, Ocaido M, Menezes CN, Ahmed ME, Mbae C, Grobusch MP, Kern P. 2011. Echinococcosis in sub-Saharan Africa: emerging complexity. Vet Parasitol 181:43–47. https://doi.org/10.1016/j.vetpar.2011.04.022.
- 248. Comte S, Raton V, Raoul F, Hegglin D, Giraudoux P, Deplazes P, Favier S, Gottschek D, Umhang G, Boue F, Combes B. 2013. Fox baiting against *Echinococcus multilocularis*: contrasted achievements among two medium size cities. Prev Vet Med 111:147–155. https://doi.org/10.1016/j.prevetmed.2013.03.016.
- 249. Janko C, Konig A. 2011. Disappearance rate of praziquantel-containing

- bait around villages and small towns in southern Bavaria, Germany. J Wildl Dis 47:373–380. https://doi.org/10.7589/0090-3558-47.2.373.
- Umhang G, Lahoreau J, Hormaz V, Boucher JM, Guenon A, Montange D, Grenouillet F, Boue F. 2016. Surveillance and management of *Echino-coccus multilocularis* in a wildlife park. Parasitol Int 65:245–250. https://doi.org/10.1016/j.parint.2016.01.008.
- 251. Bružinskaitė-Schmidhalter R, Šarkūnas M, Malakauskas A, Mathis A, Torgerson PR, Deplazes P. 2012. Helminths of red foxes (*Vulpes vulpes*) and raccoon dogs (*Nyctereutes procyonoides*) in Lithuania. Parasitology 139:120–127. https://doi.org/10.1017/S0031182011001715.
- 252. Wang Q, Yu WJ, Zhong B, Shang JY, Huang L, Mastin A, Renqing-pengcuo Huang Y, Zhang GJ, He W, Giraudoux P, Wu WP, Craig PS. 2016. Seasonal pattern of *Echinococcus* re-infection in owned dogs in Tibetan communities of Sichuan, China and its implications for control. Infect Dis Poverty 5:60. https://doi.org/10.1186/s40249-016-0155-4.
- 253. Luo A, Wang H, Li JQ, Wu HS, Yang F, Fang PQ. 2014. Epidemic factors and control of hepatic echinococcosis in Qinghai province. J Huazhong Univ Sci Technolog Med Sci 34:142–145. https://doi.org/10.1007/s11596-014-1246-8.
- 254. Craig P, Mastin A, van Kesteren F, Boufana B. 2015. *Echinococcus granulosus*: epidemiology and state-of-the-art of diagnostics in animals. Vet Parasitol 213:132–148. https://doi.org/10.1016/j.vetpar.2015.07.028.
- Maurelli MP, Bosco A, Pepe P, Ianniello D, Amadesi A, Cringoli G, Rinaldi L. 2018. Innovative tools for the diagnosis of *Echinococcus granulosus* in definitive hosts. Parasitol Res 117:2607–2612. https://doi.org/10.1007/ s00436-018-5952-1.
- 256. Oge H, Oge S, Gonenc B, Sarimehmetoglu O, Ozbakis G. 2017. Coprodiagnosis of *Echinococcus granulosus* infection in dogs from Ankara, Turkey. Vet Parasitol 242:44–46. https://doi.org/10.1016/j.vetpar.2017.05.016.
- 257. Maksimov P, Schares G, Press S, Frohlich A, Basso W, Herzig M, Conraths FJ. 2017. Comparison of different commercial DNA extraction kits and PCR protocols for the detection of *Echinococcus multilocularis* eggs in faecal samples from foxes. Vet Parasitol 237:83–93. https://doi.org/10.1016/j.vetpar.2017.02.015.
- 258. Oines O, Isaksson M, Hagstrom A, Tavornpanich S, Davidson RK. 2014. Laboratory assessment of sensitive molecular tools for detection of low levels of *Echinococcus multilocularis*-eggs in fox (*Vulpes vulpes*) faeces. Parasit Vectors 7:246. https://doi.org/10.1186/1756-3305-7-246.
- 259. Karamon J. 2014. Detection of *Echinococcus multilocularis* in faeces by nested PCR with the use of diluted DNA samples. Pol J Vet Sci 17: 79–83. https://doi.org/10.2478/pjvs-2014-0010.
- Vaniscotte A, Raoul F, Poulle ML, Romig T, Dinkel A, Takahashi K, Guislain MH, Moss J, Tiaoying L, Wang Q, Qiu J, Craig PS, Giraudoux P. 2011. Role of dog behaviour and environmental fecal contamination in transmission of *Echinococcus multilocularis* in Tibetan communities. Parasitology 138: 1316–1329. https://doi.org/10.1017/S0031182011000874.
- Zheng H, Zhang W, Zhang L, Zhang Z, Li J, Lu G, Zhu Y, Wang Y, Huang Y, Liu J, Kang H, Chen J, Wang L, Chen A, Yu S, Gao Z, Jin L, Gu W, Wang Z, Zhao L, Shi B, Wen H, Lin R, Jones MK, Brejova B, Vinar T, Zhao G, McManus DP, Chen Z, Zhou Y, Wang S. 2013. The genome of the hydatid tapeworm *Echinococcus granulosus*. Nat Genet 45:1168–1175. https://doi.org/10.1038/ng.2757.
- 262. Tsai IJ, Zarowiecki M, Holroyd N, Garciarrubio A, Sánchez-Flores A, Brooks KL, Tracey A, Bobes RJ, Fragoso G, Sciutto E, Aslett M, Beasley H, Bennett HM, Cai X, Camicia F, Clark R, Cucher M, De Silva N, Day TA, Deplazes P, Estrada K, Fernández C, Holland PWH, Hou J, Hu S, Huckvale T, Hung SS, Kamenetzky L, Keane JA, Kiss F, Koziol U, Lambert O, Liu K, Luo X, Luo Y, Macchiaroli N, Nichol S, Paps J, Parkinson J, Pouchkina-Stantcheva N, Riddiford N, Rosenzvit M, Salinas G, Wasmuth JD, Zamanian M, Zheng Y, Cai J, Soberón X, Olson PD, Laclette JP, Brehm K, Berriman M. 2013. The genomes of four tapeworm species reveal adaptations to parasitism. Nature 496:57–63. https://doi.org/10.1038/nature12031.
- 263. Smyth JD. 1990. In vitro cultivation of parasitic helminths. CRC Press, Boca Raton, FL.
- 264. Smyth JD. 1982. The insemination-fertilization problem in cestodes cultured in vitro, p 393–406. In Meerovitch E (ed), Aspects of parasitology. McGill University, Montreal, Canada.
- 265. Constantine CC, Bennet-Jenkins EM, Lymbery AJ, Jenkins DJ, Behm CA, Thompson RC. 1998. Factors influencing the development and carbohydrate metabolism of *Echinococcus granulosus* in dogs. J Parasitol 84:873–881. https://doi.org/10.2307/3284612.
- 266. Russ AP, Lampel S. 2005. The druggable genome: an update. Drug

Discov Today 10:1607–1610. https://doi.org/10.1016/S1359-6446 (05)03666-4.

- 267. Lu G, Li J, Zhang C, Li L, Bi X, Li C, Fan J, Lu X, Vuitton DA, Wen H, Lin R. 2016. Molecular cloning and characterization of a P38-like mitogenactivated protein kinase from *Echinococcus granulosus*. Korean J Parasitol 54:759–768. https://doi.org/10.3347/kjp.2016.54.6.759.
- Lin R, Lu G, Wang J, Zhang C, Xie W, Lu X, Mantion G, Martin H, Richert L, Vuitton DA, Wen H. 2011. Time course of gene expression profiling in the liver of experimental mice infected with *Echinococcus multilocularis*. PLoS One 6:e14557. https://doi.org/10.1371/journal.pone.0014557.
- 269. Gelmedin V, Spiliotis M, Brehm K. 2010. Molecular characterisation of MEK1/2- and MKK3/6-like mitogen-activated protein kinase kinases (MAPKK) from the fox tapeworm *Echinococcus multilocularis*. Int J Parasitol 40:555–567. https://doi.org/10.1016/j.ijpara.2009.10.009.
- Gelmedin V, Caballero-Gamiz R, Brehm K. 2008. Characterization and inhibition of a p38-like mitogen-activated protein kinase (MAPK) from *Echinococcus multilocularis*: antiparasitic activities of p38 MAPK inhibitors. Biochem Pharmacol 76:1068–1081. https://doi.org/10.1016/j.bcp .2008.08.020.
- 271. Yang M, Li J, Wu J, Wang H, Guo B, Wu C, Shou X, Yang N, Zhang Z, McManus DP, Zhang F, Zhang W. 2017. Cloning and characterization of an *Echinococcus granulosus* ecdysteroid hormone nuclear receptor HR3-like gene. Parasite 24:36. https://doi.org/10.1051/parasite/2017037.
- 272. Koziol U, Koziol M, Preza M, Costabile A, Brehm K, Castillo E. 2016. De novo discovery of neuropeptides in the genomes of parasitic flatworms using a novel comparative approach. Int J Parasitol 46:709–721. https://doi.org/10.1016/j.ijpara.2016.05.007.
- 273. Hemer S, Konrad C, Spiliotis M, Koziol U, Schaack D, Forster S, Gelmedin V, Stadelmann B, Dandekar T, Hemphill A, Brehm K. 2014. Host insulin stimulates *Echinococcus multilocularis* insulin signalling pathways and larval development. BMC Biol 12:5. https://doi.org/10.1186/1741-7007-12-5
- 274. Zhang C, Wang L, Wang H, Pu H, Yang L, Li J, Wang J, Lu G, Lu X, Zhang W, Vuitton DA, Wen H, Lin R. 2014. Identification and characterization of functional Smad8 and Smad4 homologues from *Echinococcus granulosus*. Parasitol Res 113:3745–3757. https://doi.org/10.1007/s00436-014
- 275. Brehm K, Spiliotis M. 2008. The influence of host hormones and cytokines on *Echinococcus multilocularis* signalling and development. Parasite 15:286–290. https://doi.org/10.1051/parasite/2008153286.
- 276. Konrad C, Kroner A, Spiliotis M, Zavala-Góngora R, Brehm K. 2003. Identification and molecular characterisation of a gene encoding a member of the insulin receptor family in *Echinococcus multilocularis*. Int J Parasitol 33:301–312. https://doi.org/10.1016/S0020-7519(02)00265-5.
- 277. Brehm K. 2010. The role of evolutionarily conserved signalling systems in *Echinococcus multilocularis* development and host-parasite interaction. Med Microbiol Immunol 199:247–259. https://doi.org/10.1007/s00430-010-0154-1.
- 278. Spiliotis M, Konrad C, Gelmedin V, Tappe D, Bruckner S, Mosch HU, Brehm K. 2006. Characterisation of EmMPK1, an ERK-like MAP kinase from *Echinococcus multilocularis* which is activated in response to human epidermal growth factor. Int J Parasitol 36:1097–1112. https://doi.org/10.1016/j.ijpara.2006.05.008.
- Spiliotis M, Tappe D, Bruckner S, Mosch HU, Brehm K. 2005. Molecular cloning and characterization of Ras- and Raf-homologues from the fox-tapeworm *Echinococcus multilocularis*. Mol Biochem Parasitol 139: 225–237. https://doi.org/10.1016/j.molbiopara.2004.11.013.
- Zavala-Gongora R, Kroner A, Wittek B, Knaus P, Brehm K. 2003. Identification and characterisation of two distinct Smad proteins from the fox-tapeworm *Echinococcus multilocularis*. Int J Parasitol 33:1665–1677. https://doi.org/10.1016/S0020-7519(03)00208-X.
- 281. Spiliotis M, Kroner A, Brehm K. 2003. Identification, molecular characterization and expression of the gene encoding the epidermal growth factor receptor orthologue from the fox-tapeworm *Echinococcus multilocularis*. Gene 323:57–65. https://doi.org/10.1016/j.gene.2003.09.007.
- 282. Joekel DE, Lundstrom-Stadelmann B, Mullhaupt B, Hemphill A, Deplazes P. 2018. Evaluation of kinase-inhibitors nilotinib and everolimus against alveolar echinococcosis in vitro and in a mouse model. Exp Parasitol 188:65–72. https://doi.org/10.1016/j.exppara.2018.04.002.
- 283. Flo M, Margenat M, Pellizza L, Grana M, Duran R, Baez A, Salceda E, Soto E, Alvarez B, Fernandez C. 2017. Functional diversity of secreted cestode Kunitz proteins: inhibition of serine peptidases and blockade of

- cation channels. PLoS Pathog 13:e1006169. https://doi.org/10.1371/journal.ppat.1006169.
- Koziol U, Jarero F, Olson PD, Brehm K. 2016. Comparative analysis of Wnt expression identifies a highly conserved developmental transition in flatworms. BMC Biol 14:10. https://doi.org/10.1186/s12915-016-0233-x.
- 285. Ranasinghe SL, Fischer K, Zhang W, Gobert GN, McManus DP. 2015. Cloning and characterization of two potent Kunitz type protease inhibitors from *Echinococcus granulosus*. PLoS Negl Trop Dis 9:e0004268. https://doi.org/10.1371/journal.pntd.0004268.
- 286. Schubert A, Koziol U, Cailliau K, Vanderstraete M, Dissous C, Brehm K. 2014. Targeting *Echinococcus multilocularis* stem cells by inhibition of the Polo-like kinase EmPlk1. PLoS Negl Trop Dis 8:e2870. https://doi.org/10.1371/journal.pntd.0002870.
- 287. Hemer S, Brehm K. 2012. In vitro efficacy of the anticancer drug imatinib on *Echinococcus multilocularis* larvae. Int J Antimicrob Agents 40:458–462. https://doi.org/10.1016/j.ijantimicag.2012.07.007.
- 288. Vuitton DA. 2003. The ambiguous role of immunity in echinococcosis: protection of the host or of the parasite? Acta Trop 85:119–132. https://doi.org/10.1016/S0001-706X(02)00230-9.
- 289. Gottstein B, Soboslay P, Ortona E, Wang J, Siracusano A, Vuitton D. 2017. Immunology of alveolar and cystic echinococcosis (AE and CE). Adv Parasitol 96:1–54. https://doi.org/10.1016/bs.apar.2016.09.005.
- 290. Wang J, Gottstein B. 2016. Immunoregulation in larval *Echinococcus multilocularis* infection. Parasite Immunol 38:182–192. https://doi.org/10.1111/pim.12292.
- 291. Wang J, Lin R, Zhang W, Li L, Gottstein B, Blagosklonov O, Lu G, Zhang C, Lu X, Vuitton DA, Wen H. 2014. Transcriptional profiles of cytokine/ chemokine factors of immune cell-homing to the parasitic lesions: a comprehensive one-year course study in the liver of *E. multilocularis*-infected mice. PLoS One 9:e91638. https://doi.org/10.1371/journal.pone.0091638.
- 292. Gottstein B, Wittwer M, Schild M, Merli M, Leib SL, Muller N, Muller J, Jaggi R. 2010. Hepatic gene expression profile in mice perorally infected with *Echinococcus multilocularis* eggs. PLoS One 5:e9779. https://doi.org/10.1371/journal.pone.0009779.
- 293. Siracusano A, Delunardo F, Teggi A, Ortona E. 2012. Cystic echinococcosis: aspects of immune response, immunopathogenesis and immune evasion from the human host. Endocr Metab Immune Disord Drug Targets 12:16–23. https://doi.org/10.2174/1871530 12799279117.
- 294. Tuxun T, Apaer S, Ma HZ, Zhao JM, Lin RY, Aji T, Shao YM, Wen H. 2018. Plasma IL-23 and IL-5 as surrogate markers of lesion metabolic activity in patients with hepatic alveolar echinococcosis. Sci Rep 8:4417. https://doi.org/10.1038/s41598-018-20301-8.
- 295. Zhang C, Shao Y, Yang S, Bi X, Li L, Wang H, Yang N, Li Z, Sun C, Li L, Lu G, Aji T, Vuitton DA, Lin R, Wen H. 2017. T-cell tolerance and exhaustion in the clearance of *Echinococcus multilocularis*: role of inoculum size in a quantitative hepatic experimental model. Sci Rep 7:11153. https://doi.org/10.1038/s41598-017-11703-1.
- 296. Wang J, Müller S, Lin R, Siffert M, Vuitton DA, Wen H, Gottstein B. 2017. Depletion of FoxP3(+) Tregs improves control of larval *Echinococcus multilocularis* infection by promoting co-stimulation and Th1/17 immunity. Immun Inflamm Dis 5:435–447. https://doi.org/10.1002/iid3.181.
- 297. Labsi M, Soufli I, Khelifi L, Amir ZC, Touil-Boukoffa C. 2018. In vivo treatment with IL-17A attenuates hydatid cyst growth and liver fibrogenesis in an experimental model of echinococcosis. Acta Trop 181: 6–10. https://doi.org/10.1016/j.actatropica.2018.01.014.
- 298. Wang J, Vuitton DA, Muller N, Hemphill A, Spiliotis M, Blagosklonov O, Grandgirard D, Leib SL, Shalev I, Levy G, Lu X, Lin R, Wen H, Gottstein B. 2015. Deletion of fibrinogen-like protein 2 (FGL-2), a novel CD4+ CD25+ Treg effector molecule, leads to improved control of *Echinococcus multilocularis* infection in mice. PLoS Negl Trop Dis 9:e0003755. https://doi.org/10.1371/journal.pntd.0003755.
- 299. Wang J, Cardoso R, Marreros N, Muller N, Lundstrom-Stadelmann B, Siffert M, Vuitton DA, Boue F, Lin R, Wen H, Gottstein B. 2018. Foxp3+ Tregs as a potential target for immunotherapy against primary infection with *Echinococcus multilocularis* eggs. Infect Immun 86:e00542-18. https://doi.org/10.1128/IAI.00542-18.
- 300. Liu H, Bakthavatsalam R, Meng Z, Li Z, Li W, Perkins JD, Reyes J. 2013. PD-L1 signal on liver dendritic cells is critical for Foxp3(+) CD4(+)CD25(+) Treg and liver tolerance induction in mice. Transplant Proc 45:1853–1855. https://doi.org/10.1016/j.transproceed.2013.03
- 301. Li Y, Xiao Y, Su M, Zhang R, Ding J, Hao X, Ma Y. 2016. Role of soluble

programmed death-1 (sPD-1) and sPD-ligand 1 in patients with cystic echinococcosis. Exp Ther Med 11:251–256. https://doi.org/10.3892/etm .2015.2876.

- 302. Zhang F, Pang N, Zhu Y, Zhou D, Zhao H, Hu J, Ma X, Li J, Wen H, Samten B, Fan H, Ding J. 2015. CCR7(lo)PD-1(hi) CXCR5(+) CD4(+) T cells are positively correlated with levels of IL-21 in active and transitional cystic echinococcosis patients. BMC Infect Dis 15:457. https://doi.org/10.1186/s12879-015-1156-9.
- 303. La X, Zhang F, Li Y, Li J, Guo Y, Zhao H, Pang N, Ma X, Wen H, Fan H, Ding J. 2015. Upregulation of PD-1 on CD4(+)CD25(+)T cells is associated with immunosuppression in liver of mice infected with *Echinococcus multilocularis*. Int Immunopharmacol 26:357–366. https://doi.org/10.1016/j.intimp.2015.04.013.
- 304. Wang J, Jebbawi F, Bellanger AP, Beldi G, Millon L, Gottstein B. 2018. Immunotherapy of alveolar echinococcosis via PD-1/PD-L1 immune checkpoint blockade in mice. Parasite Immunol 40:e12596. https://doi.org/10.1111/pim.12596.
- 305. Swaika A, Hammond WA, Joseph RW. 2015. Current state of anti-PD-L1 and anti-PD-1 agents in cancer therapy. Mol Immunol 67:4–17. https://doi.org/10.1016/j.molimm.2015.02.009.
- Li T, Zhao JM, Zhang Y, Pai ZL, Zhang W, Tuxun TH, Bai L, Wu J, Wen H.
 Suppression of acute rejective response following orthotopic liver transplantation in experimental rats infected with *Echinococcus multilocularis*. Chin Med J (Engl) 124:2818–2823.
- Wang H, Li J, Pu H, Hasan B, Ma J, Jones MK, Zheng K, Zhang X, Ma H, McManus DP, Lin R, Wen H, Zhang W. 2014. *Echinococcus granulosus* infection reduces airway inflammation of mice likely through enhancing IL-10 and down-regulation of IL-5 and IL-17A. Parasit Vectors 7:522. https://doi.org/10.1186/s13071-014-0522-6.
- Apaer S, Tuxun T, Ma HZ, Zhang H, Aierken A, Aini A, Li YP, Lin RY, Wen H. 2016. Parasitic infection as a potential therapeutic tool against rheumatoid arthritis. Exp Ther Med 12:2359–2366. https://doi.org/10.3892/etm.2016.3660.
- 309. Khelifi L, Soufli I, Labsi M, Touil-Boukoffa C. 2017. Immune-protective effect of echinococcosis on colitis experimental model is dependent of down regulation of TNF-alpha and NO production. Acta Trop 166:7–15. https://doi.org/10.1016/j.actatropica.2016.10.020.
- Wang J, Goepfert C, Mueller N, Piersigilli A, Lin R, Wen H, Vuitton DA, Vuitton L, Mueller C, Gottstein B. 2018. Larval *Echinococcus multilocularis* infection reduces dextran sulphate sodium-induced colitis in mice by attenuating T helper type 1/type 17-mediated immune reactions. Immunology 154:76–88. https://doi.org/10.1111/imm.12860.
- 311. Soufli I, Toumi R, Rafa H, Amri M, Labsi M, Khelifi L, Nicoletti F, Touil-Boukoffa C. 2015. Crude extract of hydatid laminated layer from *Echinococcus granulosus* cyst attenuates mucosal intestinal damage and inflammatory responses in dextran sulfate sodium induced colitis in mice. J Inflamm (Lond) 12:19. https://doi.org/10.1186/s12950-015-0063-6
- 312. Gottstein B, Hemphill A. 2008. *Echinococcus multilocularis*: the parasite-host interplay. Exp Parasitol 119:447–452. https://doi.org/10.1016/j.exppara.2008.03.002.

- 313. Siracusano A, Delunardo F, Teggi A, Ortona E. 2012. Host-parasite relationship in cystic echinococcosis: an evolving story. Clin Dev Immunol 2012:639362. https://doi.org/10.1155/2012/639362.
- 314. Lightowlers MW, Heath DD. 2004. Immunity and vaccine control of *Echinococcus granulosus* infection in animal intermediate hosts. Parassitologia 46:27–31.
- 315. Larrieu E, Herrero E, Mujica G, Labanchi JL, Araya D, Grizmado C, Calabro A, Talmon G, Ruesta G, Perez A, Gatti A, Santillan G, Cabrera M, Arezzo M, Seleiman M, Cavagion L, Cachau MG, Alvarez Rojas CA, Gino L, Gauci CG, Heath DD, Lamberti R, Lightowlers MW. 2013. Pilot field trial of the EG95 vaccine against ovine cystic echinococcosis in Rio Negro, Argentina: early impact and preliminary data. Acta Trop 127: 143–151. https://doi.org/10.1016/j.actatropica.2013.04.009.
- 316. Larrieu E, Mujica G, Gauci CG, Vizcaychipi K, Seleiman M, Herrero E, Labanchi JL, Araya D, Sepulveda L, Grizmado C, Calabro A, Talmon G, Poggio TV, Crowley P, Cespedes G, Santillan G, Garcia Cachau M, Lamberti R, Gino L, Donadeu M, Lightowlers MW. 2015. Pilot field trial of the EG95 vaccine against ovine cystic echinococcosis in Rio Negro, Argentina: second study of impact. PLoS Negl Trop Dis 9:e0004134. https://doi.org/10.1371/journal.pntd.0004134.
- Heath DD, Robinson C, Shakes T, Huang Y, Gulnur T, Shi B, Zhang Z, Anderson GA, Lightowlers MW. 2012. Vaccination of bovines against Echinococcus granulosus (cystic echinococcosis). Vaccine 30:3076–3081. https://doi.org/10.1016/j.vaccine.2012.02.073.
- 318. Chow C, Gauci CG, Cowman AF, Lightowlers MW. 2004. *Echinococcus granulosus*: oncosphere-specific transcription of genes encoding a host-protective antigen. Exp Parasitol 106:183–186. https://doi.org/10.1016/j.exppara.2004.03.009.
- 319. Zhang ZZ, Guo G, Li J, Shi BX, Zhao L, Guo BP, Zhang X, Wang JW, Zheng XT, Qi WJ, He L, Zhang WB. 2018. Dog vaccination with EgM proteins against *Echinococcus granulosus*. Infect Dis Poverty 7:61. https://doi.org/10.1186/s40249-018-0425-4.
- 320. Zhang W, Zhang Z, Shi B, Li J, You H, Tulson G, Dang X, Song Y, Yimiti T, Wang J, Jones MK, McManus DP. 2006. Vaccination of dogs against *Echinococcus granulosus*, the cause of cystic hydatid disease in humans. J Infect Dis 194:966–974. https://doi.org/10.1086/506622.
- 321. Cuesta-Astroz Y, Oliveira FS, Nahum LA, Oliveira G. 2017. Helminth secretomes reflect different lifestyles and parasitized hosts. Int J Parasitol 47:529–544. https://doi.org/10.1016/j.ijpara.2017.01.007.
- 322. Behrendt P, Arnold P, Brueck M, Rickert U, Lucius R, Hartmann S, Klotz C, Lucius R. 2016. A helminth protease inhibitor modulates the lipopolysaccharide-induced proinflammatory phenotype of microglia in vitro. Neuroimmunomodulation 23:109–121. https://doi.org/10.1159/000444756.
- 323. Morais SB, Figueiredo BC, Assis NRG, Alvarenga DM, de Magalhaes MTQ, Ferreira RS, Vieira AT, Menezes GB, Oliveira SC. 2018. *Schistosoma mansoni* SmKl-1 serine protease inhibitor binds to elastase and impairs neutrophil function and inflammation. PLoS Pathog 14:e1006870. https://doi.org/10.1371/journal.ppat.1006870.

Hao Wen, M.D., Ph.D., FACS, is an honorary fellow of the French Academy of Surgery, Head and Senior Principal Research Fellow of the State Key Laboratory of Pathogenesis, Prevention and Treatment of High Incidence Diseases in Central Asia, and Professor of Surgery and Senior Consultant Hepatobiliary & Pancreatic and Transplant Surgeon at The First Affiliated Hospital, Xinjiang Medical University, Urumqi, China. He completed his Ph.D. at the Faculty of Life Science, Salford



University, United Kingdom, and completed postdoctoral study at the Institute of Medicine and Pharmacy, Franche-Comte University, France, from 1994 to 1995. He has been Vice Chairman of the Chinese Medical Doctor Association Organ Transplant Branch since 2014, Vice Chairman of the World Association of Echinococcosis since 2009, and Director of the WHO Collaborating Centre for Echinococcosis Prevention and Management since 2016. Professor Wen has published 150 refereed papers/ articles in international journals in his research career. He has spent more than 30 years working on the prevention, diagnosis, and treatment of echinococcosis in China and has pioneered *ex vivo* liver resection and autotransplantation in end-stage alveolar echinococcosis globally. Most of his publications focus on clinical management, diagnosis, epidemiology, molecular biology, and immunology of echinococcosis.

Lucine Vuitton, M.D., Ph.D., is a Hospital Physician in the Gastroenterology and Endoscopy Department at the University Hospital of Besançon, France, and a postdoctoral fellow at University Bourgogne Franche-Comté (UBFC). She trained in gastroenterology in Besançon and Marseille, France, and had a 1-year research fellow position at the Inflammatory Bowel Disease (IBD) Unit of Nancy University Hospital in 2017. Her daily clinical activity is related to perendoscopic



interventions and care of IBD patients. Her research deals with clinical endoscopy, IBD care management, and the relationship between IBD and infection, namely, human papillomaviruses and *E. multilocularis*. Dr. Vuitton has published 58 scientific articles, which have been cited more than 600 times. She is a member of the French Society of Digestive Endoscopy and of the French Endosonography Group. As a member of the WHO Collaborating Centre for Prevention and Treatment of Human Echinococcosis, she has coordinated the European survey on the endoscopic management of echinococcosis by endoscopic retrograde cholangiography. She is a member of the board of the GETAID (French IBD Therapy Study Working Group) and of the GETAID's educational committee.

Tuerhongjiang Tuxun, M.D., Ph.D., is an Associate Professor of Surgery at The First Affiliated Hospital of Xinjiang Medical University, Urumqi, China. He is a research fellow of the State Key Laboratory of Pathogenesis, Prevention and Treatment of High Incidence Diseases in Central Asia, Xinjiang Medical University, Urumqi, China, and the Clinical Medical Research Institute. He received his M.Sc. from Xinjiang University in 2010 and completed his doctoral study at Xinjiang



Medical University in 2016. He has been a member of the Youth Committee of the Chinese Medical Doctor Association Hydatid Surgery Branch since 2014. He was the winner of the Shulan Talent Foundation award for "outstanding young transplant surgeon" in 2017. He has spent more than 10 years working on the treatment and control of echinococcosis. Most of his publications focus on the clinical management, diagnosis, and immunology of echinococcosis. Associate Professor Tuxun has published 100 refereed papers/articles, including 30 in international journals, in his research career.

Jun Li, B.Sc., Ph.D., is a Professor at the Xinjiang Medical University and a senior research fellow of the State Key Laboratory of Pathogenesis, Prevention and Treatment of High Incidence Diseases in Central Asia, Xinjiang Medical University, Urumqi, China. She received her B.Sc. from Xinjiang Medical University, and in 2004 she obtained her Ph.D. at the University of Queensland, Australia, working on the development of diagnostic tools for detecting cystic echinococcosis. She



then spent 3 years working at the commercial company PanBio developing diagnostic kits for infectious diseases. From 2008 to 2013, she worked on the molecular biology of *Echinococcus* as a senior research officer in the Molecular Parasitology Laboratory, Infectious Diseases Division, QIMR Berghofer Medical Research Institute, Brisbane, Queensland, Australia. She has published over 30 papers/articles in international journals in her research career.

Dominique A. Vuitton, M.D., Ph.D., is a Professor Emeritus of Clinical Immunology at University Bourgogne Franche-Comté, France, and a scientific consultant at the State Key Laboratory of Pathogenesis, Prevention and Treatment of High Incidence Diseases in Central Asia, Urumqi, China. A major part of her research work has been dedicated to the interactions between *Echinococcus* spp. and their hosts, to the epidemiology and of risk factors for cystic and alveolar echinococcosis in



the areas of endemicity in western China, and to the care management of patients with echinococcosis; she was involved in the first clinical trials of mebendazole and albendazole, in the first attempts at liver transplantation for AE, and in the definition of international classifications and recommendations for the care management of echinococcosis patients. Professor Vuitton has published 355 scientific articles, which have been cited 8,680 times. She was the coordinator of the WHO Informal Working Group on Echinococcosis (WHO, Geneva) and the head of the WHO Collaborating Centre for Prevention and Treatment of Human Echinococcosis (Besançon, France). She received the Chinese "Friendship Prize" in 2010 and the French "Légion d'Honneur" in 2011 for her professional achievements and was elected Fellow of the French National Academy of Medicine in 2017.

Wenbao Zhang, B.Sc., Ph.D., is a senior research fellow of the State Key Laboratory of Pathogenesis, Prevention and Treatment of High Incidence Diseases in Central Asia, Xinjiang Medical University, Urumqi, China, and of the Clinical Medical Research Institute, The First Affiliated Hospital of Xinjiang Medical University, Urumqi, China. He received his B.Sc. from Xinjiang University in 1983. From 1999 to 2003, he was an Australian International Postgraduate Research Scholar,



and he completed his Ph.D. at the University of Queensland. He has spent more than 30 years working on the treatment and control of zoonoses, including echinococcosis, tuberculosis, brucellosis, and schistosomiasis. Most of his publications focus on the molecular biology, immunology, diagnosis, and epidemiology of these diseases. Professor Zhang has published 150 refereed papers/articles, including 58 in international journals, in his research career.

Donald P. McManus, B.Sc., Ph.D., D.Sc. (Wales), is an NHMRC Senior Principal Research Fellow, Senior Principal Research Fellow, and Senior Scientist at Berghofer Medical Research Institute, Professor of Tropical Health, University of Queensland, and Professor, Australian National University. He researches the molecular biology, immunology, diagnosis, and epidemiology of parasitic worms. Professor McManus has published over 600 articles, which have been cited over 21,000 times. He



was made Honorary International Fellow of the American Society of Tropical Medicine and Hygiene in 2010 and received honorary membership in the American Society of Parasitologists in 2012. He was elected Fellow of the Royal Society of Biology (United Kingdom) in 2013 and was the recipient of the 2014 Ralph Doherty QIMR Berghofer Prize for Outstanding Achievement and Leadership in Medical Research. In 2015 he was elected Fellow of the Australian Academy of Health and Medical Sciences. He is the winner of the 2018 Sornchai Looareesuwan Medal "for outstanding achievements in experimental and clinical topical medicine research."